

## SUPPORTING INFORMATION

### Truncation of the constant domain drives amyloid formation by immunoglobulin light chains

Francesca Lavatelli<sup>1,2#</sup>, Antonino Natalello<sup>3#</sup>, Loredana Marchese<sup>4</sup>, Diletta Ami<sup>3</sup>, Alessandra Corazza<sup>5,6</sup>, Sara Raimondi<sup>1</sup>, Maria Chiara Mimmi<sup>7</sup>, Silvia Malinverni<sup>1</sup>, P. Patrizia Mangione<sup>1,2</sup>, Manel Terrones Palmer<sup>3</sup>, Alessio Lampis<sup>1</sup>, Monica Concardi<sup>7</sup>, Guglielmo Verona<sup>1,8</sup>, Diana Canetti<sup>8</sup>, Eloisa Arbustini<sup>7</sup>, Vittorio Bellotti<sup>2</sup>, Sofia Giorgetti<sup>1,2#</sup>.

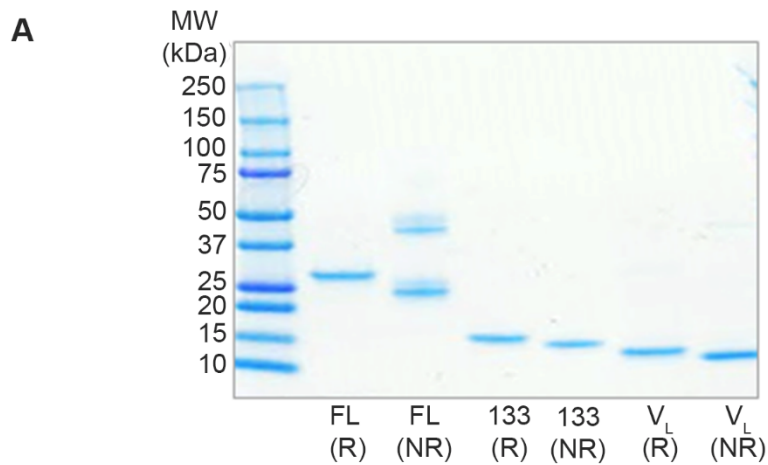
From the <sup>1</sup>Department of Molecular Medicine, Institute of Biochemistry, University of Pavia, 27100 Pavia, Italy; <sup>2</sup>Research Area, Fondazione IRCCS Policlinico San Matteo, 27100 Pavia, Italy; <sup>3</sup>Department of Biotechnology and Biosciences, University of Milano-Bicocca, 20126 Milan, Italy; <sup>4</sup>Pathology Unit, Fondazione IRCCS Policlinico San Matteo, Pavia, Italy; <sup>5</sup>Department of Medicine (DAME), University of Udine, 33100 Udine, Italy; <sup>6</sup>Istituto Nazionale Biostrutture e Biosistemi, 00136 Roma, Italy; <sup>7</sup>Transplant Research Area and Centre for Inherited Cardiovascular Diseases, Fondazione IRCCS Policlinico San Matteo, 27100 Pavia, Italy; <sup>8</sup>Centre for Amyloidosis, Division of Medicine, University College London, London NW3 2PF, UK.

*Running title: Amyloidogenicity of immunoglobulin light chain fragments*

<sup>#</sup>To whom correspondence should be addressed: Department of Molecular Medicine, Institute of Biochemistry, Via Taramelli 3b, 27100 Pavia, Italy. Tel: +390382987783; E-mail: [francesca.lavatelli@unipv.it](mailto:francesca.lavatelli@unipv.it); [s.giorgetti@unipv.it](mailto:s.giorgetti@unipv.it)  
Department of Biotechnology and Biosciences, Piazza della Scienza 2, 20126 Milan, Italy. Tel: +39026448 3459; E-mail: [antonino.natalello@unimib.it](mailto:antonino.natalello@unimib.it)

**MATERIAL INCLUDED:**

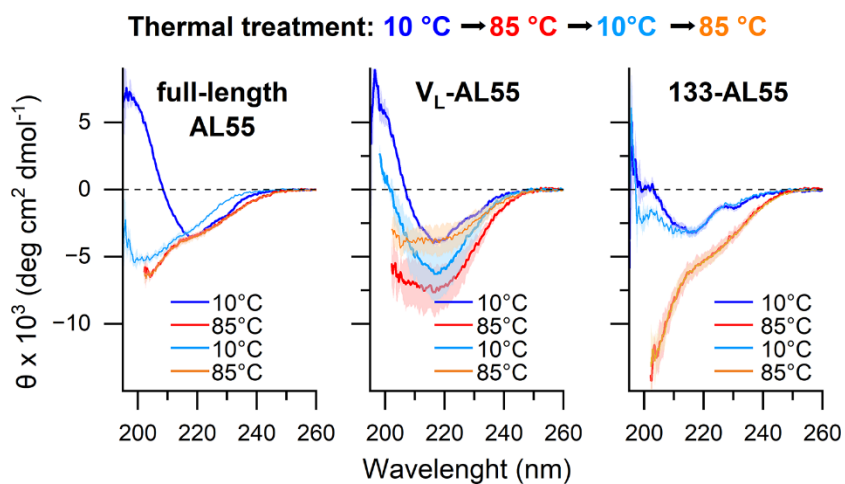
**FIGURES S1-S6**



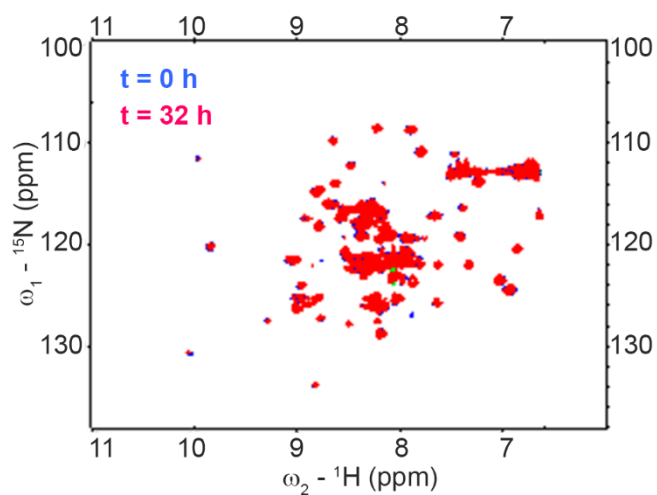
**B**

	full-length AL55	133-AL55	VL-AL55
<b>Amino acid</b>	217 (+ N-term Met)	133 (+ N-term Met)	111 (+ N-term Met)
<b>Predicted MW (Da)</b>	23306.63	14309.59	12290.35

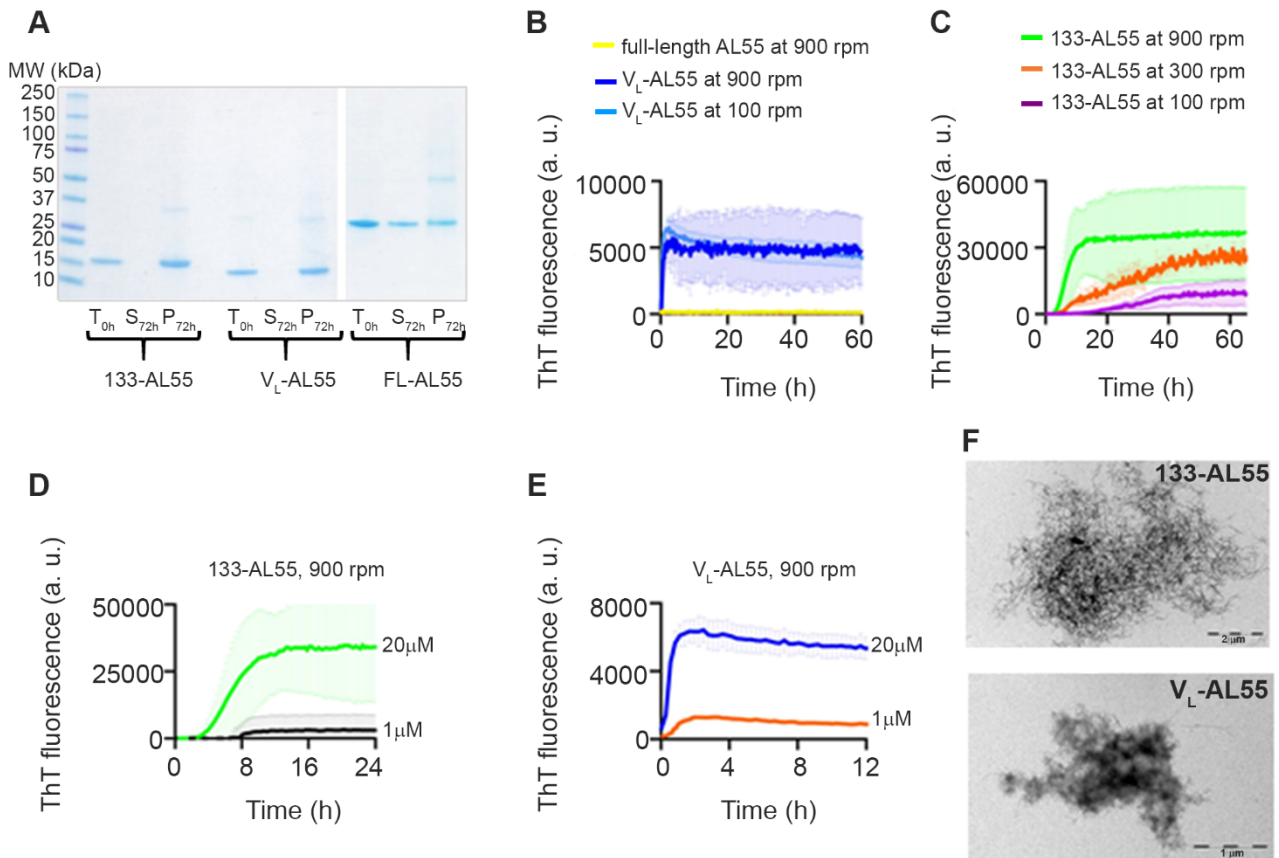
**Figure S1. Recombinant full-length AL55, VL-AL55 and 133-AL55.** (A) SDS 4-20 % PAGE analysis under reducing (R) and non-reducing (NR) conditions. The two bands visible for the full-length under non-reducing conditions correspond to the monomer (~ 25 kDa) and to the disulfide-linked dimer (~ 50 kDa). (B) main biochemical parameters of the three proteins calculated using the ExPasy ProtParam tool. MW, molecular weight.



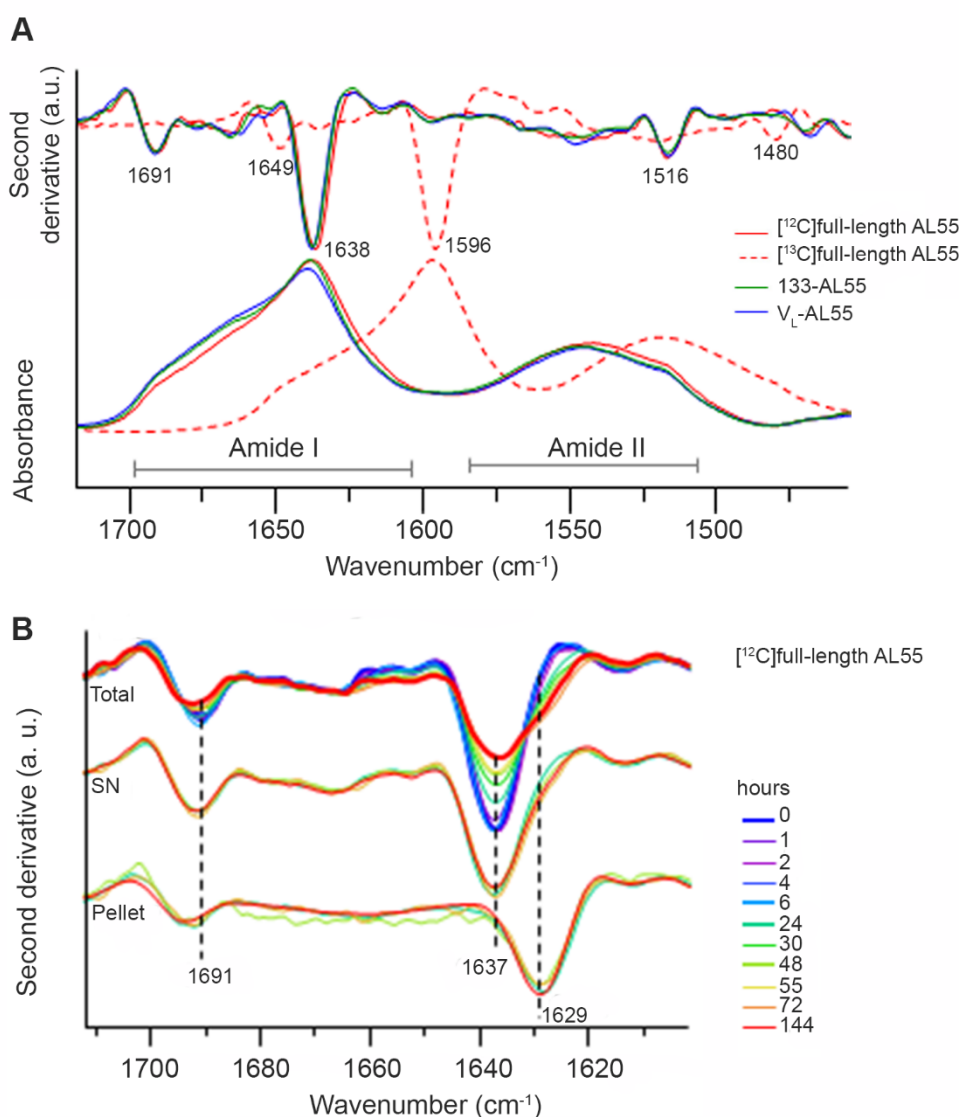
**Figure S2. Assessment of the thermal unfolding reversibility of full-length and truncated AL55 species.** Overlay of CD far UV spectra of each protein samples recorded during the first thermal treatment at 10°C (blue) and 85°C (red) respectively. Spectra recorded after cooling (light blue) and after a second thermal treatment (orange) are shown.



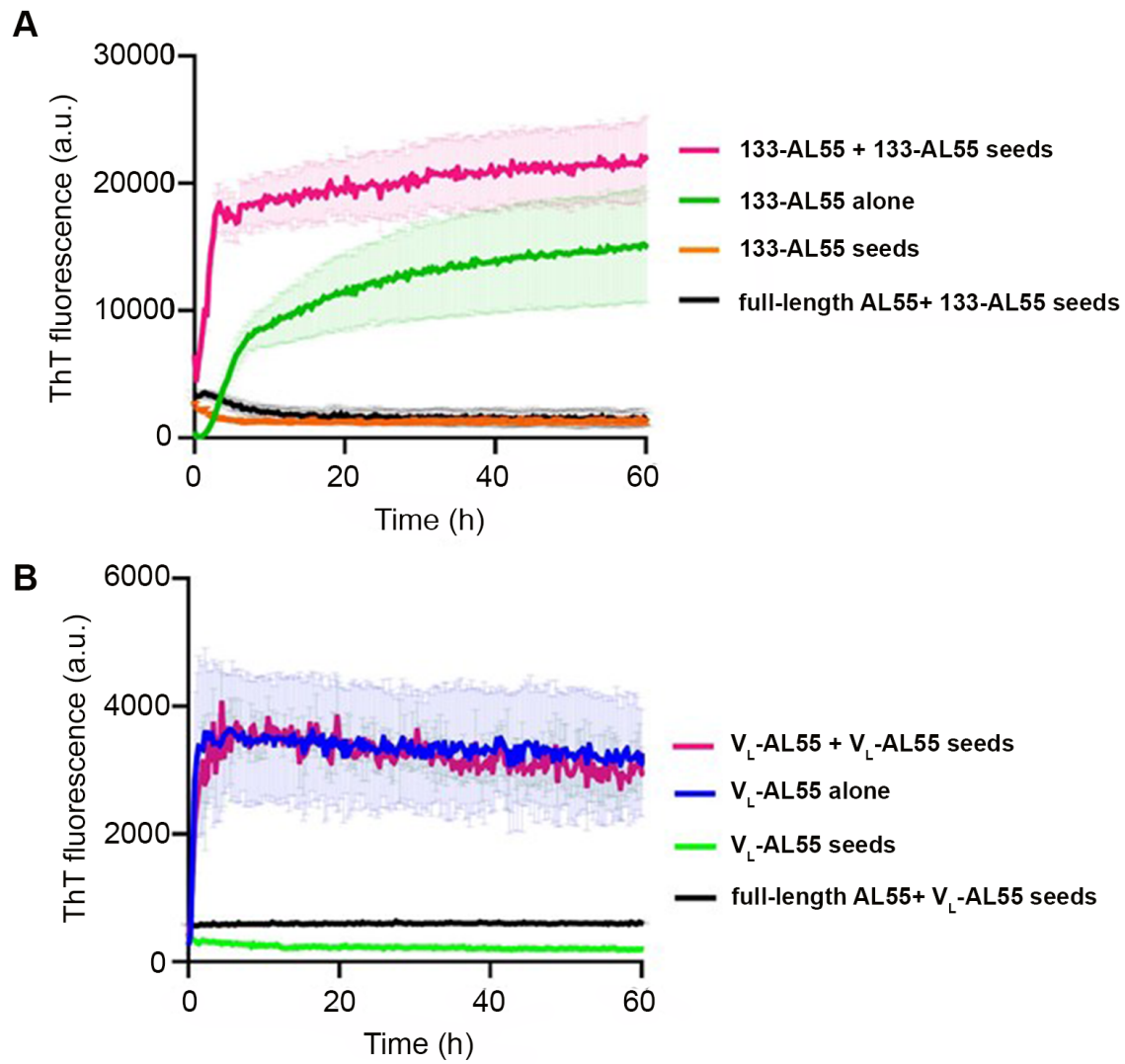
**Figure S3. Conformational stability of 133-AL55 at 15 °C monitored by solution NMR.** The “fingerprint” spectra, i.e. 2D [ $^1\text{H}$ - $^{15}\text{N}$ ] HSQC spectra, of uniformly [ $^{13}\text{C}$ ,  $^{15}\text{N}$ ] labeled 133-AL55 at 0.468 mM in PBS pH 7.4 containing 5%  $\text{D}_2\text{O}$  were recorded immediately after dissolution (blue trace) and after 32 hours at 15 °C (red trace).



**Figure S4. Fibrillogenesis of full-length AL55,  $V_L$ -AL55 and 133-AL55.** (A) SDS 4-20 %-PAGE analysis under reducing conditions of pellets (P) separated by centrifugation from supernatants (S) after incubation of proteins for 72 h under agitation at 100 rpm. The starting material is indicated as  $T_{0h}$ . (B), fibrillogenesis of  $V_L$ -AL55 and full-length AL55 (20  $\mu$ M) at 100 rpm and 900 rpm, monitored by ThT assay as described in Material and Methods. Shown  $V_L$ -AL55 at 900 rpm (blue),  $V_L$ -AL55 at 100 rpm (light blue); full-length AL55 at 900 rpm (yellow); full-length AL55 at 100 rpm not shown as it overlaps with the one at 900 rpm. (C), fibrillogenesis of 133-AL55 (20  $\mu$ M) at 100 rpm (purple), 300 rpm (orange) and 900 rpm (green), monitored by ThT assay. (D), fibrillogenesis of 133-AL55 at 900 rpm at 20  $\mu$ M (green) and 1  $\mu$ M (black) respectively. (E), fibrillogenesis of  $V_L$ -AL55 at 900 rpm, 20  $\mu$ M (blue) and 1  $\mu$ M (orange) respectively. All data shown as mean and SD of at least three independent experimental replicates. (F), TEM images of the aggregates by AL-133 and  $V_L$ -AL55, at low magnification. Scale bars are indicated.



**Figure S5. FTIR spectroscopy.** (A), FTIR absorption spectra and second derivatives of the three native unlabeled [<sup>12</sup>C] species and the labeled [<sup>13</sup>C]full-length AL55, are shown with corresponding main peaks positions. (B), time course of unlabeled full-length AL55. Second derivatives of the absorption spectra collected at different incubation times, as indicated. Aliquots of each sample (20  $\mu$ l) at 24, 48, 72 and 144 hours of incubation were centrifuged and corresponding FTIR spectra of supernatants (SN, middle spectra) and pellets (bottom spectra) were acquired. Representative spectra with the main peak positions are shown.



**Figure S6. Seeding and cross-seeding experiments**, performed by incubating soluble proteins with preformed aggregates by either 133-AL55 (A) or  $V_L$ -AL55 (B). Data are shown as mean and standard deviation of three independent experiments.