

Article

In or Out of the Checklist? DNA Barcoding and Distribution Modelling Unveil a New Species of *Crocidura* Shrew for Italy

Emiliano Mori ^{1,*}, Mattia Brambilla ^{2,3}, Fausto Ramazzotti ⁴, Leonardo Ancillotto ⁵, Giuseppe Mazza ⁶, Danilo Russo ⁷, Giovanni Amori ⁸ and Andrea Galimberti ⁴

¹ Consiglio Nazionale delle Ricerche, Istituto di Ricerca sugli Ecosistemi Terrestri, Via Madonna del Piano 10, 50019 Sesto Fiorentino, Italy

² Fondazione Lombardia per l'Ambiente, Settore Biodiversità e Aree Protette, Largo 10 luglio 1976, 1, I-20822 Seveso, Italy; bramilla.mattia@gmail.com

³ Museo delle Scienze Sezione di Zoologia dei Vertebrati, Corso del Lavoro e della Scienza, 3, I-38122 Trento, Italy

⁴ ZooPlantLab, Department of Biotechnology and Biosciences, University of Milano-Bicocca, P.za della Scienza 2, 20126-I Milano, Italy; fausto.ramazzotti@gmail.com (F.R.); andrea.galimberti@unimib.it (A.G.)

⁵ Museo di Storia Naturale, Università Degli Studi di Firenze, via Romana 17, 50125 Firenze, Italy; leonardo.ancillotto@unina.it

⁶ CREA Research Center for Plant Protection and Certification, Via di Lanciola 12/A, 50125 Firenze, Italy; giuseppe.mazza@crea.gov.it

⁷ Wildlife Research Unit, Dipartimento di Agraria, Università degli Studi Federico II di Napoli, via Università, 100-80055 Portici, Italy; danrusso@unina.it

⁸ Consiglio Nazionale delle Ricerche—Istituto di Ricerca Sugli Ecosistemi Terrestri, Viale dell'Università 32, 00185 Rome, Italy; giovanni.amori@cnr.it

* Correspondence: emiliano.mori@cnr.it; Tel.: +39-333-182-0342

Received: 22 September 2020; Accepted: 30 September 2020; Published: 2 October 2020



Abstract: The genus *Crocidura* (Eulipotyphla, Soricidae) is the most speciose genus amongst mammals, i.e., it includes the highest number of species. Different species are distinguished by skull morphology, which often prevents the identification of individuals in the field and limits research on these species' ecology and biology. We combined species distribution models and molecular analyses to assess the distribution of cryptic *Crocidura* shrews in Italy, confirming the occurrence of the greater white-toothed shrew *Crocidura russula* in the northwest of the country. The molecular identification ascertained the species' presence in two distinct Italian regions. Accordingly, species distribution modelling highlighted the occurrence of areas suitable for *C. russula* in the westernmost part of northern Italy. Our results confirm the role of Italy as a mammal hotspot in the Mediterranean; additionally, they also show the need to include *C. russula* in Italian faunal checklists. To conclude, we highlight the usefulness of combining different approaches to explore the presence of cryptic species outside their known ranges. Since the similar, smaller *C. suaveolens* may be displaced by the larger *C. russula* through competitive exclusion, the latter might be the species actually present where *C. suaveolens* had been reported previously. A comprehensive and detailed survey is therefore required to assess the current distribution of these species.

Keywords: BOLD Systems; *Crocidura russula*; molecular identification; white-toothed shrew

1. Introduction

Field identification at the species level may be problematic and unreliable for rodents and other small mammals, especially when species belonging to the same genus share similar morphology

and overlapping geographical ranges [1–4]. Recent advances in molecular tools have thoroughly improved the current knowledge about mammal taxonomy and redefined species distribution ranges [5–8]. In detail, species identification through DNA barcoding has shed new light on integrating biogeography and taxonomy research [9–11]. Such investigations carried out on species-rich groups of mammals in Italy, e.g., bats and rodents, has recently boosted the numbers of recognized species occurring in the country [12] by describing novel endemic or nearly-endemic taxa (*Sciurus meridionalis*, *Dryomys aspromontis*, *Arvicola italicus*, *Microtus nebrodensis* and *Myotis crypticus* [12]), or showing the presence of cryptic taxa previously unconfirmed for the country, e.g., *Plecotus gaisleri* [13]. Among mammals, Eulipotyphla represents one of the most species-rich groups and, yet, also one of the most underrepresented in studies on small mammals, leading to a lack of knowledge of their basic biology, ecology and distribution [14].

The genus *Crocidura* (Eulipotyphla, Soricidae) includes 196 species of white-toothed shrews distributed in Eurasia and Africa, whose taxonomy has been revised several times and is still debated [14–19]. Species living in Mediterranean countries share a similar morphology and can hardly be distinguished when live-captured (except for *C. leucodon*, which shows a characteristic fur pattern and a smaller body size [20]). Accordingly, most field identifications are carried out through morphology or using known geographical distribution as a criterion, as many *Crocidura* species are endemic to islands (e.g., *C. sicula* in Sicily and the Maltese archipelago, *C. canariensis* in the Canary Islands or *C. zimmermanni* in Crete island) or live in areas with no congeners (e.g., *C. pachyura* in Pantelleria island [19] and *C. suaveolens* on small islands, e.g., Astypalaia, Ischia, Giglio, Capraia and Elba Island [21–23]). Loy et al. [12] reported four species of *Crocidura* as currently present in Italy: the bicolored shrew *C. leucodon*, the lesser white-toothed shrew *C. suaveolens*, the Mediterranean shrew *C. pachyura* and the Sicilian shrew *C. sicula*. Following recent studies, *C. suaveolens* might be, in fact, a species complex and the Italian peninsula may host *C. gueldenstaedtii mimula*, although further detailed molecular analyses are needed to clarify its taxonomic status [12,14].

Another *Crocidura* species occurs in Mediterranean Europe, also in regions that are close to the Italian borders, i.e., the greater white-toothed shrew *C. russula*. The taxonomic overlap of *C. russula* and *C. suaveolens* has been rejected by molecular analyses that showed these are different, divergent species that do not generate hybrids [18,24–26]. After this splitting, *C. russula* remained, for a few decades, in the Italian checklist due to the misclassification of shrews from Sardinia and Pantelleria, and was then removed [21,27,28]. Molecular data showed that the latter populations belong to a different species, *C. pachyura*, also present in eastern Maghreb (Algeria–Tunisia) and on Ibiza island [19]. *Crocidura russula* is the sister taxon of *C. pachyura*, and its known range spans from Morocco to Spain and France, with its easternmost limits corresponding to the southern border between Italy and France [20].

Loy et al. [12] suggested that *C. russula* might occur in Italy, but additional evidence was required to assess its presence at the border with France. Accordingly, a skull from the Nervia river valley (Dolceacqua, province of Imperia, northwest Italy) and other skulls from Lunigiana (provinces of La Spezia and Massa-Carrara, northwest Italy) were considered as potentially belonging to *C. russula* [29,30]. However, because of similar skull sizes compared with *C. suaveolens*, they might belong to the latter species [20]. In sympatry, *C. russula* and *C. suaveolens* (western Europe) are easily identifiable because the former is larger [31]. However, *C. suaveolens* may grow larger in size where *C. russula* is absent (e.g., in eastern Europe) [20]. Thus, given the resemblance between *C. russula* and *C. suaveolens*, as well as the morphological similarity, molecular analyses of field-collected individuals remain the only surefire method for species recognition and distribution assessment, at least in the overlap of their distribution ranges.

Given the suspected presence of *C. russula* in surrounding geographic regions (the Provence-Alpes-Côte d’Azur region), we hypothesized that the species was also present in Italy. Therefore, in this work, we used a DNA barcoding approach to identify western Palearctic shrew species, with special regard to novel samples from the northwest regions of the Italian Peninsula. We then provided the first evidence of the presence of *C. russula* on the Italian territory and reviewed, on such a basis,

the currently known distribution of *Crocidura* shrews occurring in Italy. To conclude, using species distribution modelling, we provide evidence for the potential occurrence of *C. russula* in Italy and for areas of potential overlap with *C. suaveolens*.

2. Materials and Methods

2.1. DNA Barcoding

Four individuals showing morphological features that match those reported for *C. russula* [20] were killed by domestic cats in northwest Italy, where the presence of this species was suspected [21]. Specimens of other Italian species (*C. suaveolens*, *C. leucodon* and *C. pachyura*) were accidentally found dead by the authors or other collaborators (see Acknowledgements: Table 1; Figure 1).

Table 1. Italian *Crocidura* specimens analysed in our work, including those resembling *C. russula*. Specimens of *C. suaveolens*, *C. leucodon* and *C. pachyura* were included for comparison with *C. russula*. Specimen details and GenBank accession numbers are reported for each sample. a.n., accession number.

Taxon	Voucher Sample	COI a.n.	Sampling Locality	Coordinates
<i>C. cfr. russula</i>	MIB:ZPL:08649	LR877375	Levaldigi (CN), Piedmont	44.554216° N–7.633327° E
<i>C. cfr. russula</i>	MIB:ZPL:08650	LR877376	Dolceacqua (IM), Liguria	43.860976° N–7.620309° E
<i>C. cfr. russula</i>	MIB:ZPL:08651	LR877377	Rocchetta Nervina (IM), Liguria	43.890148° N–7.598244° E
<i>C. cfr. russula</i>	MIB:ZPL:08652	LR877378	Rocchetta Nervina (IM), Liguria	43.890927° N–7.597591° E
<i>C. suaveolens</i>	MIB:ZPL:08653	LR877379	Ghedi (BS), Lombardy	45.404473° N–10.267402° E
<i>C. leucodon</i>	MIB:ZPL:08654	LR877380	Oasi “Le Bine” (MN), Lombardy	45.146581° N–10.445089° E
<i>C. pachyura</i>	MIB:ZPL:08655	LR877381	Thiesi (SS), Sardinia	40.526080° N–8.663751° E

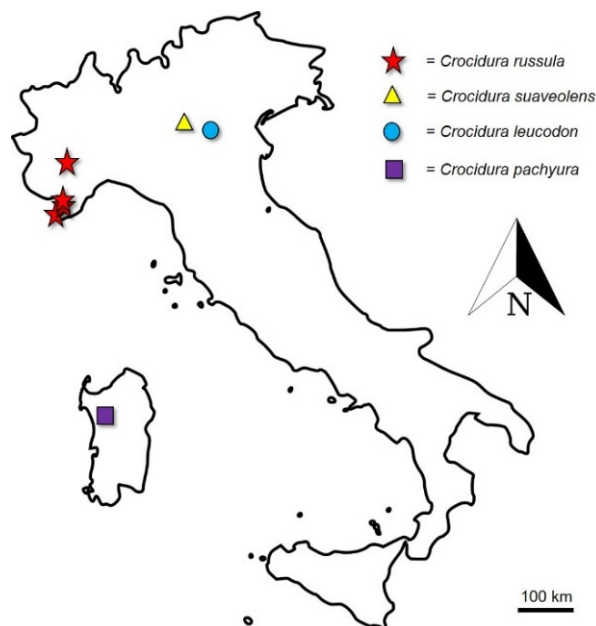


Figure 1. Locations of origin of *Crocidura* samples used in our study (Italy).

Total DNA was extracted from the *Crocidura* tail samples collected in Italy (Table 1) by using the DNeasy Blood & Tissue Kit (Qiagen, Milan, Italy) following the manufacturer’s instructions. Purified DNA concentrations and sample quality were estimated fluorometrically with a NanoDrop™1000 Spectrophotometer (Thermo Fisher Scientific, Waltham, MA, USA). To produce DNA barcoding sequences for the morphologically identified *Crocidura* species (i.e., *C. suaveolens*, *C. leucodon* and *C. pachyura*) and identify the four presumed *C. russula* samples, the standard DNA barcode 5’-end region of the mitochondrial COI gene (658 bp) was amplified and sequenced using the universal primers LCO1490 and HCO2918 [32], with the thermal profile described by [33] and chemical reaction

parameters in [34]. After sequencing, primer-nucleotide-sequence trimming, and a sequence-quality check, the presence of stop codons was verified through the online tool EMBOSS Transeq (http://www.ebi.ac.uk/Tools/st/emboss_transeq/). This analysis was conducted to exclude the amplification of COI pseudogenes that could affect the identification results or the discovery of divergent intraspecific lineages [10]. Sequence data were submitted to the European Bioinformatics Institute of the European Molecular Biology Laboratory (EMBL-EBI) and assigned to the accession numbers provided in Table 1.

To confirm the taxonomic assignment of the presumed *Crocidura russula* samples from northwest Italy, the obtained sequences were first queried against the BOLD Systems (IDS tool) database [35]. Second, to assess the genetic divergence among these specimens and other Italian and western Palearctic *Crocidura* species, we assembled a multiple alignment for the COI fragment including the sequences obtained in this study and the available sequences from BOLD (Table 1). Multiple sequence alignments were produced using MAFFT online (<https://mafft.cbrc.jp/alignment/server> [36]), set to default parameters. Due to the different lengths of the available sequences, the alignment was trimmed to the same final lengths. Genetic K2P distances were calculated using MEGA X [37].

2.2. Species Distribution Modelling

We collected occurrence records (with photos validated through an expert-based approach) for the period 2007–2020 from multiple data sources: iNaturalist (www.inaturalist.org: downloaded on 10 August 2020), Ornitho (www.ornitho.it: downloaded on 19 August 2020), the Facebook group on Italian Mammals (<https://www.facebook.com/groups/mammiferi/>, updated up to 12 August 2020) and the authors' personal records. Validated data from iNaturalist are also included in the GBIF (Global Biodiversity Information Facility) platform, the same used by the International Union for Conservation of Nature (IUCN) to assess the range of animal species. Several of them were also included in previous work on the genetics/skull morphology of Italian shrews [12,21,26,29–31] or stored in Italian museums (Museo di Storia Naturale di Firenze “La Specola”; Museo di Storia Regionale della Maremma). Several others were directly captured, and the body measurements appeared to be within the variability of just one of the two species in sympatric areas [20]. We kept, for analyses, occurrences from areas including sympatric populations of both *C. russula* and *C. suaveolens* only if the identification was deemed reliable [31], as well as occurrences from areas where previous genetic work confirmed the presence of only one of these species [18,19,24,26,38]. Occurrences that could not be unambiguously assigned to one species were excluded. From the resulting dataset, we removed all records with a spatial accuracy coarser than 1 km.

We obtained 117 reliable records of *C. russula* and 138 of *C. suaveolens*. We modelled the potential distribution of the two *Crocidura* species as a function of climate, topography and land-cover within 1 km × 1 km cells of a grid superimposed on central and southern Europe. We selected bioclimatic variables, downloaded from the CHELSA database [39], at a 30 arc-second resolution. Topographic data were derived from the European Digital Elevation Model with a 25 m resolution (EU-DEM v1.0, made available by the European Environment Agency). We calculated the slope (average slope over the cell in degrees) and solar radiation (global radiation calculated for 21 March, 21 June and 22 September, averaged over time and cell) in GRASS [40]. Land cover variables were expressed as the respective proportional cover within the cell and derived from the Corine Land Cover [41]. Some land use variables, which were likely to have similar importance for the target species based on their ecology, were merged together to reduce the number of predictors and increase their potential importance. This process led to the following categories: urban (all artificial surfaces except for green urban areas), complex crops (closely associated crops of different types or crops interspersed with natural vegetation), inland water (watercourses and waterbodies), salt water (coastal lagoons, estuaries and seas), bare ground or sparse vegetation (all habitats with no or sparse vegetation) (see Table S1 in Supplementary Material for details). Our occurrence data were thus opportunistic and non-systematic. Therefore, we adopted a maximum entropy approach, as implemented in Maxent, as this method outperforms

others when non-standardized data are used [42,43]. Maxent works with an occurrence-background framework, and it is essential to place background points, to mirror the environmental conditions that had been sampled with data collection (i.e., to avoid the inclusion into the background of conditions where the target species had not been found just because such conditions were not sampled) [44]. In our study, we did not have any information about the sampling efforts, so we restricted the background to the areas containing occurrence records by creating a 20 km buffer around the occurrence records, within which we randomly placed 30,000 points. These resulted in 28,452 actual background points after overlap with environmental layers. To build models as robust as possible, for each species, we subdivided the occurrence data into four spatially independent partitions using the *checkerboard 2* method in ENMeval [45]. We thus used the subset containing data from three partitions to train the model and the fourth to test it on spatially independent data. We used only linear and quadratic features and selected the regularization multiplier on the basis of the Akaike's Information Criterion, corrected for a small sample size (AIC_c). We did this first for a "basic" full model, which was then used for the selection between correlated variables: among pairs of correlated predictors ($r > 0.7$), we kept only the one included in the most supported model based on AUC (Area Under the Curve of the Receiver Operating Characteristic) values, using SDMtune [46]. We then removed all variables with $\Lambda = 0$, suggesting no noticeable effect on species occurrence. Finally, we performed model tuning, selecting—according to the AIC_c value—the regularization multiplier, the features to be used (linear and/or quadratic), the number of iterations and the inclusion/exclusion of environmental variables (starting from the removal of the variable with the lowest value of permutation importance [47]). The values selected by the tuning procedure were the following ones: *C. russula*: regularization multiplier: 1, features: linear and quadratic, number of iterations: 540; *C. suaveolens*: regularization multiplier: 0.5, features: linear and quadratic, number of iterations: 360. We then calculated, for each species, the AUC and TSS (True Skill Statistic) values over the training and test datasets, and the omission rates at selected thresholds (10th percentile and minimum training presence) for the test data to ensure that models were not subject to overfitting.

3. Results

3.1. DNA Barcoding Identification

The mitochondrial COI-5' end region was successfully sequenced for all the *Crocidura* samples collected in Italy, and no stop codons were found. The four presumed *C. russula* shared the same haplotype, and the BOLD-IDS search returned a 100% maximum identity match with 13 German reference specimens and one Belgian reference specimen of *C. russula*. The identity of Italian specimens of *C. suaveolens* and *C. leucodon* was also confirmed, whereas for *C. pachyura*, the DNA barcode sequence produced in this study was the first one ever published for this species.

The multiple alignments encompassing all the publicly available DNA barcode sequences of western Palearctic *Crocidura* species contained neither stop codons nor indels, and after trimming, these were 511 bp long. The K2P-distance values confirmed the marked genetic divergence of *C. russula* from the other *Crocidura* species, supporting the unambiguous identification of the northwest Italian samples (Table 2).

Table 2. Genetic K2P distance divergence values among *Crocidura* species.

Taxon I	Taxon II	Distance	Standard Error
<i>C. cfr. russula</i> (Italy)	<i>C. russula</i>	0.0000	0.0000
<i>C. cfr. russula</i> (Italy)	<i>C. suaveolens</i>	0.1592	0.0162
<i>C. cfr. russula</i> (Italy)	<i>C. leucodon</i>	0.1823	0.0205
<i>C. cfr. russula</i> (Italy)	<i>C. pachyura</i>	0.0892	0.0129
<i>C. cfr. russula</i> (Italy)	<i>C. olivieri</i>	0.1472	0.0168
<i>C. cfr. russula</i> (Italy)	<i>C. viaria</i>	0.1541	0.0177
<i>C. russula</i>	<i>C. suaveolens</i>	0.1592	0.0162
<i>C. russula</i>	<i>C. leucodon</i>	0.1823	0.0205
<i>C. russula</i>	<i>C. pachyura</i>	0.0892	0.0129
<i>C. russula</i>	<i>C. olivieri</i>	0.1472	0.0168
<i>C. russula</i>	<i>C. viaria</i>	0.1541	0.0177
<i>C. suaveolens</i>	<i>C. leucodon</i>	0.1376	0.0151
<i>C. suaveolens</i>	<i>C. pachyura</i>	0.1570	0.0160
<i>C. suaveolens</i>	<i>C. olivieri</i>	0.1466	0.0148
<i>C. suaveolens</i>	<i>C. viaria</i>	0.1364	0.0147
<i>C. leucodon</i>	<i>C. pachyura</i>	0.1632	0.0189
<i>C. leucodon</i>	<i>C. olivieri</i>	0.1676	0.0183
<i>C. leucodon</i>	<i>C. viaria</i>	0.1683	0.0186
<i>C. pachyura</i>	<i>C. olivieri</i>	0.1581	0.0172
<i>C. pachyura</i>	<i>C. viaria</i>	0.1607	0.0178
<i>C. olivieri</i>	<i>C. viaria</i>	0.0302	0.0048

3.2. Species Distribution Modelling

For both *C. russula* and *C. suaveolens*, the distribution models appeared sufficiently robust. The model's performances were similar for the training and test datasets for *C. russula* (TSS of 0.55 vs. 0.57, and AUC of 0.84 vs. 0.83, for the training and test datasets, respectively; $N = 88$ and 29), as well as for *C. suaveolens* (TSS of 0.52 vs. 0.5, and AUC of 0.81 vs. 0.79, for the training and test datasets, respectively; $N = 101$ and 37). The omission rates confirmed the lack of overfitting, these being very close (or slightly lower than, in the case of 10th percentile for *C. russula*) to the expected values.

The environmental suitability for both species depended on several variables (Table 3) and was positively associated with medium-high values of urban cover and negatively so with arable land. The most important factors for *C. suaveolens* were BIO4 (temperature seasonality) and BIO6 (the minimum temperature of the coldest month), which revealed a potential association of the species with climates characterized by high seasonality and relatively mild winters. The suitability for *C. russula* was affected first by the urban cover and then by BIO7 (the temperature annual range) and BIO10 (the mean temperature of the warmest quarter), suggesting the avoidance of climates with extreme temperature differences between seasons and of temperature extremes during the summer. The suitability for *C. suaveolens* was favoured by partial (quadratic) cover of broadleaved forest and wetlands, whereas *C. russula* likely preferred vineyards and orchards, avoiding bare ground or sparse vegetation and a high cover of broadleaved forest.

The potential distribution of habitats suitable for *C. russula* (Figure 2) highlighted the occurrence of suitable environments in northwest Italian regions. These areas perfectly overlapped with the records of the species and also included some nearby areas (see the inset in Figure 2). Italy showed a high suitability for *C. suaveolens* (Figure 3). All the confirmed occurrences were included in areas suitable for both species in Italy. The spatial variation of the environmental suitability for both species is shown in Figures S1 and S2, in the Supplementary Material.

Table 3. Variables included in the distribution models, relative permutation importance (and SD calculated over 10 permutations) and description of the variable effects. The quadratic effect always refers to higher suitability at intermediate values.

Variable	Permutation Importance	SD	Effect
<i>C. russula</i>			
urban	29	0.01	positive/quadratic
temperature annual range (BIO7)	16.8	0.02	negative/quadratic
mean temp. of warmest quarter (BIO10)	15.3	0.01	quadratic
broad-leaved forest	12.7	0.02	negative/quadratic
precipitation of warmest quarter (BIO18)	6.6	0.01	negative
non-irrigated arable land	6.1	0.01	negative
bare ground and sparse vegetation	4.0	0.01	negative
sclerophyllous vegetation	2.7	0.01	positive/quadratic
complex crops	2.7	0.00	negative
vineyards	2.5	0.00	positive
fruit trees and berry plantations	1.8	0.00	positive
<i>C. suaveolens</i>			
temperature seasonality (BIO4)	53.7	0.02	positive
min. temp. of the coldest month (BIO6)	21.6	0.01	positive
urban	7.5	0.01	positive/quadratic
broad-leaved forest	6.9	0.01	quadratic
mixed forest	5.4	0.02	quadratic/negative
slope	3.9	0.01	positive
inland marshes	0.9	0.00	quadratic
non-irrigated arable land	0.0	0.00	negative

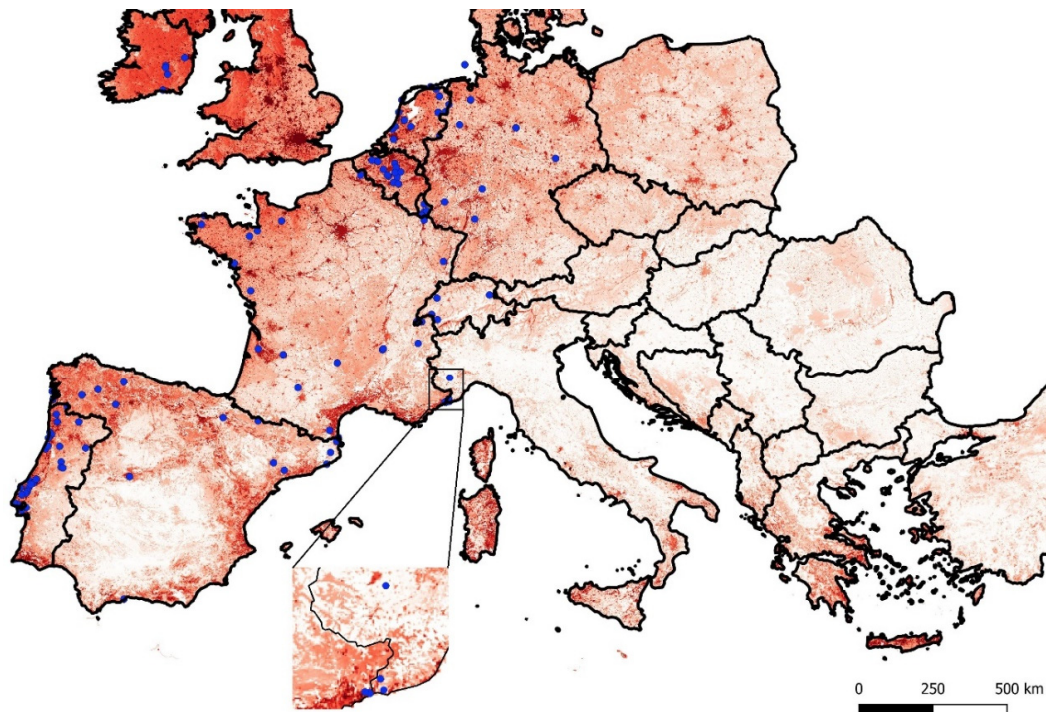


Figure 2. Potential distribution of *C. russula* with the occurrence records shown in light blue. White colour denotes unsuitable areas, whereas suitable areas are shown by a red gradient from poorly suitable sites in light red to highly suitable ones in dark red. The inset shows the habitat suitability in detail for eastern France and western Italy.

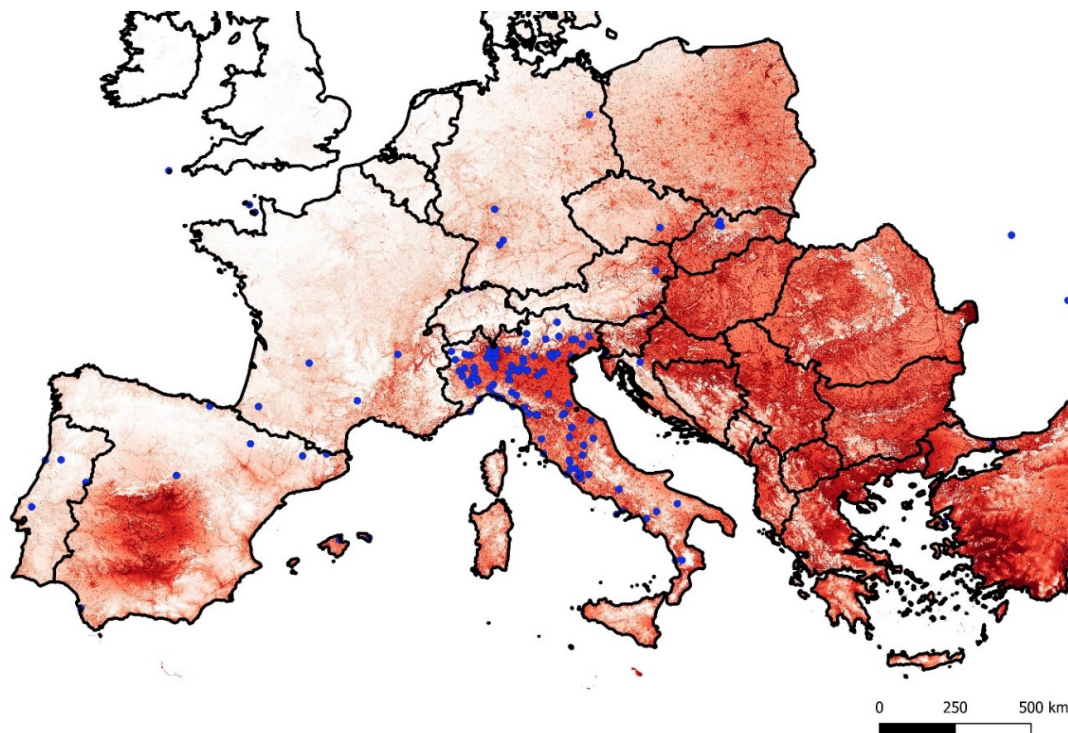


Figure 3. Potential distribution of *C. suaveolens* with the occurrence records shown in dark blue. White colour denotes unsuitable areas, whereas suitable areas are shown by a red gradient from poorly suitable sites in light red to highly suitable ones in dark red.

4. Discussion

In our work, we provided the first genetic evidence of the presence *C. russula* in Italy, near the border with France (Nervia river valley) and in western Piedmont (province of Cuneo), thus confirming previous suggestions by other authors based on skull morphology and owl pellet analyses [21,29,30]. Old distributional data, on which basis *C. russula* was thought to be present across the whole country [15,48], are, in fact, misleading because, at that time, the taxon also included *C. suaveolens* and *C. pachyura* [49], because of the high overlap of most of the morphological features and measurements among these taxa (Table 4: [20,21]). Further studies should be conducted to define the current range of *C. russula* in Italy, at least in the northwestern regions (Piedmont, Liguria and Lunigiana in northwestern Tuscany [21]).

Table 4. Body measurements of *Crocidura* species occurring in Italy and neighbouring countries [20,21].

Species	Body-Tail Length	Tail Length	Hind Foot	Condiloincise Length	Weight
<i>Crocidura pachyura</i>	62–78 mm	30–40 mm	11–13 mm	18–22 mm	8–16 g
<i>Crocidura russula</i>	44–86 mm	24–47 mm	10–14 mm	18–20 mm	5–16 g
<i>Crocidura sicula</i>	50–77 mm	28–42 mm	11–14 mm	18–20 mm	4–10 g
<i>Crocidura suaveolens</i>	50–92 mm	25–58 mm	10–16 mm	16–21 mm	3–16 g
<i>Crocidura leucodon</i>	50–90 mm	27–46 mm	11–15 mm	17–24 mm	7–20 g

The species–environment relationships suggested both similarities and divergences in the ecology of the two shrew species we examined. The suitability for both species increases almost linearly with the urban cover (with a decrease in suitability for complete urban cover) and decreases with arable land. Habitat suitability for *C. suaveolens* was positively influenced by moderate cover of broadleaved forest and wetlands, in agreement with other authors [31], and was especially associated with areas

characterized by high seasonality in temperature and not reaching excessively low temperatures in winters. The suitability for *C. russula*, instead, is negatively affected by extreme temperature differences between seasons and temperature extremes in summer. Such differences between species suggest that *C. suaveolens* is more specialized for Mediterranean and continental, relatively warm, climates than *C. russula*, whereas the latter could be adapted to more oceanic climates.

Assessing the presence of small-sized, elusive mammals may be a difficult task, and cryptic species may remain long undetected [6]. Distinguishing the occurrences of *C. russula* and *C. suaveolens* is challenging because skulls are needed to fulfil the task. Apart from the skull analysis of sacrificed specimens, examining skulls from owl pellets may provide an answer [31], but in those cases, the exact location is uncertain due to the predator's mobility (up to several kilometres between roosts/nests and feeding sites [50–52]). In this work, we provided evidence that DNA barcoding is a reliable litmus test for identifying *Crociodura* shrews where they are sympatric or close by [10].

The potential distributions depicted by our models matched fairly well with the known patterns of occurrence. Very few occurrence records of *C. suaveolens* are available for eastern Europe, whereas most of this area was depicted by the model as highly suitable for the species, coherently with the known distribution [14,38]. The model for *C. russula* identified a higher suitability for western Europe, in agreement with the known occurrence pattern [53]. The British Isles were also identified as potentially suitable for the species but were not included within the species range according to IUCN maps, as the occurrence of *C. russula* in Ireland was confirmed after the IUCN assessment [54,55]. Interestingly, the model detected a continuous high suitability for *C. russula* from southeast France to northwest Italy. This finding further reinforces the point we make that some populations of this species occur in the northwest corner of Italy: suitable habitats are predicted to occur in sites with genetically confirmed occurrence records and in nearby areas, where new investigations might lead to the discovery of other populations.

Not only do our results provide strong evidence of the occurrence of a to-date-overlooked species in Italy but also highlight the potential of applying an integrated approach in the study of cryptic species for shedding light on their distribution and ecology, both fundamental tools for local and global conservation actions. Finally, our study further emphasizes the importance of the Mediterranean basin and of Italy in particular as biodiversity hotspots. Accordingly, several other vertebrate taxa show similar high speciation rates in the region, occur in sympatry or in close proximity in Italy [56–59], and thus deserve further research efforts in the future to properly assess their distribution and conservation status.

Supplementary Materials: The following are available online at <http://www.mdpi.com/1424-2818/12/10/380/s1>. Table S1. Full list of variables used for distribution modelling. Figure S1. Environmental suitability for *C. russula* (georeferenced JPEG, coordinate system EPSG:3035—projected coordinate system for Europe). Figure S2. Environmental suitability for *C. suaveolens* (georeferenced JPEG, coordinate system EPSG:3035—projected coordinate system for Europe).

Author Contributions: Conceptualization, E.M., L.A. and A.G.; methodology, A.G., M.B. and F.R.; software, M.B. and A.G.; validation, D.R., A.G. and G.A.; formal analysis, M.B. and A.G.; investigation, E.M., G.M. and L.A.; resources, A.G.; data curation, E.M. and G.M.; writing—original draft preparation, E.M., M.B., L.A. and A.G.; writing—review and editing, G.M., D.R. and G.A. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Acknowledgments: We thank Luca Mazzini, Clio Tracanzan, Giuliano Milana and Francesco Cecere for their help in sampling and data collection, and Gianpiero Calvi for advising on the distribution models. Paolo Agnelli and Andrea Sforzi allowed us to check for museum specimens, respectively, in Museo di Storia Naturale di Firenze “La Specola” and Museo di Storia Regionale della Maremma. Two anonymous reviewers provided us with useful comments. Vasco Sfondrini kindly revised the first version of our manuscript and improved the English grammar and syntax.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Castiglia, R.; Annesi, F.; Aloise, G.; Amori, G. Systematics of the *Microtus savii* complex (Rodentia, Cricetidae) via mitochondrial DNA analyses: Paraphyly and pattern of sex chromosome evolution. *Mol. Phylogenet. Evol.* **2008**, *46*, 1157–1164. [[CrossRef](#)] [[PubMed](#)]
- Abiadh, A.; Chetoui, M.; Lamine-Cheniti, T.; Capanna, E.; Colangelo, P. Molecular phylogenetics of the genus *Gerbillus* (Rodentia, Gerbillinae): Implications for systematics, taxonomy and chromosomal evolution. *Mol. Phylogenet. Evol.* **2010**, *56*, 513–518. [[CrossRef](#)] [[PubMed](#)]
- Ancillotto, L.; Mori, E.; Sozio, G.; Solano, E.; Bertolino, S.; Russo, D. A novel approach to field identification of cryptic *Apodemus* wood mice: Calls differ more than morphology. *Mamm. Rev.* **2016**, *47*, 6–10. [[CrossRef](#)]
- Halley, D.J.; Saveljev, A.P.; Rosell, F. Population and distribution of beavers *Castor fiber* and *Castor canadensis* in Eurasia. *Mamm. Rev.* **2020**. [[CrossRef](#)]
- Castiglia, R.; Aloise, G.; Amori, G.; Annesi, F.; Bertolino, S.; Capizzi, D.; Mori, E.; Colangelo, P. The Italian peninsula hosts a divergent mtDNA lineage of the water vole, *Arvicola amphibius* s.l., including fossorial and aquatic ecotypes. *Hystrix* **2016**, *27*, 99–103.
- Ancillotto, L.; Mori, E.; Bosso, L.; Agnelli, P.; Russo, D. The Balkan long-eared bat (*Plecotus kolombatovici*) occurs in Italy—First confirmed record and potential distribution. *Mamm. Biol.* **2019**, *96*, 61–67. [[CrossRef](#)]
- Mori, E.; Nerva, L.; Lovari, S. Reclassification of the serows and the gorals: The end of a never-ending story? *Mamm. Rev.* **2019**, *49*, 256–262. [[CrossRef](#)]
- Li, G.; Sun, N.; Swa, K.; Zhang, M.; Lwin, Y.H.; Quan, R.C. Phylogenetic reassessment of gorals with new evidence from northern Myanmar reveals five distinct species. *Mamm. Rev.* **2020**. [[CrossRef](#)]
- Galimberti, A.; Spada, M.; Russo, D.; Mucedda, M.; Agnelli, P.; Crottini, A.; Ferri, E.; Martinoli, A.; Casiraghi, M. Integrated operational taxonomic units (IOTUs) in echolocating bats: A bridge between molecular and traditional taxonomy. *PLoS ONE* **2012**, *7*, e40122. [[CrossRef](#)]
- Galimberti, A.; Sandionigi, A.; Bruno, A.; Bellati, A.; Casiraghi, M. DNA barcoding in mammals: What's new and where next? *Hystrix* **2015**, *26*, 13–24.
- Sales, N.G.; Kaizer, M.D.C.; Coscia, I.; Perkins, J.C.; Highlands, A.; Boubli, J.P.; Magnusson, W.E.; Da Silva, M.N.F.; Benvenuto, C.; Mcdevitt, A.D. Assessing the potential of environmental DNA metabarcoding for monitoring Neotropical mammals: A case study in the Amazon and Atlantic Forest, Brazil. *Mamm. Rev.* **2020**. [[CrossRef](#)]
- Loy, A.; Aloise, G.; Ancillotto, L.; Angelici, F.M.; Bertolino, S.; Capizzi, D.; Castiglia, R.; Colangelo, P.; Contoli, L.; Cozzi, B.; et al. Mammals of Italy: An annotated checklist. *Hystrix* **2019**, *30*, 87–106.
- Ancillotto, L.; Bosso, L.; Smeraldo, S.; Mori, E.; Mazza, G.; Herkt, M.; Galimberti, A.; Ramazzotti, F.; Russo, D. An African bat in Europe, *Plecotus gaisleri*: Biogeographic and ecological insights from molecular taxonomy and species distribution models. *Ecol. Evol.* **2020**. [[CrossRef](#)] [[PubMed](#)]
- Wilson, D.E.; Mittermeier, R.A. *Handbook of the Mammals of the World (Vol. 8), Insectivores, Sloths and Colugos*; Lynx Editions: Barcelona, Spain, 2018.
- Miller, G.S. *Catalogue of the Mammals of Western Europe*; Trustees of British Museum Editions: London, UK, 1912.
- Meylan, A.; Hausser, J. Position cytotaxonomique de quelques musaraignes du genre *Crocidura* au Tessin (Mammalia, Insectivora). *Rev. Suisse Zool.* **1974**, *81*, 701–710. [[CrossRef](#)]
- Rzebik-Kowalska, B. Studies on the genus *Crocidura* (Insectivora, Mammalia) in Algeria. *Acta Zool. Cracovia* **1988**, *31*, 167–192.
- Cosson, J.F.; Hutterer, R.; Libois, R.; Sarà, M.; Taberlet, P.; Vogel, P. Phylogeographic footprints of the Strait of Gibraltar and Quaternary climatic fluctuations in the Western Mediterranean: A case study with the greater white-toothed shrew *Crocidura russula* (Mammalia: Soricidae). *Mol. Ecol.* **2005**, *14*, 1151–1162. [[CrossRef](#)]
- Nicolas, V.; Hamani, A.; Amrouche, L.; Bensidhoum, M.; Boukhemza, M.; Doumandji, S.; Denys, C. First molecular evidence for the presence of *Crocidura pachyura* (Mammalia, Soricidae) in Kabylie (Algeria). *Mammalia* **2014**, *78*, 245–249. [[CrossRef](#)]
- Aulagnier, S.; Haffner, P.; Mitchell-Jones, A.J.; Moutou, F.; Zima, J. *Guide des Mammifères d'Europe, d'Afrique du Nord et du Moyen-Orient*; Delachaux & Niestlé SA Editions: Paris, France, 2010.
- Amori, G.; Contoli, L.; Nappi, A. *Mammalia II: Erinaceomorpha, Soricomorpha, Lagomorpha, Rodentia*; Il Sole 24 Ore; Edagricole, Calderini Editions: Bologna, Italy, 2008.

22. Amori, G.; Rizzo Pinna, V.; Sammuri, G.; Luiselli, L. Diversity of small mammal communities of the Tuscan Archipelago: Testing the effects of island size, distance from mainland and human density. *Folia Zool.* **2015**, *64*, 161–166. [[CrossRef](#)]
23. Angelici, F.M.; Cattaneo, C.; Grano, M.; Nappi, A. About the presence of *Crocidura suaveolens* group (Soricomorpha, Soricidae) on Astipalaia Island (Dodecanese, Greece). *Nat. Hist. Sci.* **2018**, *5*, 3–6. [[CrossRef](#)]
24. Catalan, J.; Poitevin, F. Les Crocidures du midi de la France: Leurs caractéristiques génétiques et morphologiques: La place de populations corses. *C.R. Acad. Sci. Paris (III)* **1981**, *292*, 1017–1020.
25. Catzeflis, F. Relations génétiques entre trois espèces du genre *Crocidura* (Soricidae, Mammalia) en Europe. *Mammalia* **1983**, *47*, 387–400. [[CrossRef](#)]
26. Castiglia, R.; Annesi, F.; Amori, G.; Solano, E.; Aloise, G. The phylogeography of *Crocidura suaveolens* from Southern Italy reveals the absence of an endemic lineage and supports a Trans-Adriatic connection with the Balkanic refugium. *Hystrix* **2017**, *28*, 104–106.
27. Amori, G.; Angelici, F.M.; Boitani, L. Mammals of Italy: A revised checklist of species and subspecies. *Senckenberg. Biol.* **1999**, *79*, 271–286.
28. Gippoliti, S. Checklist delle specie di mammiferi italiani (esclusi Mysticeti e Odontoceti): Contributo per la conservazione della biodiversità. *Boll. Mus. Civ. Stor. Nat. Verona* **2013**, *37*, 7–28.
29. Sigillo, P.M. Micromammiferi Nelle Borre di Barbagianni (*Tyto alba*) Nella Liguria Occidentale. Master's Thesis, Biological Sciences, University of Rome "La Sapienza", Rome, Italy, 1983.
30. Rosi, R.E. Analisi e confronto fra alcuni caratteri morfologici e morfometrici di *Crocidura* sp. della Lunigiana e di *Crocidura leucodon* (Hermann, 1780) e *Crocidura russula* (Hermann, 1780) della Francia. *Quad. Stor. Nat. Cent. "Malaspina"* **2006**, *1*, 35–36.
31. Biedma, L.; Román, J.; Godoy, J.A.; Calzada, J. Using owl pellets to infer habitat associations and clarify the regional distribution of a cryptic shrew. *J. Zool.* **2019**, *308*, 139–148. [[CrossRef](#)]
32. Folmer, O.; Black, M.; Hoeh, W.; Lutz, R.; Vrijenhoek, R. DNA primers for amplification of mitochondrial cytochrome-c oxidase subunit I from diverse metazoan invertebrates. *Mol. Mar. Biol. Biotechnol.* **1994**, *3*, 294–299.
33. Bellati, A.; Tiberti, R.; Cocca, W.; Galimberti, A.; Casiraghi, M.; Bogliani, G.; Galeotti, P. A dark shell hiding great variability: A molecular insight into the evolution and conservation of melanic *Daphnia* populations in the Alps. *Zool. J. Linn. Soc.* **2014**, *171*, 697–715. [[CrossRef](#)]
34. Mazzamuto, M.V.; Galimberti, A.; Cremonesi, G.; Pisanu, B.; Chapuis, J.L.; Stuyck, J.; Amori, G.; Su, H.J.; Aloise, G.; Preatoni, D.G.; et al. Preventing species invasion: A role for integrative taxonomy? *Integr. Zool.* **2016**, *11*, 214–228. [[CrossRef](#)]
35. Ratnasingham, S.; Hebert, P.D. BOLD: The Barcode of Life Data System (<http://www.barcodinglife.org>). *Mol. Ecol. Notes* **2007**, *7*, 355–364. [[CrossRef](#)]
36. Katoh, K.; Standley, D.M. MAFFT multiple sequence alignment software version 7: Improvements in performance and usability. *Mol. Biol. Evol.* **2013**, *30*, 772–780. [[CrossRef](#)] [[PubMed](#)]
37. Kumar, S.; Stecher, G.; Li, M.; Nnyaz, C.; Tamura, K. MEGA X: Molecular evolutionary genetics analysis across computing platforms. *Mol. Biol. Evol.* **2018**, *35*, 1547–1549. [[CrossRef](#)] [[PubMed](#)]
38. Vogel, P.; Cosson, J.F.; Jurado, L.F.L. Taxonomic status and origin of the shrews (Soricidae) from the Canary islands inferred from a mtDNA comparison with the European *Crocidura* species. *Mol. Phylogenet. Evol.* **2003**, *27*, 271–282. [[CrossRef](#)]
39. Karger, D.N.; Conrad, O.; Böhrner, J.; Kawohl, T.; Kreft, H.; Soria-Auza, R.W.; Zimmermann, N.E.; Linder, H.P.; Kessler, M. Climatologies at high resolution for the earth's land surface areas. *Sci. Data* **2017**, *4*, 170122. [[CrossRef](#)]
40. Neteler, M.; Bowman, M.H.; Landa, M.; Metz, M. GRASS GIS: A multi-purpose open source GIS. *Environ. Model. Softw.* **2012**, *31*, 124–130. [[CrossRef](#)]
41. European Environment Agency. Corine Land Cover 2012. Available online: <https://www.eea.europa.eu/data-and-maps/data/external/corine-land-cover-2012> (accessed on 17 August 2020).
42. Elith, J.; Phillips, S.J.; Hastie, T.; Dudík, M.; Chee, Y.E.; Yates, C.J. A statistical explanation of MaxEnt for ecologists. *Divers. Distrib.* **2011**, *17*, 43–57. [[CrossRef](#)]
43. Merow, C.; Smith, M.J.; Silander, J.A. A practical guide to MaxEnt for modeling species' distributions: What it does, and why inputs and settings matter. *Ecography* **2013**, *36*, 1058–1069. [[CrossRef](#)]

44. Brambilla, M.; Resano-Mayor, J.; Arlettaz, R.; Bettega, C.; Binggeli, A.; Bogliani, G.; Braunisch, V.; Celada, C.; Chamberlain, D.; Chiffard Carricaburu, J.; et al. Potential distribution of a climate sensitive species, the White-winged Snowfinch *Montifringilla nivalis* in Europe. *Bird Conserv. Int.* **2020**. [CrossRef]
45. Muscarella, R.; Galante, P.J.; Soley-Guardia, M.; Boria, R.A.; Kass, J.M.; Uriarte, M.; Anderson, R.P. ENMeval: An R package for conducting spatially independent evaluations and estimating optimal model complexity for Maxent ecological niche models. *Methods Ecol. Evol.* **2014**, *5*, 1198–1205. [CrossRef]
46. Vignali, S.; Barras, A.; Braunisch, V. SDMtune: Species Distribution Model Selection. R Package Version 1.1.1. Available online: <https://github.com/ConsBiol-unibern/SDMtune> (accessed on 17 August 2020).
47. Phillips, S.J.; Anderson, R.P.; Schapire, R.E. Maximum entropy modeling of species geographic distributions. *Ecol. Model.* **2006**, *190*, 231–259. [CrossRef]
48. Toschi, A. Insectivora Gray, 1827. In *Mammalia. Generalità, Insectivora, Chitopteera. Collana "Fauna d'Italia"*; Toschi, A., Lanza, B., Eds.; Edizioni Calderini: Bologna, Italy, 1959; Volume IV, pp. 65–186.
49. Richter, H. Zur Taxonomie und Verbreitung der palaearktischen Crociduren (*Mammalia* Insectivora, Soricidae). *Zool. Abh. Staatl. Mus. Tierk. Dresd.* **1970**, *31*, 294–304.
50. Aebischer, A.; Nyffeler, P.; Arlettaz, R. Wide-range dispersal in juvenile Eagle Owls (*Bubo bubo*) across the European Alps calls for transnational conservation programmes. *J. Ornithol.* **2010**, *151*, 1–9. [CrossRef]
51. Massa, C.; Gabelli, F.M.; Cueto, G.R. Using GPS to determine movement patterns and foraging habitat selection of the common barn owl (*Tyto alba*). *Hornero* **2015**, *30*, 7–12.
52. Mori, E.; Mazzetto, F.; Menchetti, M.; Bodino, N.; Grasso, E.; Sposimo, P. Feeding ecology of the scops owl, *Otus scops* (Aves: Strigiformes), in the island of Pianosa (Tuscan Archipelago, Central Italy) outside the breeding period. *Ital. J. Zool.* **2016**, *83*, 417–422. [CrossRef]
53. Jaquierey, J.; Guèlat, J.; Broquet, T.; Berset-Brandli, L.; Pellegrini, E.; Moresi, R.; Hirzel, A.H.; Perrin, N. Habitat-quality effects on metapopulation dynamics in greater white-toothed shrews, *Crocidura russula*. *Ecology* **2008**, *89*, 2777–2785. [CrossRef]
54. Aulagnier, S.; Hutterer, R.; Amori, G.; Kryštufek, B.; Yigit, N.; Mitsain, G.; Palomo, L.J. *Crocidura russula*. *The IUCN Red List of Threatened Species 2016: E.T29652A115169607*; IUCN Editors: Gland, Switzerland, 2016.
55. Gagan, L.M.; Cornette, R.; Yearsley, J.M.; Montgomery, W.I.; Paupèrio, J.; Alves, P.C.; Butler, F.; Pascal, M.; Tresset, A.; Herrel, A.; et al. Molecular and morphological insights into the origin of the invasive greater white-toothed shrew (*Crocidura russula*) in Ireland. *Biol. Invasions* **2016**, *18*, 857–871. [CrossRef]
56. Amori, G.; Castiglia, R. Mammal endemism in Italy: A review. *Biogeographia* **2018**, *33*, 19–31. [CrossRef]
57. Brambilla, M.; Vitulano, S.; Spina, F.; Baccetti, N.; Gargallo, G.; Fabbri, E.; Guidali, F.; Randi, E. A molecular phylogeny of the *Sylvia cantillans* complex: Cryptic species within the Mediterranean basin. *Mol. Phylogenet. Evol.* **2008**, *48*, 461–472. [CrossRef]
58. Gvoždík, V.; Canestrelli, D.; García-París, M.; Moravec, J.; Nascetti, G.; Recuero, E.; Teixeira, J.; Kotlík, P. Speciation history and widespread introgression in the European short-call tree frogs (*Hyla arborea sensu lato*, *H. intermedia* and *H. sarda*). *Mol. Phylogenet. Evol.* **2015**, *83*, 143–155. [CrossRef]
59. Mulargia, M.; Corti, C.; Lunghi, E. The herpetofauna of the Monte Albo, Sardinia (Italy). *Rus. J. Herpetol.* **2018**, *25*, 172–176. [CrossRef]



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).