

## Finding the seed of recurrence: Hepatocellular carcinoma circulating tumor cells and their potential to drive the surgical treatment

Francesca Carissimi, Matteo Nazzareno Barbaglia, Livia Salmi, Cristina Ciulli, Linda Roccamatise, Giuseppe Cordaro, Venkata Ramana Mallela, Rosalba Minisini, Biagio Eugenio Leone, Matteo Donadon, Guido Torzilli, Mario Pirisi, Fabrizio Romano, Simone Famularo

**ORCID number:** Francesca Carissimi 0000-0002-1647-8397; Matteo Nazzareno Barbaglia 0000-0001-9271-7508; Livia Salmi 0000-0002-8638-0897; Cristina Ciulli 0000-0003-0911-5114; Linda Roccamatise 0000-0002-0306-1277; Giuseppe Cordaro 0000-0002-3414-4948; Venkata Ramana Mallela 0000-0001-9664-481X; Rosalba Minisini 0000-0002-7548-9731; Biagio Eugenio Leone 0000-0002-6978-2405; Matteo Donadon 0000-0003-0296-7648; Guido Torzilli 0000-0001-5798-5021; Mario Pirisi 0000-0001-9740-0155; Fabrizio Romano 0000-0001-5341-0706; Simone Famularo 0000-0002-9721-6304.

**Author contributions:** Carissimi F, Famularo S, Barbaglia MN, and Salmi L wrote the manuscript; Carissimi F, Famularo S, Barbaglia MN, Salmi L, Roccamatise L, Cordaro G, Ciulli C, and Mallela VR collected and analyzed data; Carissimi F, Famularo S, Barbaglia MN, Salmi L, Roccamatise L, Cordaro G, Ciulli C, Mallela VR, and Minisini R reviewed the literature; Salmi L, and Barbaglia MN realized figures; Famularo S, Carissimi F, Minisini R, Leone BE, Donadon M, Torzilli G, Pirisi M, and Romano F designed the study.

**Conflict-of-interest statement:** The authors declare that they have no

**Francesca Carissimi, Cristina Ciulli, Linda Roccamatise, Giuseppe Cordaro, Biagio Eugenio Leone, Fabrizio Romano, Simone Famularo,** School of Medicine and Surgery, University of Milano-Bicocca, Monza 20900, Italy

**Matteo Nazzareno Barbaglia, Livia Salmi, Venkata Ramana Mallela, Rosalba Minisini, Mario Pirisi,** Department of Translational Medicine, Università del Piemonte Orientale, Novara 28100, Italy

**Biagio Eugenio Leone,** Unit of Pathology, San Gerardo Hospital, Monza 20900, Italy

**Matteo Donadon, Guido Torzilli, Simone Famularo,** Department of Hepatobiliary and General Surgery, IRCCS Humanitas Clinical and Research Hospital-Department of Biomedical Science, Humanitas University, Pieve Emanuele 20090, Italy

**Fabrizio Romano,** Department of Medicine and Surgery, University of Milan-Bicocca, Monza 20900, Italy

**Corresponding author:** Fabrizio Romano, MD, Professor, Surgeon, Surgical Oncologist, Department of Medicine and Surgery, University of Milan-Bicocca, via pergolesi 33, Monza 20900, Italy. [fabrizio.romano@unimib.it](mailto:fabrizio.romano@unimib.it)

### Abstract

The treatment for hepatocellular carcinoma (HCC) relies on liver resection, which is, however, burdened by a high rate of recurrence after surgery, up to 60% at 5 years. No pre-operative tools are currently available to assess the recurrence risk tailored to every single patient. Recently liquid biopsy has shown interesting results in diagnosis, prognosis and treatment allocation strategies in other types of cancers, since its ability to identify circulating tumor cells (CTCs) derived from the primary tumor. Those cells were advocated to be responsible for the majority of cases of recurrence and cancer-related deaths for HCC. In fact, after being modified by the epithelial-mesenchymal transition, CTCs circulate as “seeds” in peripheral blood, then reach the target organ as dormant cells which could be subsequently “awakened” and activated, and then initiate metastasis. Their presence may justify the disagreement registered in terms of efficacy of anatomic vs non-anatomic resections, particularly in the case of microvascular invasion, which has been recently pointed as a histological sign of the spread of those cells. Thus, their presence, also in the early stages, may justify the recurrence event also

conflict of interest.

**Open-Access:** This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

**Manuscript source:** Invited manuscript

**Specialty type:** Surgery

**Country/Territory of origin:** Italy

**Peer-review report's scientific quality classification**

Grade A (Excellent): 0

Grade B (Very good): 0

Grade C (Good): C

Grade D (Fair): 0

Grade E (Poor): 0

**Received:** March 21, 2021

**Peer-review started:** March 21, 2021

**First decision:** April 29, 2021

**Revised:** May 25, 2021

**Accepted:** August 10, 2021

**Article in press:** August 10, 2021

**Published online:** September 27, 2021

**P-Reviewer:** Fernandes SA

**S-Editor:** Ma YJ

**L-Editor:** A

**P-Editor:** Wu RR



in the contest of liver transplant. Understanding the mechanism behind the tumor progression may allow improving the treatment selection according to the biological patient-based characteristics. Moreover, it may drive the development of novel biological tailored tests which could address a specific patient to neoadjuvant or adjuvant strategies, and in perspective, it could also become a new method to allocate organs for transplantation, according to the risk of relapse after liver transplant. The present paper will describe the most recent evidence on the role of CTCs in determining the relapse of HCC, highlighting their potential clinical implication as novel tumor behavior biomarkers able to influence the surgical choice.

**Key Words:** Hepatocellular carcinoma; Liquid biopsy; Circulating tumor cells; Liver surgery; Microvascular invasion; Hepatocellular carcinoma recurrence

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core Tip:** In recent years many studies have shown that surgery is the first choice treatment for hepatocellular carcinoma (HCC) patients; although undergoing surgery at early stage many patients develop relapse during follow-up. Currently, there are no tools sensitive enough to identify recurrence risk factors. Recent studies have identified liquid biopsy as a valid method for diagnosis, prognosis and treatment allocation in HCC patients thanks to its effectiveness in identification of circulating tumor cells (CTCs), advocated to be responsible for relapse. In this manuscript we describe the main markers expressed by CTCs and how their presence in blood sample may be implied in the progression mechanism, and how they can modify the surgical strategy.

**Citation:** Carissimi F, Barbaglia MN, Salmi L, Ciulli C, Roccamatisi L, Cordaro G, Mallela VR, Minisini R, Leone BE, Donadon M, Torzilli G, Pirisi M, Romano F, Famularo S. Finding the seed of recurrence: Hepatocellular carcinoma circulating tumor cells and their potential to drive the surgical treatment. *World J Gastrointest Surg* 2021; 13(9): 967-978

**URL:** <https://www.wjgnet.com/1948-9366/full/v13/i9/967.htm>

**DOI:** <https://dx.doi.org/10.4240/wjgs.v13.i9.967>

## INTRODUCTION

Hepatocellular carcinoma (HCC) is the sixth most common cancer by diagnosis and the fourth highest cause of cancer-related death worldwide[1]. Surgical resection or liver transplantation are curative options, but unfortunately less than 40% of patients are eligible due to advanced stage at diagnosis[2].

Liver resection remains the mainstay among the curative treatments, but it is affected by a recurrence rate of up to 60% at 5 years, even for early stage tumours[3]. The route of recurrence is still a matter of debate. The relapse of HCC may be driven by precancerous status of the remaining diseased liver: namely "multicentric de-novo occurrence", these tumours are always primitive[4].

However, the majority of recurrence is attributed to intra-hepatic metastasis, driven by the acquisition of the cancer hallmark of invasiveness[5]. Clinically speaking, no tools have been developed to recognise the two different patterns before treatment, although basic and pre-clinical studies have identified several genetic signatures[6,7]. While multicentric occurrence cannot be controlled by liver resection alone, intra-hepatic metastasis could be avoided by an appropriate resection: almost thirty years ago, Makuuchi *et al*[8] stasisation could be avoided by an appropriate resection: almost thirty years ago, Makuuchi *et al*[8] reported a high rate of recurrence after surgery when microvascular invasion and satellitosis were present in the histological specimen. This evidence and that from other experiments[8-10], ed portal vein dissemination to be considered to be the main route of intra-hepatic metastasis, developing the notion of anatomical resection that relies on the complete removal of the whole segmental portal-flow area of the liver segment hosting the tumour. This technical approach was expected to allow better control of the area with the highest risk of tumour spread, reducing recurrence rates. However, several authors

compared recurrence rates among anatomical and non-anatomical resections, without a clear conclusion[11]. In recent years, the challenge of recurrence even after liver transplantation, a better knowledge of the tumour blood flow area[12], and the modern knowledge derived from molecular studies, have forced us to rethink the portal theory of HCC recurrence. In fact, intra-hepatic metastasis seems to be caused by local dissemination among the tumour blood flow, or by the systemic dissemination of tumour cells. These circulating tumour cells (CTCs) have also been identified in other types of tumours[13], and may have the ability to rehome themselves in the liver[14], and consequently could explain cases of relapse even after organ transplantation. In the present paper, we aimed to critically review the literature regarding HCC recurrence and CTC identification, and their role in surgery. We interpret our previous data in light of results from other studies, aiming to suggest a possible general picture to inform future research in the field.

## RECONSIDERING THE ROUTE OF RECURRENCE: EVIDENCE-DRIVEN HYPOTHESIS

Nakashima *et al*[15] have proposed that the portal vein (PV) may act as the efferent vessel during the oncoprogression of HCC, particularly in the setting of cirrhotic patients, where the hepatic veins are compromised. In this theory, the hepatic artery is the feeding vessel, and the PV, as an efferent vessel, penetrates the tumour capsule, and becomes the path of minor resistance for tumour infiltration or expansion[9] and the drainage pathway of the neoplasm. This mechanism was described to explain the high rate of tumour thrombi observed, and the presence of satellitosis near the primitive tumour. Those considerations led to the proposal of the anatomic resection (AR) to completely remove the parenchymal area fed by the portal branch (namely, the liver segment), in which there may be an increased risk of recurrence. However, the superiority of AR has been never proven, and several reports are available in favour or against this hypothesis[11,16]. More importantly, according to the theory, AR should completely eliminate the risk of local recurrence (relapse at the surgical edge), by eliminating the area where the tumour may have spread. However, our and others data[17,18] have reported a comparable rate of local recurrence among AR and non-anatomical resections, questioning the ability of a radical segment resection to control the oncological burden. Thus, the highest rate of intra-hepatic recurrence occurred in other liver segments than the one carrying the primitive nodule, suggesting a different or at least a concomitant route of the tumour cells. More recently, we tried to identify the risk factors for either local or intra-hepatic distant recurrence in a large European series[19], observing that local relapse occurred frequently in cases of positive surgical margin (and consequently as a kind of surgical failure), while the presence of microvascular invasion and satellitosis were hallmarks of increased risk of intra-hepatic distant relapse. These data suggest that, when those histological features occurred, the tumour may have already invaded the blood circulation, with a metastatisation potential in other locations that may not be explained by the local portal flow, and that cannot be controlled by modifying the extent of surgery. In this sense, the tumour micro-thrombi assessed by histology near the primitive nodule could not be considered only a local extension of the disease (as supposed by the portal flow theory), but a sign of systemic dissemination. Another ‘brick in the wall’ was suggested by the clinical data: recently, Hidaka *et al*[20] reported that the complete removal of the portal-bearing area did not modify the risk of recurrence in cases of microvascular invasion, and this data was confirmed in our recent meta-analysis [11].

Sakon *et al*[12,21] studied the tumour blood flow (TBF) area, discovering that this coincided with the segmental portal area only in 18% of their cohort. In up to 75% of cases, the TBF was independent of the PV area, and the rate of recurrence was reduced only in cases where the TBF was completely included in the resection area, regardless of the removal of the liver segment. The authors proposed a subclassification of HCC recurrence based on two different mechanisms: local recurrence, which is driven by the invasion of the local tumour blood flow with a peritumoural dissemination, and a systemic dissemination driven by the spread of CTCs derived from the primitive nodule, which may be able to “rehome” after passing through the systemic circulation. While the first mechanism could be controlled by an effective radical resection, the second relies on the oncological progression of the tumour, and could explain cases of intra-hepatic relapse at a distance from the original site, but also recurrence after transplantation. In 2018, a very interesting study was conducted by Sun *et al*[22], who

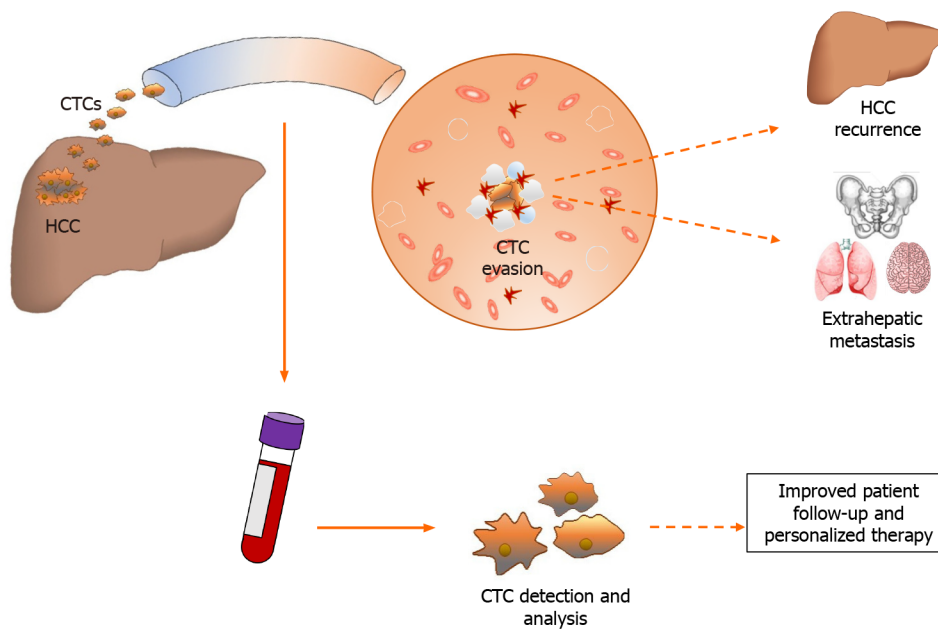
tested the spatial heterogeneity of phenotypic and molecular characteristics of CTCs within the circulatory system, discovering that a higher number of CTCs were detected in sites other than the PV. In particular, the percentages of CTCs detected in blood sampled from a peripheral vein, peripheral artery, hepatic veins, infrahepatic inferior vena cava, and PV before HCC resection were 68.5%, 45.2%, 80.8%, 39.7%, and 58.9%, respectively. Moreover, CTC and circulating tumour microemboli burden detected in hepatic veins and peripheral circulation, but not in the PV, were associated with postoperative lung metastasis and intrahepatic recurrence, respectively. These pieces of evidence suggest that the classical recurrence theory for HCC cannot explain many real-scenario observations. A novel approach, integrating the discovery of CTCs and their role in tumour biology with clinical experience, will allow a novel and tailored approach to select the best candidates for curative strategies, but will also be able to provide a novel biomarker with the ability to summarise the biological data of the tumour, using a very simple blood sample analysis (Figure 1). The detection of these cells and their possible role in surgery will be further explored in the following sections.

## CTCS MOLECULAR CHARACTERISTICS

In the context of cancer pathogenesis, and especially for carcinomas, the “epithelial-mesenchymal transition” (EMT) is a fundamental mechanism playing a key role in the metastatic process[23]. Several authors agree that this rearrangement of cell status is neither stable nor binary, and neoplastic epithelial cells that have activated an EMT program very rarely advance to a fully mesenchymal state[24]. Also, the reverse process known as mesenchymal-epithelial transition (MET) is required for metastatic colonisation in the same or other tissues[25]. Both of the above-mentioned mechanisms can actively operate in the generation of CTCs. Since CTCs are a phenotypically distinct subpopulation that originate from the tumour microenvironment, the idea behind the identification of CTCs is to discover characteristic markers of both EMT/MET transition and of the primary tumour.

CTC identification is technically difficult due to the low concentration of these cells in blood[26]. In recent years, research has focused on improving the specificity and sensitivity of CTC detection and facilitating accurate molecular characterisation[14]. Based on physical and/or biological properties of the cells, several strategies and systems have been developed to improve CTC enrichment. Filter membranes, such as the CanPatrol™ system, and microfluidic devices, such as CTC-iChip and Labyrinth-chip, allow separation of cells based on their sizes[14,27]. Alternatively, Ficoll-type density gradient methods make it easier to separate blood cells, exploiting their different density[28].

One of the most used methods is the Cell Search® system, which is based on immunomagnetic enrichment[29]. This CTC isolation strategy exploits the expression on the cell surface of the protein EpCAM, which is the most accredited marker for positive affinity-selection of CTCs. The Cell Search® system is the only system approved by the Food and Drug Administration (FDA) to predict the outcome of patients affected by breast cancer[30]. Nevertheless, enumeration of EpCAM<sup>+</sup> CTCs alone has demonstrated modest clinical sensitivity and, for instance, in cancers with low EpCAM expression, the Cell Search® system showed a lower CTC recovery rate compared to microfluidic devices[31]. In 2018, Pang *et al*[32] developed a method which exploits the surface-enhanced Raman scattering (SERS) technology and nanoparticles linked to antibodies directed against the specific hepatic proteins asialoglycoprotein receptor (ASGPR) and glypican-3 (GPC3), allowing isolation of EpCAM<sup>+</sup> CTCs. Moreover, not all CTCs have metastatic or relapsing potential, so simple quantification without better molecular characterisation could lead to incorrect clinical conclusions. The use of isolation and enrichment devices is supported by other laboratory techniques such as immunofluorescence staining of different markers [fluorescence-activated cell sorting (FACS) and fluorescent in situ hybridization (FISH)] and/or gene expression analysis [real time PCR (qPCR) and single cell RNA sequencing (scRNA-seq)] in order to obtain in depth CTC characterisation[33]. Considering all of these biological /phenotypic and experimental issues, the application of this method in common clinical practice has proved to be difficult. Making this strategy even harder is the heterogeneity of the tumour itself and, among protein markers, cytokeratins (CKs), vimentin, CD44, CD133 and CD90 are the most used so far[34]. CKs, like EpCAM, are epithelial markers, but unlike the latter, they are intracellular proteins, thus they are identified mainly using immunocytochemistry. Cells are usually stained for CK8, 18



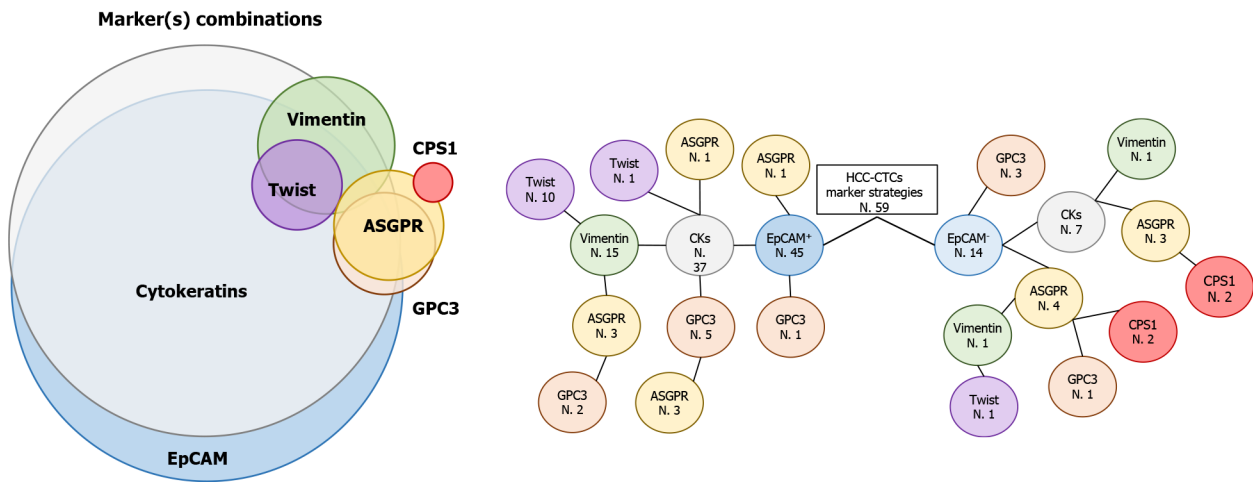
**Figure 1 Hepatocellular carcinoma releases tumor cells into circulation where they become circulating tumor cells.** Circulating tumor cells after evading the immune system can cause hepatocellular carcinoma recurrence or distant metastasis. The liquid biopsy intercepts these cells into the bloodstream and allows to study the tumor characteristics and guarantee personalized therapy. CTCs: Circulating tumor cells; HCC: Hepatocellular carcinoma.

and 19, but recently other markers such as human epidermal growth factor receptor 2 (HER2) and the estrogen receptor (ER) have been examined to facilitate detection of CTCs with metastatic potential[35].

As mentioned above, the major drawback of using epithelial markers is their inability to detect CTCs that no longer express them after undergoing EMT, a process which is strongly associated with overexpression of vimentin and CD44[36,37]. CD44 is often used as a marker in combination with the stem-like markers CD133 and CD90 [38]. However, plasma membrane and cytoplasmic proteins are not the only markers used to detect potential CTCs; complex studies have tried to generate the mRNA expression profiles of CTCs in different diseases[39]. In particular, D'Avola *et al*[33] recently developed a new method that sequentially combines image flow cytometry and high density scRNA-seq in order to identify CTCs in patients with HCC. The authors suggest the advantages of genome-wide transcriptome profiling to confidently detect CTCs and its potential role in monitoring HCC heterogeneity and detecting HCC driver genes, which could ultimately help customize therapeutic interventions in these patients.

## CTCS IDENTIFICATION IN HCC PATIENTS

To date, there are no specific or accredited CTC-related protocols for detection of HCC that are agreed upon by the scientific community. Several studies have been performed to deeply investigate and introduce the use of CTC enumeration/characterisation in HCC monitoring in clinical practice. Most of these studies are primarily based on the previously validated EpCAM/CK markers, with secondary examination of other markers or features (Figure 2). In recent years, several authors in the HCC field have taken advantage of combining the markers vimentin and twist; as mentioned above, these mesenchymal markers have followed the common epithelial markers EpCAM/CKs. Ou *et al*[40] observed that the presence of mesenchymal CTCs tended to occur in advanced stage patients and was associated with earlier recurrence in a large cohort of HCC patients. ASGPR and carbamoyl-phosphate synthetase 1 (CPS1) are interesting in the context of HCC. Liu *et al*[41] demonstrated that CTC enrichment, combined with identification using an antibody cocktail against ASGPR and CPS1, not only significantly improves sensitivity for CTC enrichment, but also provides high specificity for CTC detection in patients with HCC, thereby minimising false negative/positive results. The combination of ASGPR and CPS1 was used also by Li *et al*[42], confirming the increased sensitivity for HCC CTC detection.



**Figure 2** Frequently used markers for circulating tumor cell detection hepatocellular carcinoma-related. Epithelial cell adhesion molecule+ circulating tumor cell identification is often combined with cytokeratins, vimentin, twist, Glypican-3 and asialoglycoprotein receptor (ASGPR). In some cases, ASGPR is also used with the hepatocellular marker carbamoyl-phosphate synthetase 1 (Venn diagram). Tree diagram show the number of articles (N.) that use different marker combinations for circulating tumor cell isolation. Bibliography counts articles related to hepatocellular carcinoma field and published from 2009 to 2020. EpCAM: Epithelial cell adhesion molecule; CKs: Cytokeratins; ASGPR: Asialoglycoprotein receptor; GPC3: Glypican-3; CPS1: Carbamoyl-phosphate synthetase 1.

In 2016, Zhang *et al*[43] isolated ASGPR+/CPS1+ CTCs from HCC patients, which were then cultured and expanded to form spheroid-like structures in a 3D cell culture assay. They suggested that this method could aid physicians in the selection of appropriate drug therapies for HCC patients. The role of CTCs expressing mesenchymal features in predicting HCC early recurrence was confirmed in the same year by Qi *et al*[44], in a monocentric study with 112 enrolled patients. However, mesenchymal CTC use in clinical practice is controversial, since their analysis in a different cohort of HCC patients who underwent liver transplantation was not able to predict HCC recurrence[45]. Conversely, in liver transplantation, the entire organ is replaced with a healthy liver deriving from a donor. Thus, the HCC recurrence is likely due to circulating and/or dormant tumour cells, which have acquired the ability to escape from the host’s immune system. Clusters of CTCs were first predicted and then observed as intravascular tumour microemboli, represented by multicellular epithelial tumour cells. In a mouse model experiment, in which human-derived CTCs were used, it was observed that CTC clusters are not derived from intravascular aggregation of single CTCs or from the progeny of a single primary tumour cell that proliferates in the vascular space, but instead, evidence showed that CTC clusters derive from groupings of primary tumour cells that enter the bloodstream together [46].

### CTCS AS A MOLECULAR SIGNATURE OF THE HISTOLOGICAL CHARACTERISTICS OF THE PRIMITIVE HCC

Recently it has been reported that CTCs positive for EpCAM, N-Cadherin and CD90 expression (triple positive CTCs) are more frequently associated with microvascular invasion (MVI), as detected in a histological specimen after liver resection[47]. The histopathological finding of MVI is a feature of advanced HCC, associated with a higher probability of recurrence and metastasis[48]; however, with the imaging tests and biomarkers currently available, the preoperative identification of MVI remains difficult[49]. Rodriguez-Perálvarez *et al*[50] showed that MVI incidence was between 15.0% and 57.1% at histopathological examination after liver resection and transplantation, in a systematic review. Thus, different tumour stages and HCC invasive characteristics affect MVI incidence. The possibility given by the triple positive CTCs, associated with the actual diagnostics tool, to pre-operatively identify MVI may play a role in the use of preoperative predictive models in therapeutic decision-making in patients with HCC.

## CTCS VARIATIONS AFTER LIVER RESECTION

After surgical tumor excision, CTC levels drop dramatically and post-operative CTC levels can be used as tools to verify surgical resection as a monitor for tumor burden [14]. Yu *et al*[51] evaluated the effect of surgical liver resection on CTCs in patients with HCC, demonstrating that a lower CTC level after surgical resection is an independent prognostic factor for better disease-free-survival (HR 0.620; 95%CI: 0.479–0.803;  $P < 0.001$ ) and overall-survival (HR 0.608; 95%CI: 0.443–0.834;  $P = 0.002$ ). Ou *et al*[40] demonstrated that increased CTC numbers were observed in patients with high levels ( $> 400$  mcg/L) of alpha fetoprotein (AFP), advanced TNM and BCLC stage, and the presence of embolus or microembolus. They also investigated CTC heterogeneity, noting a significant correlation between mesenchymal CTCs and high AFP levels, multiple tumours, advanced TNM and BCLC stage, presence of embolus or microembolus, and earlier recurrence.

These CTCs could be considered as a very early sign of tumour migration: invisible micro-metastasis, impossible to detect with standard methods but playing a fundamental role in patients' clinical evolution[52]. Sun *et al*[53] analysed the diagnostic value of CTCs in HCC patients, performing a meta-analysis on 20 studies of a total of 998 HCC patients. From their work, it emerges that CTC positivity is associated with a lower overall survival (HR 2.417; 95%CI: 1.421–3.250;  $P < 0.001$ ) and disease free survival (HR 3.59; 95%CI: 1.984–6.495;  $P < 0.001$ ). CTC analysis determines the tumour molecular characteristics before any treatment, evaluating cancer differentiation and identifying markers as possible molecular therapy targets or mechanisms of resistance to therapy[54]. The selective pressure that develops over time since starting the treatment leads to increased cellular heterogeneity of the tumour, production of drug resistant subclones, and the selection of rare mutants[55], essentially the tumour is characterised by different genetic backgrounds at different times. Therefore, the tumour genome during follow-up could differ significantly from its initial state, and this difference cannot be assessed unless repeated sampling is performed. However, repeat biopsy is rarely feasible and, without knowledge of the genetic changes, complete treatment personalisation and targeted therapy is impossible[56]. In comparison, liquid biopsy is easily repeatable during follow-up, making knowledge of all tumour genome changes possible. In the future, it will be desirable to use quantitative and qualitative analysis of CTCs to develop personalised therapy for each patient. The phenotyping of those cells, and their quantification in the peripheral blood, may allow identification of patients with a more severe and more aggressive disease, who could be the target population in which adjuvant therapies as Sorafenib may play a role. Currently, it is not evident what features are associated with response to such treatments[57]. The presence and characteristics of the CTCs identified in peripheral blood may become a molecular marker to decide the follow-up schedule, and estimations of risk could be updated at each visit by repeating the test. In other words, CTCs could become a new predictive marker to better stratify patients and assign them to the best individual treatment plan, improving long-term cancer outcomes.

## CTCS AND LIVER TRANSPLANT FOR HCC

Orthotopic liver transplantation (OLT) is the most favourable option for the treatment of HCC, with a 5-year overall survival rate of 75% and disease-free survival rate of 83% [58]. Despite stringent criteria in patient selection for transplantation, HCC recurrence still remains a significant problem, with a rate of 15%–20% [59,60]. Due to organ shortage and recurrence risk even after transplantation, it is important to be able to select patients for LT in order not to misallocate a limited resource.

Tumour size, AFP levels, and micro- and macro-vascular invasion are the main prognostic factors for recurrence risk after transplantation[61]. The aim of patient selection criteria should be to prevent transplantation in those patients with an expected HCC recurrence and to improve transplantation for those patients who have a high likelihood of being cured. The present parameters are based on morphology, but in the modern molecular era, new information could be available to better understand the patient's tumour biology in a tailored fashion.

Xu *et al*[62] highlighted how the CTC-positive rate and number of CTCs present is higher in patients beyond the Milan criteria than in patients within the criteria (91% *vs* 69%,  $P = 0.009$ ; and  $27 \pm 27$  *vs*  $6 \pm 9$ ,  $P < 0.001$ ; respectively). This suggests that including the CTC count in pre-transplant evaluation could revolutionise the eligi-

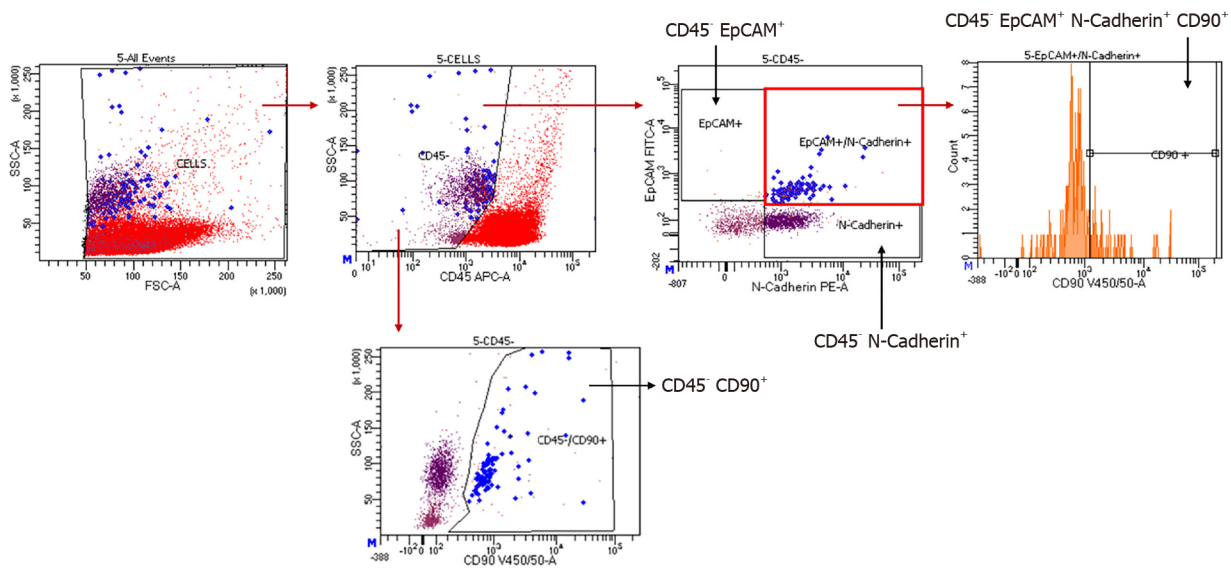


Figure 3 Representative example of a patient's PBMC analysis within the FINDINGBIOREC protocol.

bility criteria for transplantation. Chen *et al*[63] analysed preoperative CTCs in HCC patients who underwent LT and followed them up for at least one year, or until relapse or death occurred. They found that recurrence is associated with presence of preoperative CTCs ( $P = 0.013$ ); multivariate analysis confirmed that CTCs are an independent risk factor for the onset of recurrence after LT (HR: 5.411; 95%CI: 1.132–25,874;  $P = 0.034$ ). These data reflect the 1-year DFS rate, which is 91.6% for the CTC-negative and 61.5% for the CTC-positive group ( $P = 0.020$ ). On the other hand, the 1-year overall survival rate for the CTC-negative and CTC-positive group is 91.7% and 88.5%, respectively, with no significant difference. Very few data are available about the potential role of CTCs as preoperative predictors of HCC recurrence after LT, and it is still a controversial issue. However, their application could drastically change the allocation protocols, enabling a more tailored algorithm with potentially better ability to predict the risk of relapse and, consequently, differentiate the cases that could benefit from transplant from the ones that could not.

## CTCS IDENTIFICATION IN A REAL-CLINICAL SCENARIO: THE FINDINGBIOREC PROTOCOL

In light of the previously mentioned data, the University of Milano-Bicocca, the University of Piemonte Orientale and Humanitas University have decided to collaborate by creating a study with the aim of "finding the seeds of recurrence", using liquid biopsy to detect CTCs as markers of disease and prognosis in HCC.

Our hypothesis is that CTCs may spread from the original tumours as a hallmark of advanced cancer, which has already developed the characteristic of invasiveness. It is our opinion that early stage tumours do not release CTCs into the bloodstream at the same rate or quality as advanced tumours. Patients with a positive CTC liquid biopsy may have a worse prognosis, due to an increased relapse rate. Finally, from a pathophysiological point of view, we want to demonstrate that recurrence is due to CTC seeding, in order to gain a better understanding of HCC carcinogenesis. The "FINDINGBIOREC" study (clinicaltrial.gov ID: NCT04800497) was developed: a prospective, observational cohort study, conducted in two tertiary referral centres for liver cancer, in which each enrolled patient is submitted to liquid biopsy prior to surgery and then every 3 mo during the follow-up schedules, for 3 years. Patients with a first diagnosis of HCC, no previous treatment for this condition, no other oncological history, and BCLC stage 0-A-B are prospectively enrolled. The samples are processed and the CTCs are detected using FACSsymphony™ with subsequent identification of the following markers: EpCAM, N-cadherin (N-cad) and CD90 (Figure 3). Patients are followed up with clinical assessments; CT or, where necessary, MRI and AFP level, together with liquid biopsy. With this protocol, we aim to better highlight the trends of CTCs at different time-points and their correlation with the oncologic prognosis in very early



and early HCC. The study is currently enrolling, and it will be closed in 2023.

## CONCLUSION

HCC may produce early CTCs, which seem to be the seed of the recurrence. Their presence in the blood stream has been correlated with the presence of MVI, suggesting that the latter is a surrogate sign of a systemic disease that cannot be controlled by classical liver segment resection alone. Those cells could be detected and studied by liquid biopsy, which is a safe method to obtain information on the patient's disease status. This allows tumour molecular characterisation during different disease phases, and could become a new method for patient stratification. The study of CTCs allows selection of patients and the type of treatment they will receive in order to optimize HCC therapy. During the follow-up, an increase in CTCs makes it possible to identify tumour recurrence and implement further therapy early. In future, liquid biopsy could be implemented in the pre- and post-operative routine of HCC patients in order to gain more accurate information on tumour type and stage, and guarantee the most personalised therapy possible for the patients.

## REFERENCES

- 1 **Lin L**, Yan L, Liu Y, Qu C, Ni J, Li H. The Burden and Trends of Primary Liver Cancer Caused by Specific Etiologies from 1990 to 2017 at the Global, Regional, National, Age, and Sex Level Results from the Global Burden of Disease Study 2017. *Liver Cancer* 2020; **9**: 563-582 [PMID: 33083281 DOI: 10.1159/000508568]
- 2 **Forner A**, Llovet JM, Bruix J. Hepatocellular carcinoma. *Lancet* 2012; **379**: 1245-1255 [PMID: 22353262 DOI: 10.1016/S0140-6736(11)61347-0]
- 3 **Tabrizian P**, Jibara G, Shrager B, Schwartz M, Roayaie S. Recurrence of hepatocellular cancer after resection: patterns, treatments, and prognosis. *Ann Surg* 2015; **261**: 947-955 [PMID: 25010665 DOI: 10.1097/SLA.0000000000000710]
- 4 **Abdelaziz AO**, Nabil MM, Abdelmaksoud AH, Shousha HI, Cordie AA, Hassan EM, Omran DA, Leithy R, Elbaz TM. De-novo vs recurrent hepatocellular carcinoma following direct-acting antiviral therapy for hepatitis C virus. *Eur J Gastroenterol Hepatol* 2018; **30**: 39-43 [PMID: 29064851 DOI: 10.1097/MEG.0000000000001004]
- 5 **Yamamoto S**, Midorikawa Y, Nagae G, Tatsuno K, Ueda H, Moriyama M, Takayama T, Aburatani H. Spatial and temporal expansion of intrahepatic metastasis by molecularly-defined clonality in multiple liver cancers. *Cancer Sci* 2020; **111**: 601-609 [PMID: 31845427 DOI: 10.1111/cas.14282]
- 6 **Wang B**, Xia CY, Lau WY, Lu XY, Dong H, Yu WL, Jin GZ, Cong WM, Wu MC. Determination of clonal origin of recurrent hepatocellular carcinoma for personalized therapy and outcomes evaluation: a new strategy for hepatic surgery. *J Am Coll Surg* 2013; **217**: 1054-1062 [PMID: 24246620 DOI: 10.1016/j.jamcollsurg.2013.07.402]
- 7 **Carone C**, Olivani A, Dalla Valle R, Manuguerra R, Silini EM, Trenti T, Missale G, Cariani E. Immune Gene Expression Profile in Hepatocellular Carcinoma and Surrounding Tissue Predicts Time to Tumor Recurrence. *Liver Cancer* 2018; **7**: 277-294 [PMID: 30319985 DOI: 10.1159/000486764]
- 8 **Makuuchi M**, Hasegawa H, Yamazaki S. Ultrasonically guided subsegmentectomy. *Surg Gynecol Obstet* 1985; **161**: 346-350 [PMID: 2996162 DOI: 10.1055/s-2007-1022639]
- 9 **Mitsunobu M**, Toyosaka A, Oriyama T, Okamoto E, Nakao N. Intrahepatic metastases in hepatocellular carcinoma: the role of the portal vein as an efferent vessel. *Clin Exp Metastasis* 1996; **14**: 520-529 [PMID: 8970582 DOI: 10.1007/BF00115112]
- 10 **Yamanaka N**, Okamoto E, Fujihara S, Kato T, Fujimoto J, Oriyama T, Mitsunobu M, Toyosaka A, Uematsu K, Yamamoto K. Do the tumor cells of hepatocellular carcinomas dislodge into the portal venous stream during hepatic resection? *Cancer* 1992; **70**: 2263-2267 [PMID: 1327495 DOI: 10.1002/1097-0142(19921101)70:9<2263::aid-cnrcr2820700909>3.0.co;2-m]
- 11 **Famularo S**, Ceresoli M, Gianti A, Ciulli C, Pinotti E, Romano F, Braga M, De Carlis L, Gianotti L. Is It Just a Matter of Surgical Extension to Achieve the Cure of Hepatocarcinoma? *J Gastrointest Surg* 2021; **25**: 94-103 [PMID: 31898106 DOI: 10.1007/s11605-019-04494-5]
- 12 **Sakon M**, Ogawa H, Fujita M, Nagano H. Hepatic resection for hepatocellular carcinoma based on tumor hemodynamics. *Hepatol Res* 2013; **43**: 155-164 [PMID: 23194466 DOI: 10.1111/hepr.12001]
- 13 **Ye Q**, Ling S, Zheng S, Xu X. Liquid biopsy in hepatocellular carcinoma: circulating tumor cells and circulating tumor DNA. *Mol Cancer* 2019; **18**: 114 [PMID: 31269959 DOI: 10.1186/s12943-019-1043-x]
- 14 **Okajima W**, Komatsu S, Ichikawa D, Miyamae M, Ohashi T, Imamura T, Kiuchi J, Nishibeppu K, Arita T, Konishi H, Shiozaki A, Morimura R, Ikoma H, Okamoto K, Otsuji E. Liquid biopsy in patients with hepatocellular carcinoma: Circulating tumor cells and cell-free nucleic acids. *World J Gastroenterol* 2017; **23**: 5650-5668 [PMID: 28883691 DOI: 10.3748/wjg.v23.i31.5650]
- 15 **Okuda K**, Peters RL. Hepatocellular Carcinoma. John Wiley & Sons. Available from:

- [https://books.google.com/books/about/Hepatocellular\\_Carcinoma.html?hl=&id=YX9rAAAAAAAJ](https://books.google.com/books/about/Hepatocellular_Carcinoma.html?hl=&id=YX9rAAAAAAAJ)
- 16 **Shindoh J**, Kobayashi Y, Umino R, Kojima K, Okubo S, Hashimoto M. Successful Anatomic Resection of Tumor-Bearing Portal Territory Delays Long-Term Stage Progression of Hepatocellular Carcinoma. *Ann Surg Oncol* 2021; **28**: 844-853 [PMID: 32712886 DOI: [10.1245/s10434-020-08927-3](https://doi.org/10.1245/s10434-020-08927-3)]
  - 17 **Famularo S**, Di Sandro S, Giani A, Lauterio A, Sandini M, De Carlis R, Buscemi V, Uggeri F, Romano F, Gianotti L, De Carlis L. Recurrence Patterns After Anatomic or Parenchyma-Sparing Liver Resection for Hepatocarcinoma in a Western Population of Cirrhotic Patients. *Ann Surg Oncol* 2018; **25**: 3974-3981 [PMID: 30244421 DOI: [10.1245/s10434-018-6730-0](https://doi.org/10.1245/s10434-018-6730-0)]
  - 18 **Marubashi S**, Gotoh K, Akita H, Takahashi H, Sugimura K, Miyoshi N, Motoori M, Kishi K, Noura S, Fujiwara Y, Ohue M, Nakazawa T, Nakanishi K, Ito Y, Yano M, Ishikawa O, Sakon M. Analysis of Recurrence Patterns After Anatomical or Non-anatomical Resection for Hepatocellular Carcinoma. *Ann Surg Oncol* 2015; **22**: 2243-2252 [PMID: 25373536 DOI: [10.1245/s10434-014-4214-4](https://doi.org/10.1245/s10434-014-4214-4)]
  - 19 **Famularo S**, Piardi T, Molfino S, Di Martino M, Ferrari C, Ielpo B, Diago MV, Giani A, Griseri G, Terés LB, Gianotti L, Baiocchi GL, Sommacale D, Romano F. Factors Affecting Local and Intra Hepatic Distant Recurrence After Surgery for Hcc: An Alternative Perspective on Microvascular Invasion and Satellitosis - A Western European Multicentre Study. *J Gastrointest Surg* 2021; **25**: 104-111 [PMID: 31965441 DOI: [10.1007/s11605-019-04503-7](https://doi.org/10.1007/s11605-019-04503-7)]
  - 20 **Hidaka M**, Eguchi S, Okuda K, Beppu T, Shirabe K, Kondo K, Takami Y, Ohta M, Shiraishi M, Ueno S, Nanashima A, Noritomi T, Kitahara K, Fujioka H. Impact of Anatomical Resection for Hepatocellular Carcinoma With Microportal Invasion (vp1): A Multi-institutional Study by the Kyushu Study Group of Liver Surgery. *Ann Surg* 2020; **271**: 339-346 [PMID: 30048313 DOI: [10.1097/SLA.0000000000002981](https://doi.org/10.1097/SLA.0000000000002981)]
  - 21 **Sakon M**, Kobayashi S, Wada H, Eguchi H, Marubashi S, Takahashi H, Akita H, Gotoh K, Yamada D, Asukai K, Hasegawa S, Ohue M, Yano M, Nagano H. "Logic-Based Medicine" Is More Feasible than "Evidence-Based Medicine" in the Local Treatment for Hepatocellular Carcinoma. *Oncology* 2020; **98**: 259-266 [PMID: 32045926 DOI: [10.1159/000505554](https://doi.org/10.1159/000505554)]
  - 22 **Sun YF**, Guo W, Xu Y, Shi YH, Gong ZJ, Ji Y, Du M, Zhang X, Hu B, Huang A, Chen GG, Lai PBS, Cao Y, Qiu SJ, Zhou J, Yang XR, Fan J. Circulating Tumor Cells from Different Vascular Sites Exhibit Spatial Heterogeneity in Epithelial and Mesenchymal Composition and Distinct Clinical Significance in Hepatocellular Carcinoma. *Clin Cancer Res* 2018; **24**: 547-559 [PMID: 29070526 DOI: [10.1158/1078-0432.CCR-17-1063](https://doi.org/10.1158/1078-0432.CCR-17-1063)]
  - 23 **van Zijl F**, Zulehner G, Petz M, Schneller D, Kornauth C, Hau M, Machat G, Grubinger M, Huber H, Mikulits W. Epithelial-mesenchymal transition in hepatocellular carcinoma. *Future Oncol* 2009; **5**: 1169-1179 [PMID: 19852728 DOI: [10.2217/fon.09.91](https://doi.org/10.2217/fon.09.91)]
  - 24 **Dongre A**, Weinberg RA. New insights into the mechanisms of epithelial-mesenchymal transition and implications for cancer. *Nat Rev Mol Cell Biol* 2019; **20**: 69-84 [PMID: 30459476 DOI: [10.1038/s41580-018-0080-4](https://doi.org/10.1038/s41580-018-0080-4)]
  - 25 **Gunasinghe NP**, Wells A, Thompson EW, Hugo HJ. Mesenchymal-epithelial transition (MET) as a mechanism for metastatic colonisation in breast cancer. *Cancer Metastasis Rev* 2012; **31**: 469-478 [PMID: 22729277 DOI: [10.1007/s10555-012-9377-5](https://doi.org/10.1007/s10555-012-9377-5)]
  - 26 **Edd JF**, Mishra A, Dubash TD, Herrera S, Mohammad R, Williams EK, Hong X, Mutlu BR, Walsh JR, Machado de Carvalho F, Aldikacti B, Nieman LT, Stott SL, Kapur R, Maheswaran S, Haber DA, Toner M. Microfluidic concentration and separation of circulating tumor cell clusters from large blood volumes. *Lab Chip* 2020; **20**: 558-567 [PMID: 31934715 DOI: [10.1039/c9lc01122f](https://doi.org/10.1039/c9lc01122f)]
  - 27 **Wan S**, Kim TH, Smith KJ, Delaney R, Park GS, Guo H, Lin E, Plegue T, Kuo N, Steffes J, Leu C, Simeone DM, Razimulava N, Parikh ND, Nagrath S, Welling TH. New Labyrinth Microfluidic Device Detects Circulating Tumor Cells Expressing Cancer Stem Cell Marker and Circulating Tumor Microemboli in Hepatocellular Carcinoma. *Sci Rep* 2019; **9**: 18575 [PMID: 31819089 DOI: [10.1038/s41598-019-54960-y](https://doi.org/10.1038/s41598-019-54960-y)]
  - 28 **Gertler R**, Rosenberg R, Fuehrer K, Dahm M, Nekarda H, Siewert JR. Detection of circulating tumor cells in blood using an optimized density gradient centrifugation. *Recent Results Cancer Res* 2003; **162**: 149-155 [PMID: 12790329 DOI: [10.1007/978-3-642-59349-9\\_13](https://doi.org/10.1007/978-3-642-59349-9_13)]
  - 29 **Wang PX**, Sun YF, Zhou KQ, Cheng JW, Hu B, Guo W, Yin Y, Huang JF, Zhou J, Fan J, Cheung TT, Qu XD, Yang XR. Circulating tumor cells are an indicator for the administration of adjuvant transarterial chemoembolization in hepatocellular carcinoma: A single-center, retrospective, propensity-matched study. *Clin Transl Med* 2020; **10**: e137 [PMID: 32702202 DOI: [10.1002/ctm2.137](https://doi.org/10.1002/ctm2.137)]
  - 30 **Cristofanilli M**, Budd GT, Ellis MJ, Stopeck A, Matera J, Miller MC, Reuben JM, Doyle GV, Allard WJ, Terstappen LW, Hayes DF. Circulating tumor cells, disease progression, and survival in metastatic breast cancer. *N Engl J Med* 2004; **351**: 781-791 [PMID: 15317891 DOI: [10.1056/NEJMoa040766](https://doi.org/10.1056/NEJMoa040766)]
  - 31 **Sánchez-Lorencio MI**, Ramirez P, Saenz L, Martínez Sánchez MV, De La Orden V, Mediero-Valeros B, Veganzones-De-Castro S, Baroja-Mazo A, Revilla Nuin B, Gonzalez MR, Cascales-Campos PA, Noguera-Velasco JA, Minguela A, Díaz-Rubio E, Pons JA, Parrilla P. Comparison of Two Types of Liquid Biopsies in Patients With Hepatocellular Carcinoma Awaiting Orthotopic Liver Transplantation. *Transplant Proc* 2015; **47**: 2639-2642 [PMID: 26680058 DOI: [10.1016/j.transproceed.2015.10.003](https://doi.org/10.1016/j.transproceed.2015.10.003)]
  - 32 **Pang Y**, Wang C, Xiao R, Sun Z. Dual-Selective and Dual-Enhanced SERS Nanoprobes Strategy for

- Circulating Hepatocellular Carcinoma Cells Detection. *Chemistry* 2018; **24**: 7060-7067 [PMID: 29521467 DOI: 10.1002/chem.201801133]
- 33 **D'Avola D**, Villacorta-Martin C, Martins-Filho SN, Craig A, Labgaa I, von Felden J, Kimaada A, Bonaccorso A, Tabrizian P, Hartmann BM, Sebra R, Schwartz M, Villanueva A. High-density single cell mRNA sequencing to characterize circulating tumor cells in hepatocellular carcinoma. *Sci Rep* 2018; **8**: 11570 [PMID: 30068984 DOI: 10.1038/s41598-018-30047-y]
  - 34 **Yagci T**, Cetin M, Ercin PB. Cancer Stem Cells in Hepatocellular Carcinoma. *J Gastrointest Cancer* 2017; **48**: 241-245 [PMID: 28643126 DOI: 10.1007/s12029-017-9960-7]
  - 35 **Masuda T**, Hayashi N, Iguchi T, Ito S, Eguchi H, Mimori K. Clinical and biological significance of circulating tumor cells in cancer. *Mol Oncol* 2016; **10**: 408-417 [PMID: 26899533 DOI: 10.1016/j.molonc.2016.01.010]
  - 36 **Chen C**, Zhao S, Karnad A, Freeman JW. The biology and role of CD44 in cancer progression: therapeutic implications. *J Hematol Oncol* 2018; **11**: 64 [PMID: 29747682 DOI: 10.1186/s13045-018-0605-5]
  - 37 **Satelli A**, Li S. Vimentin in cancer and its potential as a molecular target for cancer therapy. *Cell Mol Life Sci* 2011; **68**: 3033-3046 [PMID: 21637948 DOI: 10.1007/s00018-011-0735-1]
  - 38 **Guo W**, Sun YF, Shen MN, Ma XL, Wu J, Zhang CY, Zhou Y, Xu Y, Hu B, Zhang M, Wang G, Chen WQ, Guo L, Lu RQ, Zhou CH, Zhang X, Shi YH, Qiu SJ, Pan BS, Cao Y, Zhou J, Yang XR, Fan J. Circulating Tumor Cells with Stem-Like Phenotypes for Diagnosis, Prognosis, and Therapeutic Response Evaluation in Hepatocellular Carcinoma. *Clin Cancer Res* 2018; **24**: 2203-2213 [PMID: 29374055 DOI: 10.1158/1078-0432.CCR-17-1753]
  - 39 **Mostert B**, Sieuwerts AM, Bolt-de Vries J, Kraan J, Lalmahomed Z, van Galen A, van der Spoel P, de Weerd V, Ramírez-Moreno R, Smid M, Verhoef C, IJzermans JN, Gratama JW, Sleijfer S, Foekens JA, Martens JW. mRNA expression profiles in circulating tumor cells of metastatic colorectal cancer patients. *Mol Oncol* 2015; **9**: 920-932 [PMID: 25655581 DOI: 10.1016/j.molonc.2015.01.001]
  - 40 **Ou H**, Huang Y, Xiang L, Chen Z, Fang Y, Lin Y, Cui Z, Yu S, Li X, Yang D. Circulating Tumor Cell Phenotype Indicates Poor Survival and Recurrence After Surgery for Hepatocellular Carcinoma. *Dig Dis Sci* 2018; **63**: 2373-2380 [PMID: 29926241 DOI: 10.1007/s10620-018-5124-2]
  - 41 **Liu HY**, Qian HH, Zhang XF, Li J, Yang X, Sun B, Ma JY, Chen L, Yin ZF. Improved method increases sensitivity for circulating hepatocellular carcinoma cells. *World J Gastroenterol* 2015; **21**: 2918-2925 [PMID: 25780289 DOI: 10.3748/wjg.v21.i10.2918]
  - 42 **Li J**, Chen L, Zhang X, Zhang Y, Liu H, Sun B, Zhao L, Ge N, Qian H, Yang Y, Wu M, Yin Z. Detection of circulating tumor cells in hepatocellular carcinoma using antibodies against asialoglycoprotein receptor, carbamoyl phosphate synthetase 1 and pan-cytokeratin. *PLoS One* 2014; **9**: e96185 [PMID: 24763545 DOI: 10.1371/journal.pone.0096185]
  - 43 **Zhang Y**, Zhang X, Zhang J, Sun B, Zheng L, Li J, Liu S, Sui G, Yin Z. Microfluidic chip for isolation of viable circulating tumor cells of hepatocellular carcinoma for their culture and drug sensitivity assay. *Cancer Biol Ther* 2016; **17**: 1177-1187 [PMID: 27662377 DOI: 10.1080/15384047.2016.1235665]
  - 44 **Qi LN**, Xiang BD, Wu FX, Ye JZ, Zhong JH, Wang YY, Chen YY, Chen ZS, Ma L, Chen J, Gong WF, Han ZG, Lu Y, Shang JJ, Li LQ. Circulating Tumor Cells Undergoing EMT Provide a Metric for Diagnosis and Prognosis of Patients with Hepatocellular Carcinoma. *Cancer Res* 2018; **78**: 4731-4744 [PMID: 29915159 DOI: 10.1158/0008-5472.CAN-17-2459]
  - 45 **Wang S**, Zheng Y, Liu J, Huo F, Zhou J. Analysis of circulating tumor cells in patients with hepatocellular carcinoma recurrence following liver transplantation. *J Invest Med* 2018; **66**: 1-6 [PMID: 29632031 DOI: 10.1136/jim-2017-000655]
  - 46 **Aceto N**, Bardia A, Miyamoto DT, Donaldson MC, Wittner BS, Spencer JA, Yu M, Pely A, Engstrom A, Zhu H, Brannigan BW, Kapur R, Stott SL, Shioda T, Ramaswamy S, Ting DT, Lin CP, Toner M, Haber DA, Maheswaran S. Circulating tumor cell clusters are oligoclonal precursors of breast cancer metastasis. *Cell* 2014; **158**: 1110-1122 [PMID: 25171411 DOI: 10.1016/j.cell.2014.07.013]
  - 47 **Nam SJ**, Yeo HY, Chang HJ, Kim BH, Hong EK, Park JW. A New Cell Block Method for Multiple Immunohistochemical Analysis of Circulating Tumor Cells in Patients with Liver Cancer. *Cancer Res Treat* 2016; **48**: 1229-1242 [PMID: 27034142 DOI: 10.4143/crt.2015.500]
  - 48 **Hirokawa F**, Hayashi M, Asakuma M, Shimizu T, Inoue Y, Uchiyama K. Risk factors and patterns of early recurrence after curative hepatectomy for hepatocellular carcinoma. *Surg Oncol* 2016; **25**: 24-29 [PMID: 26979637 DOI: 10.1016/j.suronc.2015.12.002]
  - 49 **Xu X**, Zhang HL, Liu QP, Sun SW, Zhang J, Zhu FP, Yang G, Yan X, Zhang YD, Liu XS. Radiomic analysis of contrast-enhanced CT predicts microvascular invasion and outcome in hepatocellular carcinoma. *J Hepatol* 2019; **70**: 1133-1144 [PMID: 30876945 DOI: 10.1016/j.jhep.2019.02.023]
  - 50 **Rodríguez-Perálvarez M**, Luong TV, Andrea L, Meyer T, Dhillon AP, Burroughs AK. A systematic review of microvascular invasion in hepatocellular carcinoma: diagnostic and prognostic variability. *Ann Surg Oncol* 2013; **20**: 325-339 [PMID: 23149850 DOI: 10.1245/s10434-012-2513-1]
  - 51 **Yu JJ**, Xiao W, Dong SL, Liang HF, Zhang ZW, Zhang BX, Huang ZY, Chen YF, Zhang WG, Luo HP, Chen Q, Chen XP. Effect of surgical liver resection on circulating tumor cells in patients with hepatocellular carcinoma. *BMC Cancer* 2018; **18**: 835 [PMID: 30126375 DOI: 10.1186/s12885-018-4744-4]
  - 52 **Schulze K**, Gasch C, Staufer K, Nashan B, Lohse AW, Pantel K, Riethdorf S, Wege H. Presence of

- EpCAM-positive circulating tumor cells as biomarker for systemic disease strongly correlates to survival in patients with hepatocellular carcinoma. *Int J Cancer* 2013; **133**: 2165-2171 [PMID: 23616258 DOI: 10.1002/ijc.28230]
- 53 **Sun C**, Liao W, Deng Z, Li E, Feng Q, Lei J, Yuan R, Zou S, Mao Y, Shao J, Wu L, Zhang C. The diagnostic value of assays for circulating tumor cells in hepatocellular carcinoma: A meta-analysis. *Medicine (Baltimore)* 2017; **96**: e7513 [PMID: 28723763 DOI: 10.1097/MD.00000000000007513]
- 54 **Micalizzi DS**, Maheswaran S, Haber DA. A conduit to metastasis: circulating tumor cell biology. *Genes Dev* 2017; **31**: 1827-1840 [PMID: 29051388 DOI: 10.1101/gad.305805.117]
- 55 **McGranahan N**, Swanton C. Biological and therapeutic impact of intratumor heterogeneity in cancer evolution. *Cancer Cell* 2015; **27**: 15-26 [PMID: 25584892 DOI: 10.1016/j.ccell.2014.12.001]
- 56 **Zhou J**, Huang A, Yang XR. Liquid Biopsy and its Potential for Management of Hepatocellular Carcinoma. *J Gastrointest Cancer* 2016; **47**: 157-167 [PMID: 26969471 DOI: 10.1007/s12029-016-9801-0]
- 57 **Pinyol R**, Montal R, Bassaganyas L, Sia D, Takayama T, Chau GY, Mazzaferro V, Roayaie S, Lee HC, Kokudo N, Zhang Z, Torrecilla S, Moeini A, Rodriguez-Carunchio L, Gane E, Verslype C, Croitoru AE, Cillo U, de la Mata M, Lupo L, Strasser S, Park JW, Camps J, Solé M, Thung SN, Villanueva A, Pena C, Meinhardt G, Bruix J, Llovet JM. Molecular predictors of prevention of recurrence in HCC with sorafenib as adjuvant treatment and prognostic factors in the phase 3 STORM trial. *Gut* 2019; **68**: 1065-1075 [PMID: 30108162 DOI: 10.1136/gutjnl-2018-316408]
- 58 **Ramirez P**, Sáenz L, Cascales-Campos PA, González Sánchez MR, Llàcer-Millán E, Sánchez-Lorencio MI, Díaz-Rubio E, De La Orden V, Mediero-Valeros B, Navarro JL, Revilla Nuin B, Baroja-Mazo A, Noguera-Velasco JA, Sánchez BF, de la Peña J, Pons-Miñano JA, Sánchez-Bueno F, Robles-Campos R, Parrilla P. Oncological Evaluation by Positron-emission Tomography, Circulating Tumor Cells and Alpha Fetoprotein in Patients With Hepatocellular Carcinoma on the Waiting List for Liver Transplantation. *Transplant Proc* 2016; **48**: 2962-2965 [PMID: 27932119 DOI: 10.1016/j.transproceed.2016.07.035]
- 59 **Mazzaferro V**, Regalia E, Doci R, Andreola S, Pulvirenti A, Bozzetti F, Montalto F, Ammatuna M, Morabito A, Gennari L. Liver transplantation for the treatment of small hepatocellular carcinomas in patients with cirrhosis. *N Engl J Med* 1996; **334**: 693-699 [PMID: 8594428 DOI: 10.1056/NEJM199603143341104]
- 60 **Mazzola A**, Costantino A, Petta S, Bartolotta TV, Raineri M, Sacco R, Brancatelli G, Cammà C, Cabibbo G. Recurrence of hepatocellular carcinoma after liver transplantation: an update. *Future Oncol* 2015; **11**: 2923-2936 [PMID: 26414336 DOI: 10.2217/fon.15.239]
- 61 **Bertuzzo VR**, Cescon M, Ravaioli M, Grazi GL, Ercolani G, Del Gaudio M, Cucchetti A, D'Errico-Grigioni A, Golfieri R, Pinna AD. Analysis of factors affecting recurrence of hepatocellular carcinoma after liver transplantation with a special focus on inflammation markers. *Transplantation* 2011; **91**: 1279-1285 [PMID: 21617590 DOI: 10.1097/TP.0b013e3182187cf0]
- 62 **Xu W**, Cao L, Chen L, Li J, Zhang XF, Qian HH, Kang XY, Zhang Y, Liao J, Shi LH, Yang YF, Wu MC, Yin ZF. Isolation of circulating tumor cells in patients with hepatocellular carcinoma using a novel cell separation strategy. *Clin Cancer Res* 2011; **17**: 3783-3793 [PMID: 21527564 DOI: 10.1158/1078-0432.CCR-10-0498]
- 63 **Chen Z**, Lin X, Chen C, Chen Y, Zhao Q, Wu L, Wang D, Ma Y, Ju W, Chen M, He X. Analysis of preoperative circulating tumor cells for recurrence in patients with hepatocellular carcinoma after liver transplantation. *Ann Transl Med* 2020; **8**: 1067 [PMID: 33145286 DOI: 10.21037/atm-20-2751]



Published by **Baishideng Publishing Group Inc**  
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

**Telephone:** +1-925-3991568

**E-mail:** [bpgoffice@wjgnet.com](mailto:bpgoffice@wjgnet.com)

**Help Desk:** <https://www.f6publishing.com/helpdesk>

<https://www.wjgnet.com>

