



Review

Cardiac Biomarkers and Autoantibodies in Endurance Athletes: Potential Similarities with Arrhythmogenic Cardiomyopathy Pathogenic Mechanisms

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Citation: Stadiotti, I.; Lippi, M.; Maione, A.S.; Compagnucci, P.; Andreini, D.; Casella, M.; Pompilio, G.; Sommariva, E. Cardiac Biomarkers and Autoantibodies in Endurance Athletes: Potential Similarities with Arrhythmogenic Cardiomyopathy Pathogenic Mechanisms. *Int. J. Mol. Sci.* **2021**, *22*, 6500. <https://doi.org/10.3390/ijms22126500>

Academic Editor: Michael T. Chin

Received: 11 May 2021

Accepted: 15 June 2021

Published: 17 June 2021

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Abstract: The “Extreme Exercise Hypothesis” states that when individuals perform training beyond the ideal exercise dose, a decline in the beneficial effects of physical activity occurs. This is due to significant changes in myocardial structure and function, such as hemodynamic alterations, cardiac chamber enlargement and hypertrophy, myocardial inflammation, oxidative stress, fibrosis, and conduction changes. In addition, an increased amount of circulating biomarkers of exercise-induced damage has been reported. Although these changes are often reversible, long-lasting cardiac damage may develop after years of intense physical exercise. Since several features of the athlete’s heart overlap with arrhythmogenic cardiomyopathy (ACM), the syndrome of “exercise-induced ACM” has been postulated. Thus, the distinction between ACM and the athlete’s heart may be challenging. Recently, an autoimmune mechanism has been discovered in ACM patients linked to their characteristic junctional impairment. Since cardiac junctions are similarly impaired by intense physical activity due to the strong myocardial stretching, we propose in the present work the novel hypothesis of an autoimmune response in endurance athletes. This investigation may deepen the knowledge about the pathological remodeling and relative activated mechanisms induced by intense endurance exercise, potentially improving the early recognition of whom is actually at risk.

Keywords: arrhythmogenic cardiomyopathy; athletes; autoantibodies; physical exercise; desmosomes

1. Introduction

Although physical exercise is recommended for the maintenance of a healthy lifestyle and the reduction of cardiovascular disease incidence [1,2], prolonged and intense activity can be deleterious for cardiac structure and function. It can acutely and transiently increase sudden cardiac death (SCD) and myocardial infarction risk in susceptible individuals [3]. Increased myocardial fibrosis [4,5], coronary artery calcification [6], and atrial fibrillation [7,8] have been reported in endurance athletes.

Since endurance athletes exceed the usual recommendations for exercise by 15-fold to 20-fold, the “Extreme Exercise Hypothesis” has been proposed to explain how the beneficial effects of physical activity may plateau or decline when individuals perform training beyond the ideal exercise dose [9,10]. As depicted in Figure 1, the dose–response relationship

between exercise training volumes and health risk is described by a J-shaped (or U-shaped) curve [2,9]. To date, the exact amount of exercise able to impair the cardiovascular system has not been defined. The metabolic equivalents of task (METs) method is recognized as useful to evaluate the functional capacity or exercise tolerance of an individual [11]. One MET is the amount of oxygen consumed at rest and is equal to 3.5 mL of oxygen per kilogram per minute [11]. Most reports have defined “vigorous exercise” as needing at least six METs, although the maximal individual capacity could influence this threshold [3].

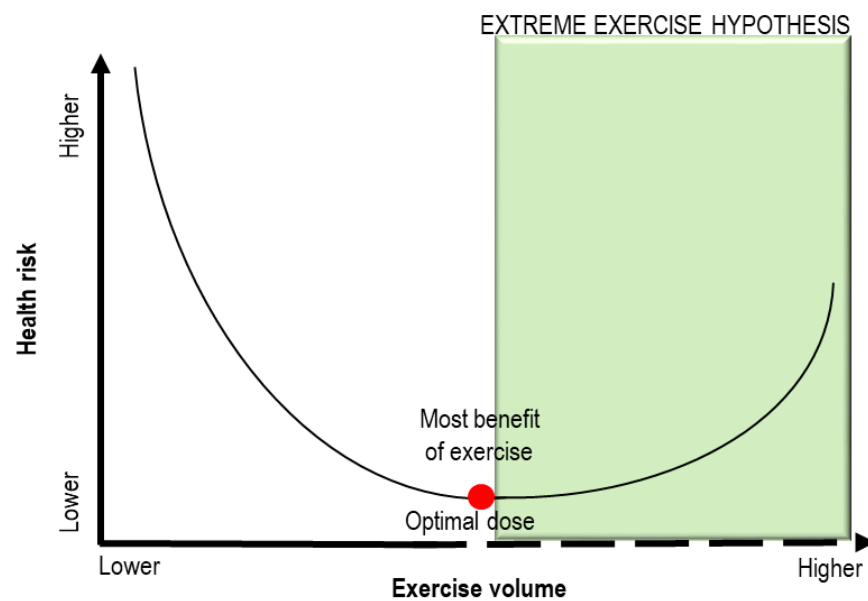


Figure 1. Schematic representation of the “Extreme Exercise Hypothesis”. A J-shaped (or U-shaped) curve describes the dose–response relationship between exercise training volumes and health risk. When the threshold of optimal exercise dose (red point) is exceeded, the health benefits of training can be reduced. Adapted from [9].

Most exercise-associated adverse effects often occur in subjects with occult or diagnosed structural cardiac diseases [3,12]. Among young individuals, the concomitant pathological conditions are often hereditary or congenital cardiovascular abnormalities, such as arrhythmogenic cardiomyopathy, hypertrophic cardiomyopathy, coronary artery anomalies, and bicuspid aortic valve [13–15]. Among older subjects who die during physical exercise, coronary artery disease is the most frequent pathological finding [16,17].

SCD overall incidence during exercise is estimated at 1:50,000 [18–20]. Exercise-related SCD seems to depend on the interaction between the physical activity acute trigger and an underlying disease, but it can be further elicited by other concomitant processes, including emotional stress, hemodynamic changes, and impaired parasympathetic tone [3]. It has been reported that physical activity may increase the risk of SCD by 2.5 times [21].

In the present review, we summarize the current knowledge about exercise-induced cardiac alterations and circulating biomarkers of damage. In addition, we propose a novel hypothesis of an autoimmune response in endurance athletes, based on the analogies with arrhythmogenic cardiomyopathy patients.

1.1. The Athlete’s Heart

Vigorous physical exercise is associated with significant changes in myocardial structure and function. The athlete’s heart has to sustain a higher cardiac output during maximal effort than untrained hearts. Thus, it is subjected to a physiological remodeling that allows its greater resistance during intense activity and sufficient oxygen delivery to exercising muscles. This physiological response is known as the Frank–Starling Mechanism or “law of the heart” [22].

Sympathetic activation is responsible for the augmented cardiac output through heart rate modulation. The heart rate spans from <40 bpm at rest to >200 bpm in a young maximally exercising individual. The stroke volume may significantly increase with sustained training because of the higher ventricular end-diastolic volume and sympathetically mediated end-systolic volume reduction [23]. The hemodynamic changes parallel cardiac chamber enlargement and hypertrophy [24–26]. These cardiac adaptations may mimic those of a diseased heart, but in most cases systolic and diastolic functions are preserved [27,28], although a transient reduction in left ventricular (LV) ejection fraction (EF) has been reported after more than 6 h of continuous exercise [29,30]. In many cases, the right ventricular (RV) function seems to be more compromised by prolonged exercise than left ventricular one, possibly for the thinner-walled structure of RV [31–33]. Indeed, RV wall stress increases more than in the LV during exercise, producing higher pressure load on the RV, not compensated by a sufficient volume increase and myocardial thickening [34]. This depends on the increase in pulmonary artery pressure relative to systemic vascular pressure, necessary to guarantee the requested cardiac output [35]. However, at equal exercise loads, the RV response shows high interindividual variability due to differential adaptations of the pulmonary circulation. Indeed, a higher vascular reserve corresponds to enhanced maximal exercise capacity [36]. In addition, conduction alterations are common, and they are usually mediated by parasympathetic activity and/or sinoatrial node slowing [37]. Endurance athletes are often affected by bradyarrhythmias, such as sinus bradycardia, junctional bradycardia, and first-degree atrioventricular block [38]. Trained athletes can experience premature beats and non-sustained ventricular tachycardia, usually of benign etiology and without long-term consequences [39,40], although no higher prevalence if compared to sedentary individuals has been assessed [41]. The main problematic effect of intense physical activity is atrial fibrillation, which has been reported more frequently among athletes than in sedentary individuals [42,43]. Syncope often manifests in the immediate post-exercise period due to neurocardiogenic mechanisms based on a sudden reduction in venous return. When syncope manifests during exercise, it can be due to malignant arrhythmias, structural cardiac disease, or myocardial ischemia, which have to be thoroughly evaluated [44].

Although these alterations are often reversible, long-lasting cardiac damage may develop after years of intense physical exercise [45,46].

Furthermore, after prolonged endurance exercise, myocardial inflammation, oxidative stress, and fibrosis have often been reported, representing a substrate for life-threatening arrhythmias [5,10,47].

During exercise, increased metabolic processes with an augmented oxygen uptake may induce a mitochondrial electron “leakage” and the consequent production of reactive oxygen species (ROS) [48,49]. Moreover, the activation of immune and inflammatory responses due to exercise-induced muscle injury may generate high amounts of ROS [50,51]. To possibly counterbalance the oxidative damage, an increase in antioxidant defenses, through the activation of antioxidant enzymes [52], has been reported in response to high volumes of exercise [48]. Thus, it seems that oxidative stress does not occur below a certain threshold of intensity, but only when exercise is strenuous [53].

The higher amount of oxidative stress can increase oxidation of different molecules, causing their damage. For example, acute bouts of exercise can increase LDL oxidation [54–57].

The oxidative radicals could impair cardiomyocyte membrane permeability, concurring with mechanical stress in cardiomyocyte remodeling [58,59]. In addition, ROS may inhibit glycolysis, producing a perturbation in calcium homeostasis, which could lead to myocardial dysfunction [60].

Exercise-induced myocardial fibrosis patterns are various and differ according to the age of the athletes [61–63]. Fibrosis is often found near the interventricular septum, especially in middle-aged and older athletes, and near the right ventricular insertion points, mainly in young athletes [4,5,61]. More rarely, a sub-endocardial ischemic pattern, a sub-epicardial pattern, and extensive mid-wall and diffuse fibrosis could be detected. Since only

specific fibrotic patterns have been associated with ventricular arrhythmias and adverse cardiac events, the clinical and prognostic significance of myocardial fibrosis in athletes is yet to be determined [63]. The dose of exercise has been reported to be associated with the extent of fibrotic substitution in few studies [5,9,64].

The impact of gender on the ventricular response to exercise constitutes a relevant open issue. Although male versus female differences have a strong impact on cardiovascular disease pathogenesis [65–67], female athletes are often underrepresented in studies of cardiac adaptation to exercise. Few studies on this topic demonstrated a similar cardiac remodeling and prevalence of arrhythmic events in both male and female athletes [68,69]. However, RV performance during exercise seems to be enhanced in women when compared to men [68]. Conversely, a recent study reported that LV remodeling is more common in males, whereas RV remodeling mainly concerns females [70]. Thus, further investigations are needed to clarify the effective impact of gender.

1.2. Effects of Myocardial Stretching

During intense exercise, the stretching of the myocardium activates different intrinsic physiologic mechanisms to adequately respond to this stimulus [71], through the so-called mechanoelectric feedback, which is able to transduce the mechanical stimulus into an electrical signal [72,73].

Each component of the heart seems to perceive mechanical stimuli, activating intracellular pathways that mediate several responses [74,75]. These pathways are often activated without binding of extracellular mediators [76]. An example is stretch-activated channels, able to modulate their permeability to ions and, consequently, electrical and mechanical properties of the myocardium [77]. This response often depends on protein phosphorylation. For example, calcium channel phosphorylation, by intensifying calcium transient, improves the contractile function [78]. Besides ion channels, other cardiomyocyte proteins concur in stretch-activated mechanisms: troponin I phosphorylation increases contractility, owing to the reduction of myofilament calcium sensitivity [79]; titin, both functioning as a mechanosensor and a molecular target, can trigger downstream signaling and modulate myocardial tension and sarcomeric length [80–82].

A central role in mechanosensing is played by intercalated discs, required to maintain mechanical and electric coupling between cardiomyocytes [83]. The two main structures of intercalated discs, fascia adherens junctions and desmosomes, contribute to adaptive responses to stretch [84] due to their connection with cytoskeletal actin and intermediate filaments, respectively [83]. In volume overload conditions, as during intense physical activity, the intercalated discs undergo dynamic changes [85]. For example, N-cadherin (N-CAD), one of the main proteins of fascia adherens junctions, is upregulated following mechanical stretch and elicits changes in cardiomyocyte shape, myofibrillar organization, and function [86]. On the contrary, N-CAD downregulation precludes the correct formation of intercalated discs, provoking cardiac morphological and functional defects [87].

Similarly, desmosomal protein loss impairs mechanotransduction responses [83]. For example, the deletion of the desmosomal protein desmoglein 2 (DSG2), necessary to assemble the extracellular domain, alters cell adhesion and signaling. It provokes the upregulation of heart failure markers, fibrosis, biventricular dilation and dysfunction, and death [88–90]. In general, desmosome deficiency leads to cardiomyocyte inability to appropriately face high mechanical stress, resulting in myocyte detachment and tissue remodeling [84].

Moreover, gap junctions, prominently localized at intercalated discs, mediate electrical propagation and are thus crucial to excitation and contraction [91]. They are composed of connexins, among which connexin 43 (CX43), the most important in the myocardium [91]. Physical exercise may affect gap junction remodeling, leading to CX43 expression downregulation during acute exercise, as demonstrated in a murine model [92], and to a possible consequent impairment of electrical conduction [93].

Additionally, costamere proteins, which are responsible for the connection between the contractile apparatus and extracellular matrix, as integrins, are involved in mechanotransduction and can be compromised when subjected to mechanical stress [94–97].

All these modifications generally have an adaptive meaning, but, depending on the strength of the stimulus, their nature, and the individual's genetic background, they can result in maladaptive pathological remodeling [98], with mechanoelectric feedback dysfunction [72] and consequent arrhythmias, cardiac hypertrophy, and heart failure [74,84]. Indeed, when the physical activity is prolonged and intense, the impairment of these processes may provoke an altered cellular response and possibly heart disease.

1.3. Circulating Biomarkers of Exercise-Induced Damage

The changes induced by endurance exercise are associated with several circulating biomarker increases. These elevations are usually modest and transient, but their clinical implications are not fully elucidated.

As for cardiac damage biomarker, cardiac troponin (cTn) levels significantly increase after only 30 min of intense physical activity [99], reaching higher levels in younger and untrained individuals [100], concomitant with cardiovascular risk factors [101], greater exercise duration and intensity [32,101–105], and dehydration [106]. cTn release likely depends on exercise-induced cardiomyocyte necrosis, or the changes in membrane permeability caused by intense activity could determine the leakage of unbound troponin [107]. Further studies are needed to understand the mechanisms mediating its elevation [59,108]. Usually, cTn levels return to baseline within 72 h [109,110], and any cardiac dysfunction associated with increased cTn has been reported transient [59].

B-type natriuretic peptide (BNP) and its cleaved form NT-proBNP are secreted in response to cardiomyocyte stress produced by volume or pressure overload [111]. Thus, they can increase after endurance exercise [32,112–117], but return to baseline within 72 h [109,118]. Exercise duration [104,115], age [113,118], and poor physical preparation [32,112,119] can impact on BNP and NT-proBNP elevation.

Creatine kinase MB (CKMB), belonging to myocardial infarction biomarkers, can also be increased after intense activity, but it possibly originates more from skeletal muscle damage than from myocardial injury [120].

Moreover, typical fibrosis biomarkers have been associated with intense physical exercise. Soluble suppression of tumorigenicity 2 (sST-2) concentrations exceed the upper reference value after endurance activity, reaching higher levels as exercise intensity increases, but its complete normalization occurs within 48 h [121].

Tissue inhibitors of matrix metalloproteinase type I (TIMP-1), carboxy-terminal telopeptide of collagen type I (CITP), and carboxy-terminal propeptide of collagen type I (PICP) are other circulating markers of collagen synthesis and degradation that are augmented in endurance athletes [59].

Similarly, galectin-3 resting levels are greater in athletes than controls, and further increase after physical activity, possibly produced mainly by skeletal muscle [122]. Indeed, no correlations with cardiac function have been detected [122].

For what concern oxidative stress markers, 13- and 9-hydroxy-octadecadienoic acid (13-HODE and 9-HODE), known oxidized linoleic acid metabolites, significantly increase immediately post-exercise, but their plasma concentrations return to baseline levels within 24 h [123]. Their production could be linked to lipoxygenase activation in response to cell injury [124].

Moreover, lipid peroxidation increases after endurance exercise, as demonstrated by higher levels of malondialdehyde (MDA) [117,125] and F(2)-isoprostanes [57]. In both cases, the augmentation is transient.

Similarly, the heat shock proteins Hsp70 and Hsp72, known inflammation markers, are upregulated in athletes' serum after physical exercise [126–128]. As for the majority of the exercise-induced circulating biomarkers, the increase is rapid but transient. Higher

levels of these proteins has been previously reported in failing hearts [129]. Thus, their release is not specifically associated with exercise-induced alterations.

The current information about circulating factors in athletes is limited and a broader analysis (e.g., microRNA) is required [130,131]. To date, no biomarkers have been associated with the adverse remodeling that can occur after endurance activity with a cause–effect specificity. Indeed, most of the described biomarkers are elevated also in concomitance with various arrhythmic and heart failure diseases [132], making the understanding of the underlying causes difficult.

2. Exercise-Induced Arrhythmogenic Cardiomyopathy

Together with polymorphic ventricular tachycardia [133] and hypertrophic cardiomyopathy [134,135], arrhythmogenic cardiomyopathy (ACM) is included among the cardiac diseases that can be induced by physical exercise.

ACM is a genetic cardiac disorder, predominantly affecting the RV [136]. It is characterized by a fibro-adipose replacement of the ventricular myocardium, malignant arrhythmias, and SCD, especially in young adults and athletes [136]. Often, ACM penetrance is incomplete and genotype-phenotype correlations are difficult to establish [137]. Among the pathogenesis cofactors that have been proposed to explain ACM variable expressivity, physical exercise exposes ACM patients to a five-fold higher risk of SCD if compared to sedentary patients [138]. Due to the critical RV degeneration and arrhythmogenicity following repetitive intense exercise [31–33], even in the absence of known genetic abnormalities [139], the syndrome of “exercise-induced ACM” was proposed few years ago [140,141].

Although only a small fraction of high-level athletes develops signs of RV cardiomyopathy, the hypothesis is that endurance exercise provokes RV insults, in line with circulating cardiac damage biomarker rise and dysfunction, that, in the long term, could have pathological implications [142,143]. According to this hypothesis, an ACM-like phenotype may be acquired and not unquestionably ascribed to a genetic predisposition. Studies with a rat model of long-term intensive exercise training supported this theory, proving training-dependent RV fibrosis and increased arrhythmia inducibility after chronic endurance exercise [144].

If definite ACM patients carry, in the majority of the cases, genetic mutations that impair desmosomes, exercise-induced ACM is likely to involve cardiac junctions as well. Indeed, in the latter case, disproportionate wall stress, caused by intense and prolonged physical exercise, disrupts “normal” desmosomes [142], and the mismatch between wall stress and desmosomal integrity can be associated with the huge hemodynamic stress [142].

A debate about the effective existence of these exercise-induced ACM forms is still open and not all the involved factors have been unraveled. Indeed, no proper studies have confirmed or denied the existence of ACM phenotypes exclusively induced by exercise [145]. Additionally, gene-elusive patients represent a heterogeneous group, since some might have ACM causative genetic variants in genes not yet identified as associated. Possibly, other factors can predispose a subpopulation of endurance athletes to this condition, such as an unrecognized genetic predisposition that phenotypically manifests only in the setting of extreme exercise [146]. Moreover, the study of the underlying molecular mechanisms is still difficult for the limitations of the current *in vivo* models of endurance exercise [147].

The distinction between ACM and the athlete’s heart is still a challenge for sports cardiologists because of the overlapping features [148–151]. Indeed, studies comparing athletes with and without recognized genetic mutations often described similar clinical phenotypes (Figure 2). The proportion of individuals experiencing a major arrhythmic event during the follow up is comparable (28% of the cases), and electrocardiographic signs, including the ACM diagnostic major criterion epsilon wave, were reported to be similarly represented [139], except for the presence of pathological Q waves only in ACM patients [149]. The impairment of signal average ECG (SAECG) parameters has been more

frequently reported in traditional ACM cases [149]. ACM patients are characterized by more severe LV and/or RV dilation and dysfunction, as confirmed by the stronger EF reduction [139,142], but the increase in RV size is similar in ACM and exercise-induced ACM individuals [152]. As for tissue remodeling, a typical feature of ACM patients is fibro-fatty substitution, whereas only fibrosis has been found in exercise-induced ACM individuals [139]. In addition, ACM patients exhibit delayed gadolinium enhancement [149]. On the other hand, histological abnormalities and cardiac inflammation and oxidative stress are shared by the two types of patients [12,142].

	Traditional ACM	Athlete heart / Exercise-induced ACM
Cardiac structure	<ul style="list-style-type: none"> Severe LV and/or RV dilation 	<ul style="list-style-type: none"> Increased RV size
Function	<ul style="list-style-type: none"> Severe LV and/or RV dysfunction RV kinetic abnormalities 	<ul style="list-style-type: none"> Mild and reversible LV and RV dysfunction
Tissue remodeling	<ul style="list-style-type: none"> Cardiac fibro-fatty substitution 	<ul style="list-style-type: none"> Myocardial inflammation Cardiac oxidative stress
Electrical abnormalities	<ul style="list-style-type: none"> Frequent non-sustained ventricular tachycardia / ventricular tachycardia 	<ul style="list-style-type: none"> Major arrhythmic events ECG alterations (T wave inversion; epsilon wave; prolonged QRS terminal activation duration) Ventricular conduction defects, mainly in the RV
Family history	<ul style="list-style-type: none"> Genetic basis 	

Figure 2. Comparison of ACM and exercise-induced ACM clinical phenotypes. The diagram illustrates differences and similarities between ACM and exercise-induced ACM from a clinical point of view.

Unfortunately, despite the intensive efforts to discriminate traditional ACM patients from exercise-induced ACM individuals, there is still the need to improve the diagnostic procedure. In addition, the comparison between the features of ACM left dominant and biventricular forms to exercise-induced remodeling could be of further help.

3. Autoimmune Response Hypothesis in Endurance Athletes

Based on the available information, a parallelism between the negative effects induced by intense exercise at cardiac level and the typical characteristics of ACM, mainly caused by genetic mutations that affect desmosomes, has been proposed by coining the term “exercise-induced ACM” [140,141]. Although a proper description of the features that can discriminate the two forms is still lacking, some lessons might be learned from the actual knowledge of ACM. Recently, circulating autoantibodies against DSG2, one of the desmosomal proteins, have been found in ACM patients, and not in healthy controls and subjects affected by other cardiomyopathies [153]. These results are promising for the development of a novel diagnostic test for ACM patients and to potentially discriminate definite ACM patients from people affected by other conditions in differential diagnosis [132]. However, can they discriminate ACM from exercise-induced ACM?

The generation of these autoantibodies is likely to be dependent on DSG2 release into the intercellular space and circulation due to an ACM causative genetic mutation [153]. DSG2 may include “cryptic” epitopes that can induce, once exposed, an immune response [153].

However, no authors have to date assessed if the same autoimmune mechanisms interest endurance athletes. Indeed, as described above, among the induced modifications, intense physical exercise can challenge cell junctions, especially fascia adherens and desmosomes [84], and the expression and localization of the constituent proteins may be altered [85]. Although their remodeling usually ensues to properly respond to a greater mechanical stress, when a threshold of intensity is exceeded, dysfunction may occur due to increased oxidative stress [48,49] and changes in cell permeability [107]. The extent of fibrotic substitution that can follow endurance activity exacerbates the reduction of cell-to-cell contact [154]. Accordingly, the disruption of epithelial tight junctions has been already described during exercise due to heat- and oxidative damage-mediated stress [155]. Therefore, similar mechanisms are likely to impact on other cell junctions also at the cardiac level. In the worst scenario, all these processes could induce the detachment of intercellular junctional proteins, such as DSG2. For these reasons, the analysis of endurance athletes' plasma for anti-DSG2 autoantibody is awaited to assess the potential of this immune biomarker in distinguishing ACM from athlete's heart remodeling.

Other than their role as circulating biomarkers, DSG2 autoantibodies may affect cardiac function, as suggested by Chatterjee and colleagues [153]. Their levels correlate with premature ventricular contraction count in ACM patients, and *in vitro* analyses revealed their ability to functionally affect gap junctions. This is in line with the increasingly recognized ability of autoantibodies in arrhythmia stimulation through ion channels interference [156]. Moreover, DSG2 autoantibodies may attack the desmosomes and the whole intercalated discs, further weakening these structures. This is likely to happen similarly to what is described for anti-DSG3 autoantibodies produced against the cutaneous isoform of desmoglein in some skin disorders [157]. These autoantibodies induce DSG3 internalization and redistribution, altering the dynamics of desmosome assembly [158] and increasing tissue fragility in this already diseased state [159].

In addition, the inflammation triggered by this faulty immune response may aggravate the exercise-induced dysfunction [160].

In this view, in addition to physical detraining, immunosuppressive measures, including immunomodulatory drugs, plasmapheresis, or immunoadsorption for autoantibody removal, may be of help for at risk individuals.

4. Conclusions

Although physical exercise is an important measure to reduce cardiovascular disease incidence, excessive endurance training can paradoxically increase SCD and myocardial infarction risk in susceptible individuals as explained by the "Extreme Exercise Hypothesis". Several exercise-induced features overlap with ACM and make the distinction between ACM and the athlete's heart challenging. A debate about the existence of ACM phenotypes exclusively induced by exercise is still open. More information about the pathological remodeling and relative activated mechanisms induced by intense endurance exercise needs to be collected. Since not all the individuals practicing sports at high levels manifest exercise-induced cardiac alterations, the early recognition of whom is actually at risk is fundamental. The improvement of pre-participation screenings and the evaluation of new potential circulating biomarkers could be of considerable help.

Author Contributions: Conceptualization, I.S. and E.S.; writing—original draft preparation, I.S. and E.S.; writing—review and editing, I.S., M.L., A.S.M., P.C., D.A., M.C., G.P., E.S.; supervision, E.S. and G.P.; funding acquisition, E.S. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by Italian Ministry of Health, grant number RC2019 EF5C ID:2754330.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Shiroma, E.J.; Lee, I.M. Physical activity and cardiovascular health: Lessons learned from epidemiological studies across age, gender, and race/ethnicity. *Circulation* **2010**, *122*, 743–752. [[CrossRef](#)]
2. Franklin, B.A.; Thompson, P.D.; Al-Zaiti, S.S.; Albert, C.M.; Hivert, M.F.; Levine, B.D.; Lobelo, F.; Madan, K.; Sharrief, A.Z.; Eijsvogels, T.M.H.; et al. Exercise-related acute cardiovascular events and potential deleterious adaptations following long-term exercise training: Placing the risks into perspective-An update: A scientific statement from the American Heart Association. *Circulation* **2020**, *141*, e705–e736. [[CrossRef](#)] [[PubMed](#)]
3. Thompson, P.D.; Franklin, B.A.; Balady, G.J.; Blair, S.N.; Corrado, D.; Estes, N.A., 3rd; Fulton, J.E.; Gordon, N.F.; Haskell, W.L.; Link, M.S.; et al. Exercise and acute cardiovascular events placing the risks into perspective: A scientific statement from the American Heart Association Council on Nutrition, Physical Activity, and Metabolism and the Council on Clinical Cardiology. *Circulation* **2007**, *115*, 2358–2368. [[CrossRef](#)] [[PubMed](#)]
4. Breuckmann, F.; Mohlenkamp, S.; Nassenstein, K.; Lehmann, N.; Ladd, S.; Schmermund, A.; Sievers, B.; Schlosser, T.; Jockel, K.H.; Heusch, G.; et al. Myocardial late gadolinium enhancement: Prevalence, pattern, and prognostic relevance in marathon runners. *Radiology* **2009**, *251*, 50–57. [[CrossRef](#)] [[PubMed](#)]
5. Wilson, M.; O'Hanlon, R.; Prasad, S.; Deighan, A.; Macmillan, P.; Oxborough, D.; Godfrey, R.; Smith, G.; Maceira, A.; Sharma, S.; et al. Diverse patterns of myocardial fibrosis in lifelong, veteran endurance athletes. *J. Appl. Physiol.* **2011**, *110*, 1622–1626. [[CrossRef](#)]
6. Mohlenkamp, S.; Lehmann, N.; Breuckmann, F.; Brocker-Preuss, M.; Nassenstein, K.; Halle, M.; Budde, T.; Mann, K.; Barkhausen, J.; Heusch, G.; et al. Running: The risk of coronary events: Prevalence and prognostic relevance of coronary atherosclerosis in marathon runners. *Eur. Heart J.* **2008**, *29*, 1903–1910. [[CrossRef](#)]
7. Mussigbrodt, A.; Weber, A.; Mandrola, J.; van Belle, Y.; Richter, S.; Doring, M.; Arya, A.; Sommer, P.; Bollmann, A.; Hindricks, G. Excess of exercise increases the risk of atrial fibrillation. *Scand. J. Med. Sci. Sports* **2017**, *27*, 910–917. [[CrossRef](#)]
8. Aizer, A.; Gaziano, J.M.; Cook, N.R.; Manson, J.E.; Buring, J.E.; Albert, C.M. Relation of vigorous exercise to risk of atrial fibrillation. *Am. J. Cardiol.* **2009**, *103*, 1572–1577. [[CrossRef](#)]
9. Eijsvogels, T.M.H.; Thompson, P.D.; Franklin, B.A. The extreme exercise hypothesis: Recent findings and cardiovascular health implications. *Curr. Treat. Options Cardiovasc. Med.* **2018**, *20*, 84. [[CrossRef](#)]
10. Franklin, B.A.; Billecke, S. Putting the benefits and risks of aerobic exercise in perspective. *Curr. Sports Med. Rep.* **2012**, *11*, 201–208. [[CrossRef](#)]
11. Jette, M.; Sidney, K.; Blumchen, G. Metabolic equivalents (METs) in exercise testing, exercise prescription, and evaluation of functional capacity. *Clin. Cardiol.* **1990**, *13*, 555–565. [[CrossRef](#)]
12. Dello Russo, A.; Pieroni, M.; Santangeli, P.; Bartoletti, S.; Casella, M.; Pelargonio, G.; Smaldone, C.; Bianco, M.; Di Biase, L.; Bellocci, F.; et al. Concealed cardiomyopathies in competitive athletes with ventricular arrhythmias and an apparently normal heart: Role of cardiac electroanatomical mapping and biopsy. *Heart Rhythm* **2011**, *8*, 1915–1922. [[CrossRef](#)] [[PubMed](#)]
13. Maron, B.J.; Thompson, P.D.; Ackerman, M.J.; Balady, G.; Berger, S.; Cohen, D.; Dimeff, R.; Douglas, P.S.; Glover, D.W.; Hutter, A.M., Jr.; et al. Recommendations and considerations related to preparticipation screening for cardiovascular abnormalities in competitive athletes: 2007 update: A scientific statement from the American Heart Association Council on Nutrition, Physical Activity, and Metabolism: Endorsed by the American College of Cardiology Foundation. *Circulation* **2007**, *115*, 1643–1655. [[CrossRef](#)] [[PubMed](#)]
14. Kochi, A.N.; Vettor, G.; Dessanai, M.A.; Pizzamiglio, F.; Tondo, C. Sudden Cardiac Death in Athletes: From the basics to the practical work-up. *Medicina* **2021**, *57*, 168. [[CrossRef](#)] [[PubMed](#)]
15. Volpato, G.; Falanga, U.; Cipolletta, L.; Conti, M.A.; Grifoni, G.; Ciliberti, G.; Urbinati, A.; Barbarossa, A.; Stronati, G.; Fogante, M.; et al. Sports Activity and Arrhythmic Risk in Cardiomyopathies and Channelopathies: A Critical Review of European Guidelines on Sports Cardiology in Patients with Cardiovascular Diseases. *Medicina* **2021**, *57*, 308. [[CrossRef](#)] [[PubMed](#)]
16. Thompson, P.D.; Stern, M.P.; Williams, P.; Duncan, K.; Haskell, W.L.; Wood, P.D. Death during jogging or running. A study of 18 cases. *JAMA* **1979**, *242*, 1265–1267. [[CrossRef](#)] [[PubMed](#)]
17. Burke, A.P.; Farb, A.; Malcom, G.T.; Liang, Y.; Smialek, J.E.; Virmani, R. Plaque rupture and sudden death related to exertion in men with coronary artery disease. *JAMA* **1999**, *281*, 921–926. [[CrossRef](#)]
18. Maron, B.J.; Poliac, L.C.; Roberts, W.O. Risk for sudden cardiac death associated with marathon running. *J. Am. Coll. Cardiol.* **1996**, *28*, 428–431. [[CrossRef](#)]
19. Harris, K.M.; Henry, J.T.; Rohman, E.; Haas, T.S.; Maron, B.J. Sudden death during the triathlon. *JAMA* **2010**, *303*, 1255–1257. [[CrossRef](#)] [[PubMed](#)]
20. Kim, J.H.; Malhotra, R.; Chiampas, G.; d'Hemecourt, P.; Troyanos, C.; Cianca, J.; Smith, R.N.; Wang, T.J.; Roberts, W.O.; Thompson, P.D.; et al. Cardiac arrest during long-distance running races. *New Engl. J. Med.* **2012**, *366*, 130–140. [[CrossRef](#)]
21. Corrado, D.; Basso, C.; Rizzoli, G.; Schiavon, M.; Thiene, G. Does sports activity enhance the risk of sudden death in adolescents and young adults? *J. Am. Coll. Cardiol.* **2003**, *42*, 1959–1963. [[CrossRef](#)] [[PubMed](#)]

22. Patterson, S.W.; Starling, E.H. On the mechanical factors which determine the output of the ventricles. *J. Physiol.* **1914**, *48*, 357–379. [[CrossRef](#)] [[PubMed](#)]
23. Iatridis, P.G. Human circulation: Regulation during physical stress. *JAMA* **1987**, *258*, 3316. [[CrossRef](#)]
24. Roeske, W.R.; O'Rourke, R.A.; Klein, A.; Leopold, G.; Karliner, J.S. Noninvasive evaluation of ventricular hypertrophy in professional athletes. *Circulation* **1976**, *53*, 286–291. [[CrossRef](#)]
25. Scharhag, J.; Schneider, G.; Urhausen, A.; Rochette, V.; Kramann, B.; Kindermann, W. Athlete's heart: Right and left ventricular mass and function in male endurance athletes and untrained individuals determined by magnetic resonance imaging. *J. Am. Coll. Cardiol.* **2002**, *40*, 1856–1863. [[CrossRef](#)]
26. Scharf, M.; Brem, M.H.; Wilhelm, M.; Schoepf, U.J.; Uder, M.; Lell, M.M. Cardiac magnetic resonance assessment of left and right ventricular morphologic and functional adaptations in professional soccer players. *Am. Heart J.* **2010**, *159*, 911–918. [[CrossRef](#)]
27. Baggish, A.L.; Wood, M.J. Athlete's heart and cardiovascular care of the athlete: Scientific and clinical update. *Circulation* **2011**, *123*, 2723–2735. [[CrossRef](#)]
28. Gilbert, C.A.; Nutter, D.O.; Felner, J.M.; Perkins, J.V.; Heymsfield, S.B.; Schlant, R.C. Echocardiographic study of cardiac dimensions and function in the endurance-trained athlete. *Am. J. Cardiol.* **1977**, *40*, 528–533. [[CrossRef](#)]
29. Carrio, I.; Serra-Grima, R.; Berna, L.; Estorch, M.; Martinez-Duncker, C.; Ordóñez, J. Transient alterations in cardiac performance after a six-hour race. *Am. J. Cardiol.* **1990**, *65*, 1471–1474. [[CrossRef](#)]
30. Whyte, G.P.; George, K.; Sharma, S.; Lumley, S.; Gates, P.; Prasad, K.; McKenna, W.J. Cardiac fatigue following prolonged endurance exercise of differing distances. *Med. Sci. Sports Exerc.* **2000**, *32*, 1067–1072. [[CrossRef](#)]
31. La Gerche, A.; Connelly, K.A.; Mooney, D.J.; MacIsaac, A.I.; Prior, D.L. Biochemical and functional abnormalities of left and right ventricular function after ultra-endurance exercise. *Heart* **2008**, *94*, 860–866. [[CrossRef](#)]
32. Neilan, T.G.; Januzzi, J.L.; Lee-Lewandrowski, E.; Ton-Nu, T.T.; Yoerger, D.M.; Jassal, D.S.; Lewandrowski, K.B.; Siegel, A.J.; Marshall, J.E.; Douglas, P.S.; et al. Myocardial injury and ventricular dysfunction related to training levels among nonelite participants in the Boston marathon. *Circulation* **2006**, *114*, 2325–2333. [[CrossRef](#)] [[PubMed](#)]
33. Trivax, J.E.; Franklin, B.A.; Goldstein, J.A.; Chinnaiyan, K.M.; Gallagher, M.J.; deJong, A.T.; Colar, J.M.; Haines, D.E.; McCullough, P.A. Acute cardiac effects of marathon running. *J. Appl. Physiol.* **2010**, *108*, 1148–1153. [[CrossRef](#)]
34. La Gerche, A.; Heidebuchel, H.; Burns, A.T.; Mooney, D.J.; Taylor, A.J.; Pflugger, H.B.; Inder, W.J.; Macisaac, A.I.; Prior, D.L. Disproportionate exercise load and remodeling of the athlete's right ventricle. *Med. Sci. Sports Exerc.* **2011**, *43*, 974–981. [[CrossRef](#)]
35. La Gerche, A.; Rakhit, D.J.; Claessen, G. Exercise and the right ventricle: A potential Achilles' heel. *Cardiovasc. Res.* **2017**, *113*, 1499–1508. [[CrossRef](#)] [[PubMed](#)]
36. Sanz-de la Garza, M.; Vaquer-Segui, A.; Duran, K.; Blanco, I.; Burgos, F.; Alsina, X.; Prat-Gonzalez, S.; Bijmens, B.; Sitges, M. Pulmonary transit of contrast during exercise is related to improved cardio-pulmonary performance in highly trained endurance athletes. *Eur. J. Prev. Cardiol.* **2020**, *27*, 1504–1514. [[CrossRef](#)] [[PubMed](#)]
37. Stein, R.; Medeiros, C.M.; Rosito, G.A.; Zimmerman, L.I.; Ribeiro, J.P. Intrinsic sinus and atrioventricular node electrophysiologic adaptations in endurance athletes. *J. Am. Coll. Cardiol.* **2002**, *39*, 1033–1038. [[CrossRef](#)]
38. Heidebuchel, H. The athlete's heart is a proarrhythmic heart, and what that means for clinical decision making. *Europace* **2018**, *20*, 1401–1411. [[CrossRef](#)] [[PubMed](#)]
39. Biffi, A.; Pelliccia, A.; Verdile, L.; Fernando, F.; Spataro, A.; Caselli, S.; Santini, M.; Maron, B.J. Long-term clinical significance of frequent and complex ventricular tachyarrhythmias in trained athletes. *J. Am. Coll. Cardiol.* **2002**, *40*, 446–452. [[CrossRef](#)]
40. Biffi, A.; Maron, B.J.; Verdile, L.; Fernando, F.; Spataro, A.; Marcello, G.; Ciardo, R.; Ammirati, F.; Colivicchi, F.; Pelliccia, A. Impact of physical deconditioning on ventricular tachyarrhythmias in trained athletes. *J. Am. Coll. Cardiol.* **2004**, *44*, 1053–1058. [[CrossRef](#)]
41. Zorzi, A.; De Lazzari, M.; Mastella, G.; Niero, A.; Trovato, D.; Cipriani, A.; Peruzza, F.; Portolan, L.; Berton, G.; Sciacca, F.; et al. Ventricular arrhythmias in young competitive athletes: Prevalence, determinants, and underlying substrate. *J. Am. Heart Assoc.* **2018**, *7*. [[CrossRef](#)]
42. Sorokin, A.V.; Araujo, C.G.; Zweibel, S.; Thompson, P.D. Atrial fibrillation in endurance-trained athletes. *Br. J. Sports Med.* **2011**, *45*, 185–188. [[CrossRef](#)]
43. Molina, L.; Mont, L.; Marrugat, J.; Berruezo, A.; Brugada, J.; Bruguera, J.; Rebato, C.; Elosua, R. Long-term endurance sport practice increases the incidence of lone atrial fibrillation in men: A follow-up study. *Europace* **2008**, *10*, 618–623. [[CrossRef](#)]
44. Koene, R.J.; Adkisson, W.O.; Benditt, D.G. Syncope and the risk of sudden cardiac death: Evaluation, management, and prevention. *J. Arrhythmia* **2017**, *33*, 533–544. [[CrossRef](#)]
45. O'Keefe, J.H.; Lavie, C.J. Run for your life...at a comfortable speed and not too far. *Heart* **2013**, *99*, 516–519. [[CrossRef](#)] [[PubMed](#)]
46. O'Keefe, J.H.; Patil, H.R.; Lavie, C.J. Exercise and life expectancy. *Lancet* **2012**, *379*, 799. [[CrossRef](#)]
47. Compagnucci, P.; Volpato, G.; Falanga, U.; Cipolletta, L.; Conti, M.A.; Grifoni, G.; Ciliberti, G.; Stronati, G.; Fogante, M.; Bergonti, M.; et al. Myocardial Inflammation, Sports Practice, and Sudden Cardiac Death: 2021 Update. *Medicina* **2021**, *57*, 277. [[CrossRef](#)] [[PubMed](#)]
48. Knez, W.L.; Coombes, J.S.; Jenkins, D.G. Ultra-endurance exercise and oxidative damage: Implications for cardiovascular health. *Sports Med.* **2006**, *36*, 429–441. [[CrossRef](#)]
49. Bailey, D.M.; Young, I.S.; McEneny, J.; Lawrenson, L.; Kim, J.; Barden, J.; Richardson, R.S. Regulation of free radical outflow from an isolated muscle bed in exercising humans. *Am. J. Physiol. Heart Circ. Physiol.* **2004**, *287*, H1689–H1699. [[CrossRef](#)] [[PubMed](#)]

50. Nieman, D.C.; Pedersen, B.K. Exercise and immune function. Recent developments. *Sports Med.* **1999**, *27*, 73–80. [[CrossRef](#)] [[PubMed](#)]
51. Hellsten, Y.; Frandsen, U.; Orthenblad, N.; Sjodin, B.; Richter, E.A. Xanthine oxidase in human skeletal muscle following eccentric exercise: A role in inflammation. *J. Physiol.* **1997**, *498*, 239–248. [[CrossRef](#)]
52. Tiidus, P.M.; Houston, M.E. Antioxidant and oxidative enzyme adaptations to vitamin E deprivation and training. *Med. Sci. Sports Exerc.* **1994**, *26*, 354–359. [[CrossRef](#)]
53. Powers, S.K.; Deminice, R.; Ozdemir, M.; Yoshihara, T.; Bomkamp, M.P.; Hyatt, H. Exercise-induced oxidative stress: Friend or foe? *J. Sport Health Sci.* **2020**, *9*, 415–425. [[CrossRef](#)]
54. Sanchez-Quesada, J.L.; Homs-Serradesanferm, R.; Serrat-Serrat, J.; Serra-Grima, J.R.; Gonzalez-Sastre, F.; Ordonez-Llanos, J. Increase of LDL susceptibility to oxidation occurring after intense, long duration aerobic exercise. *Atherosclerosis* **1995**, *118*, 297–305. [[CrossRef](#)]
55. Wetzstein, C.J.; Shern-Brewer, R.A.; Santanam, N.; Green, N.R.; White-Welkley, J.E.; Parthasarathy, S. Does acute exercise affect the susceptibility of low density lipoprotein to oxidation? *Free Radic. Biol. Med.* **1998**, *24*, 679–682. [[CrossRef](#)]
56. Liu, M.L.; Bergholm, R.; Makimattila, S.; Lahdenpera, S.; Valkonen, M.; Hilden, H.; Yki-Jarvinen, H.; Taskinen, M.R. A marathon run increases the susceptibility of LDL to oxidation in vitro and modifies plasma antioxidants. *Am. J. Physiol.* **1999**, *276*, E1083–E1091. [[CrossRef](#)] [[PubMed](#)]
57. Mastaloudis, A.; Leonard, S.W.; Traber, M.G. Oxidative stress in athletes during extreme endurance exercise. *Free Radic. Biol. Med.* **2001**, *31*, 911–922. [[CrossRef](#)]
58. Stavroulakis, G.A.; George, K.P. Exercise-induced release of troponin. *Clin. Cardiol.* **2020**, *43*, 872–881. [[CrossRef](#)]
59. Eijsvogels, T.M.; Fernandez, A.B.; Thompson, P.D. Are there deleterious cardiac effects of acute and chronic endurance exercise? *Physiol. Rev.* **2016**, *96*, 99–125. [[CrossRef](#)] [[PubMed](#)]
60. Forrester, S.J.; Kikuchi, D.S.; Hernandez, M.S.; Xu, Q.; Griendling, K.K. Reactive oxygen species in metabolic and inflammatory signaling. *Circ. Res.* **2018**, *122*, 877–902. [[CrossRef](#)]
61. van de Schoor, F.R.; Aengevaeren, V.L.; Hopman, M.T.; Oxborough, D.L.; George, K.P.; Thompson, P.D.; Eijsvogels, T.M. Myocardial fibrosis in athletes. In *Mayo Clinic Proceedings*; Elsevier: Amsterdam, The Netherlands, 2016; pp. 1617–1631. [[CrossRef](#)]
62. Zhang, C.D.; Xu, S.L.; Wang, X.Y.; Tao, L.Y.; Zhao, W.; Gao, W. Prevalence of myocardial fibrosis in intensive endurance training athletes: A systematic review and meta-analysis. *Front. Cardiovasc. Med.* **2020**, *7*, 585692. [[CrossRef](#)]
63. Domenech-Ximenes, B.; Sanz-de la Garza, M.; Prat-Gonzalez, S.; Sepulveda-Martinez, A.; Crispi, F.; Duran-Fernandez, K.; Perea, R.J.; Bijmens, B.; Sitges, M. Prevalence and pattern of cardiovascular magnetic resonance late gadolinium enhancement in highly trained endurance athletes. *J. Cardiovasc. Magn. Reson.* **2020**, *22*, 62. [[CrossRef](#)] [[PubMed](#)]
64. Tahir, E.; Starekova, J.; Muellerleile, K.; von Stritzky, A.; Munch, J.; Avanesov, M.; Weinrich, J.M.; Stehning, C.; Bohnen, S.; Radunski, U.K.; et al. Myocardial fibrosis in competitive triathletes detected by contrast-enhanced CMR correlates with exercise-induced hypertension and competition history. *JACC Cardiovasc. Imaging* **2018**, *11*, 1260–1270. [[CrossRef](#)]
65. Cedieli, G.; Codina, P.; Spitaleri, G.; Domingo, M.; Santiago-Vacas, E.; Lupon, J.; Bayes-Genis, A. gender-related differences in heart failure biomarkers. *Front. Cardiovasc. Med.* **2020**, *7*, 617705. [[CrossRef](#)] [[PubMed](#)]
66. Summerhill, V.I.; Moschetta, D.; Orekhov, A.N.; Poggio, P.; Myasoedova, V.A. Sex-specific features of calcific aortic valve disease. *Int. J. Mol. Sci.* **2020**, *21*, 5620. [[CrossRef](#)] [[PubMed](#)]
67. Group, E.U.C.C.S.; Regitz-Zagrosek, V.; Oertelt-Prigione, S.; Prescott, E.; Franconi, F.; Gerdt, E.; Foryst-Ludwig, A.; Maas, A.H.; Kautzky-Willer, A.; Knappe-Wegner, D.; et al. Gender in cardiovascular diseases: Impact on clinical manifestations, management, and outcomes. *Eur. Heart J.* **2016**, *37*, 24–34. [[CrossRef](#)]
68. Sanz-de la Garza, M.; Giraldeau, G.; Marin, J.; Grazioli, G.; Esteve, M.; Gabrielli, L.; Brambila, C.; Sanchis, L.; Bijmens, B.; Sitges, M. Influence of gender on right ventricle adaptation to endurance exercise: An ultrasound two-dimensional speckle-tracking stress study. *Eur. J. Appl. Physiol.* **2017**, *117*, 389–396. [[CrossRef](#)]
69. Quinto, G.; Neunhaeuserer, D.; Gasperetti, A.; Battista, F.; Foccardi, G.; Baiocco, V.; Gobbo, S.; Bergamin, M.; Ermolao, A. can exercise test intensity and modality affect the prevalence of arrhythmic events in young athletes? *Res. Sports Med.* **2021**, 1–9. [[CrossRef](#)]
70. Yoon, H.J.; Kim, K.H.; Hornsby, K.; Park, J.H.; Park, H.; Kim, H.Y.; Cho, J.Y.; Ahn, Y.; Jeong, M.H.; Cho, J.G. Gender difference of cardiac remodeling in university athletes: Results from 2015 Gwangju Summer Universiade. *Korean Circ. J.* **2021**, *51*, 426–438. [[CrossRef](#)]
71. Neves, J.S.; Leite-Moreira, A.M.; Neiva-Sousa, M.; Almeida-Coelho, J.; Castro-Ferreira, R.; Leite-Moreira, A.F. Acute myocardial response to stretch: What we (don't) know. *Front. Physiol.* **2015**, *6*, 408. [[CrossRef](#)]
72. Lab, M.J. Mechanoelectric feedback (transduction) in heart: Concepts and implications. *Cardiovasc. Res.* **1996**, *32*, 3–14. [[CrossRef](#)]
73. Timmermann, V.; Dejgaard, L.A.; Haugaa, K.H.; Edwards, A.G.; Sundnes, J.; McCulloch, A.D.; Wall, S.T. An integrative appraisal of mechano-electric feedback mechanisms in the heart. *Prog. Biophys. Mol. Biol.* **2017**, *130*, 404–417. [[CrossRef](#)] [[PubMed](#)]
74. Takahashi, K.; Kakimoto, Y.; Toda, K.; Naruse, K. Mechanobiology in cardiac physiology and diseases. *J. Cell Mol. Med.* **2013**, *17*, 225–232. [[CrossRef](#)] [[PubMed](#)]
75. Ugolini, G.S.; Rasponi, M.; Pavesi, A.; Santoro, R.; Kamm, R.; Fiore, G.B.; Pesce, M.; Soncini, M. On-chip assessment of human primary cardiac fibroblasts proliferative responses to uniaxial cyclic mechanical strain. *Biotechnol. Bioeng.* **2016**, *113*, 859–869. [[CrossRef](#)]

76. Storch, U.; Mederos y Schnitzler, M.; Gudermann, T. G protein-mediated stretch reception. *Am. J. Physiol. Heart Circ. Physiol.* **2012**, *302*, H1241–H1249. [[CrossRef](#)]
77. Reed, A.; Kohl, P.; Peyronnet, R. Molecular candidates for cardiac stretch-activated ion channels. *Glob. Cardiol. Sci. Pract.* **2014**, *2014*, 9–25. [[CrossRef](#)] [[PubMed](#)]
78. Berridge, M.J.; Bootman, M.D.; Roderick, H.L. Calcium signalling: Dynamics, homeostasis and remodelling. *Nat. Reviews Mol. Cell Biol.* **2003**, *4*, 517–529. [[CrossRef](#)]
79. Layland, J.; Solaro, R.J.; Shah, A.M. Regulation of cardiac contractile function by troponin I phosphorylation. *Cardiovasc. Res.* **2005**, *66*, 12–21. [[CrossRef](#)]
80. Linke, W.A.; Kruger, M. The giant protein titin as an integrator of myocyte signaling pathways. *Physiology* **2010**, *25*, 186–198. [[CrossRef](#)]
81. Puchner, E.M.; Gaub, H.E. Exploring the conformation-regulated function of titin kinase by mechanical pump and probe experiments with single molecules. *Angew. Chem. Int. Ed. Engl.* **2010**, *49*, 1147–1150. [[CrossRef](#)]
82. Ahmed, S.H.; Lindsey, M.L. Titin phosphorylation: Myocardial passive stiffness regulated by the intracellular giant. *Circ. Res.* **2009**, *105*, 611–613. [[CrossRef](#)] [[PubMed](#)]
83. Sheikh, F.; Ross, R.S.; Chen, J. Cell-cell connection to cardiac disease. *Trends Cardiovasc. Med.* **2009**, *19*, 182–190. [[CrossRef](#)]
84. Lyon, R.C.; Zanella, F.; Omens, J.H.; Sheikh, F. Mechanotransduction in cardiac hypertrophy and failure. *Circ. Res.* **2015**, *116*, 1462–1476. [[CrossRef](#)] [[PubMed](#)]
85. Yoshida, M.; Sho, E.; Nanjo, H.; Takahashi, M.; Kobayashi, M.; Kawamura, K.; Honma, M.; Komatsu, M.; Sugita, A.; Yamauchi, M.; et al. Weaving hypothesis of cardiomyocyte sarcomeres: Discovery of periodic broadening and narrowing of intercalated disk during volume-load change. *Am. J. Pathol.* **2010**, *176*, 660–678. [[CrossRef](#)]
86. Chopra, A.; Tabdanov, E.; Patel, H.; Janmey, P.A.; Kresh, J.Y. Cardiac myocyte remodeling mediated by N-cadherin-dependent mechanosensing. *Am. J. Physiol. Heart Circ. Physiol.* **2011**, *300*. [[CrossRef](#)]
87. Kostetskii, I.; Li, J.; Xiong, Y.; Zhou, R.; Ferrari, V.A.; Patel, V.V.; Molkentin, J.D.; Radice, G.L. Induced deletion of the N-cadherin gene in the heart leads to dissolution of the intercalated disc structure. *Circ. Res.* **2005**, *96*, 346–354. [[CrossRef](#)]
88. Krusche, C.A.; Holthofer, B.; Hofe, V.; van de Sandt, A.M.; Eshkind, L.; Bockamp, E.; Merx, M.W.; Kant, S.; Windoffer, R.; Leube, R.E. Desmoglein 2 mutant mice develop cardiac fibrosis and dilation. *Basic Res. Cardiol.* **2011**, *106*, 617–633. [[CrossRef](#)]
89. Kant, S.; Krull, P.; Eisner, S.; Leube, R.E.; Krusche, C.A. Histological and ultrastructural abnormalities in murine desmoglein 2-mutant hearts. *Cell Tissue Res.* **2012**, *348*, 249–259. [[CrossRef](#)] [[PubMed](#)]
90. Kant, S.; Holthofer, B.; Magin, T.M.; Krusche, C.A.; Leube, R.E. Desmoglein 2-dependent arrhythmogenic cardiomyopathy is caused by a loss of adhesive function. *Circ. Cardiovasc. Genet.* **2015**, *8*, 553–563. [[CrossRef](#)]
91. Verheule, S.; Kaese, S. Connexin diversity in the heart: Insights from transgenic mouse models. *Front. Pharmacol.* **2013**, *4*, 81. [[CrossRef](#)]
92. Tiscornia, G.C.; Moretta, R.; Argenziano, M.A.; Amorena, C.E.; Garcia Gras, E.A. Inhibition of connexin 43 in cardiac muscle during intense physical exercise. *Scand. J. Med. Sci. Sports* **2014**, *24*, 336–344. [[CrossRef](#)]
93. Poelzing, S.; Rosenbaum, D.S. Altered connexin43 expression produces arrhythmia substrate in heart failure. *Am. J. Physiol. Heart Circ. Physiol.* **2004**, *287*, H1762–H1770. [[CrossRef](#)]
94. Manso, A.M.; Li, R.; Monkley, S.J.; Cruz, N.M.; Ong, S.; Lao, D.H.; Koshman, Y.E.; Gu, Y.; Peterson, K.L.; Chen, J.; et al. Talin1 has unique expression versus talin 2 in the heart and modifies the hypertrophic response to pressure overload. *J. Biol. Chem.* **2013**, *288*, 4252–4264. [[CrossRef](#)]
95. Imanaka-Yoshida, K.; Enomoto-Iwamoto, M.; Yoshida, T.; Sakakura, T. Vinculin, Talin, Integrin alpha6beta1 and laminin can serve as components of attachment complex mediating contraction force transmission from cardiomyocytes to extracellular matrix. *Cell Motil. Cytoskeleton* **1999**, *42*, 1–11. [[CrossRef](#)]
96. Santoro, R.; Perrucci, G.L.; Gowran, A.; Pompilio, G. Unchain my heart: Integrins at the basis of iPSC cardiomyocyte differentiation. *Stem Cells Int.* **2019**, *2019*, 8203950. [[CrossRef](#)] [[PubMed](#)]
97. Perrucci, G.L.; Barbagallo, V.A.; Corliano, M.; Tosi, D.; Santoro, R.; Nigro, P.; Poggio, P.; Bulfamante, G.; Lombardi, F.; Pompilio, G. Integrin alphanubeta5 in vitro inhibition limits pro-fibrotic response in cardiac fibroblasts of spontaneously hypertensive rats. *J. Transl. Med.* **2018**, *16*, 352. [[CrossRef](#)] [[PubMed](#)]
98. Buyandelger, B.; Mansfield, C.; Knoll, R. Mechano-signaling in heart failure. *Pflugers Arch. Eur. J. Physiol.* **2014**, *466*, 1093–1099. [[CrossRef](#)]
99. Shave, R.; Ross, P.; Low, D.; George, K.; Gaze, D. Cardiac troponin I is released following high-intensity short-duration exercise in healthy humans. *Int. J. Cardiol.* **2010**, *145*, 337–339. [[CrossRef](#)]
100. Fortescue, E.B.; Shin, A.Y.; Greenes, D.S.; Mannix, R.C.; Agarwal, S.; Feldman, B.J.; Shah, M.I.; Rifai, N.; Landzberg, M.J.; Newburger, J.W.; et al. Cardiac troponin increases among runners in the Boston Marathon. *Ann. Emerg. Med.* **2007**, *49*, 137–143. [[CrossRef](#)]
101. Eijsvogels, T.; George, K.; Shave, R.; Gaze, D.; Levine, B.D.; Hopman, M.T.; Thijssen, D.H. Effect of prolonged walking on cardiac troponin levels. *Am. J. Cardiol.* **2010**, *105*, 267–272. [[CrossRef](#)]
102. Jassal, D.S.; Moffat, D.; Krahn, J.; Ahmadie, R.; Fang, T.; Eschun, G.; Sharma, S. Cardiac injury markers in non-elite marathon runners. *Int. J. Sports Med.* **2009**, *30*, 75–79. [[CrossRef](#)] [[PubMed](#)]

103. Mingels, A.; Jacobs, L.; Michielsen, E.; Swaanenburg, J.; Wodzig, W.; van Dieijen-Visser, M. Reference population and marathon runner sera assessed by highly sensitive cardiac troponin T and commercial cardiac troponin T and I assays. *Clin. Chem.* **2009**, *55*, 101–108. [[CrossRef](#)] [[PubMed](#)]
104. Serrano-Ostariz, E.; Terreros-Blanco, J.L.; Legaz-Arrese, A.; George, K.; Shave, R.; Bocos-Terraz, P.; Izquierdo-Alvarez, S.; Bancalero, J.L.; Echavarri, J.M.; Quilez, J.; et al. The impact of exercise duration and intensity on the release of cardiac biomarkers. *Scand. J. Med. Sci. Sports* **2011**, *21*, 244–249. [[CrossRef](#)]
105. Shave, R.; George, K.P.; Atkinson, G.; Hart, E.; Middleton, N.; Whyte, G.; Gaze, D.; Collinson, P.O. Exercise-induced cardiac troponin T release: A meta-analysis. *Med. Sci. Sports Exerc.* **2007**, *39*, 2099–2106. [[CrossRef](#)] [[PubMed](#)]
106. Hubble, K.M.; Fatovich, D.M.; Grasko, J.M.; Vasikaran, S.D. Cardiac troponin increases among marathon runners in the Perth Marathon: The Troponin in Marathons (TRIM) study. *Med. J. Aust.* **2009**, *190*, 91–93. [[CrossRef](#)] [[PubMed](#)]
107. Sharma, S.; Papadakis, M.; Whyte, G. Chronic ultra-endurance exercise: Implications in arrhythmogenic substrates in previously normal hearts. *Heart* **2010**, *96*, 1255–1256. [[CrossRef](#)]
108. White, H.D. Pathobiology of troponin elevations: Do elevations occur with myocardial ischemia as well as necrosis? *J. Am. Coll. Cardiol.* **2011**, *57*, 2406–2408. [[CrossRef](#)]
109. Scherr, J.; Braun, S.; Schuster, T.; Hartmann, C.; Moehlenkamp, S.; Wolfarth, B.; Pressler, A.; Halle, M. 72-h kinetics of high-sensitive troponin T and inflammatory markers after marathon. *Med. Sci. Sports Exerc.* **2011**, *43*, 1819–1827. [[CrossRef](#)]
110. Tian, Y.; Nie, J.; Huang, C.; George, K.P. The kinetics of highly sensitive cardiac troponin T release after prolonged treadmill exercise in adolescent and adult athletes. *J. Appl. Physiol.* **2012**, *113*, 418–425. [[CrossRef](#)]
111. Liang, F.; Gardner, D.G. Mechanical strain activates BNP gene transcription through a p38/NF-kappaB-dependent mechanism. *J. Clin. Investig.* **1999**, *104*, 1603–1612. [[CrossRef](#)]
112. Herrmann, M.; Scharhag, J.; Miclea, M.; Urhausen, A.; Herrmann, W.; Kindermann, W. Post-race kinetics of cardiac troponin T and I and N-terminal pro-brain natriuretic peptide in marathon runners. *Clin. Chem.* **2003**, *49*, 831–834. [[CrossRef](#)]
113. Neumayr, G.; Pfister, R.; Mitterbauer, G.; Eibl, G.; Hoertnagl, H. Effect of competitive marathon cycling on plasma N-terminal pro-brain natriuretic peptide and cardiac troponin T in healthy recreational cyclists. *Am. J. Cardiol.* **2005**, *96*, 732–735. [[CrossRef](#)]
114. Ohba, H.; Takada, H.; Musha, H.; Nagashima, J.; Mori, N.; Awaya, T.; Omiya, K.; Murayama, M. Effects of prolonged strenuous exercise on plasma levels of atrial natriuretic peptide and brain natriuretic peptide in healthy men. *Am. Heart J.* **2001**, *141*, 751–758. [[CrossRef](#)]
115. Scharhag, J.; Herrmann, M.; Urhausen, A.; Haschke, M.; Herrmann, W.; Kindermann, W. Independent elevations of N-terminal pro-brain natriuretic peptide and cardiac troponins in endurance athletes after prolonged strenuous exercise. *Am. Heart J.* **2005**, *150*, 1128–1134. [[CrossRef](#)]
116. Siegel, A.J.; Lewandrowski, E.L.; Chun, K.Y.; Sholar, M.B.; Fischman, A.J.; Lewandrowski, K.B. Changes in cardiac markers including B-natriuretic peptide in runners after the Boston marathon. *Am. J. Cardiol.* **2001**, *88*, 920–923. [[CrossRef](#)]
117. Kanter, M.M.; Lesmes, G.R.; Kaminsky, L.A.; La Ham-Saeger, J.; Nequin, N.D. Serum creatine kinase and lactate dehydrogenase changes following an eighty kilometer race. Relationship to lipid peroxidation. *Eur. J. Appl. Physiol. Occup. Physiol.* **1988**, *57*, 60–63. [[CrossRef](#)] [[PubMed](#)]
118. Konig, D.; Schumacher, Y.O.; Heinrich, L.; Schmid, A.; Berg, A.; Dickhuth, H.H. Myocardial stress after competitive exercise in professional road cyclists. *Med. Sci. Sports Exerc.* **2003**, *35*, 1679–1683. [[CrossRef](#)]
119. Scharhag, J.; Urhausen, A.; Schneider, G.; Herrmann, M.; Schumacher, K.; Haschke, M.; Krieg, A.; Meyer, T.; Herrmann, W.; Kindermann, W. Reproducibility and clinical significance of exercise-induced increases in cardiac troponins and N-terminal pro brain natriuretic peptide in endurance athletes. *Eur. J. Cardiovasc. Prev. Rehabil.* **2006**, *13*, 388–397. [[CrossRef](#)] [[PubMed](#)]
120. Brancaccio, P.; Maffulli, N.; Limongelli, F.M. Creatine kinase monitoring in sport medicine. *Br. Med. Bull.* **2007**, *81–82*, 209–230. [[CrossRef](#)] [[PubMed](#)]
121. Aengevaeren, V.L.; RRJ, V.A.N.K.; Hopman, M.T.E.; Van Royen, N.; Snider, J.V.; Januzzi, J.L.; George, K.P.; Eijsvogels, T.M.H. Exercise-induced changes in soluble ST2 concentrations in marathon runners. *Med. Sci. Sports Exerc.* **2019**, *51*, 405–410. [[CrossRef](#)] [[PubMed](#)]
122. Hattasch, R.; Spethmann, S.; de Boer, R.A.; Ruifrok, W.P.; Schattke, S.; Wagner, M.; Schroeckh, S.; Durmus, T.; Schimke, I.; Sanad, W.; et al. Galectin-3 increase in endurance athletes. *Eur. J. Prev. Cardiol.* **2014**, *21*, 1192–1199. [[CrossRef](#)] [[PubMed](#)]
123. Nieman, D.C.; Shanely, R.A.; Luo, B.; Meaney, M.P.; Dew, D.A.; Pappan, K.L. Metabolomics approach to assessing plasma 13- and 9-hydroxy-octadecadienoic acid and linoleic acid metabolite responses to 75-km cycling. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2014**, *307*, R68–R74. [[CrossRef](#)] [[PubMed](#)]
124. Spiteller, G. Linoleic acid peroxidation—the dominant lipid peroxidation process in low density lipoprotein—and its relationship to chronic diseases. *Chem. Phys. Lipids.* **1998**, *95*, 105–162. [[CrossRef](#)]
125. Lovlin, R.; Cottle, W.; Pyke, I.; Kavanagh, M.; Belcastro, A.N. Are indices of free radical damage related to exercise intensity. *Eur. J. Appl. Physiol. Occup. Physiol.* **1987**, *56*, 313–316. [[CrossRef](#)]
126. Walsh, R.C.; Koukoulas, I.; Garnham, A.; Moseley, P.L.; Hargreaves, M.; Febbraio, M.A. Exercise increases serum Hsp72 in humans. *Cell Stress Chaperones* **2001**, *6*, 386–393. [[CrossRef](#)]
127. Febbraio, M.A.; Ott, P.; Nielsen, H.B.; Steensberg, A.; Keller, C.; Krstrup, P.; Secher, N.H.; Pedersen, B.K. Exercise induces hepatosplanchnic release of heat shock protein 72 in humans. *J. Physiol.* **2002**, *544*, 957–962. [[CrossRef](#)] [[PubMed](#)]

128. Banfi, G.; Dolci, A.; Verna, R.; Corsi, M.M. Exercise raises serum heat-shock protein 70 (Hsp70) levels. *Clin. Chem. Lab. Med.* **2004**, *42*, 1445–1446. [[CrossRef](#)] [[PubMed](#)]
129. Wei, Y.J.; Huang, Y.X.; Shen, Y.; Cui, C.J.; Zhang, X.L.; Zhang, H.; Hu, S.S. Proteomic analysis reveals significant elevation of heat shock protein 70 in patients with chronic heart failure due to arrhythmogenic right ventricular cardiomyopathy. *Mol. Cell. Biochem.* **2009**, *332*, 103–111. [[CrossRef](#)]
130. Songia, P.; Chiesa, M.; Valerio, V.; Moschetta, D.; Myasoedova, V.A.; D'Alessandra, Y.; Poggio, P. Direct screening of plasma circulating microRNAs. *RNA Biology* **2018**, *15*, 1268–1272. [[CrossRef](#)]
131. D'Alessandra, Y.; Chiesa, M.; Carena, M.C.; Beltrami, A.P.; Rizzo, P.; Buzzetti, M.; Ricci, V.; Ferrari, R.; Fucili, A.; Livi, U.; et al. Differential role of circulating microRNAs to track progression and pre-symptomatic stage of chronic heart failure: A pilot study. *Biomedicine* **2020**, *8*, 597. [[CrossRef](#)]
132. Stadiotti, I.; Pompilio, G.; Maione, A.S.; Pilato, C.A.; D'Alessandra, Y.; Sommariva, E. Arrhythmogenic cardiomyopathy: What blood can reveal. *Heart Rhythm* **2018**. [[CrossRef](#)]
133. Oliveira, N.R.d.; Oliveira, W.S.d.; Porto, A.A.; Mastrocola, F.; Novaes, A.E.; Mendonça, R.M.; Sousa, J.C.V.d. Cardiac arrest and exercise-induced polymorphic ventricular tachycardia: An elusive diagnosis. *Int. J. Cardiovasc. Sci.* **2021**. [[CrossRef](#)]
134. Thompson, A.J.; Cannon, B.C.; Wackel, P.L.; Horner, J.M.; Ackerman, M.J.; O'Leary, P.W.; Eidem, B.W.; Johnson, J.N. Electrocardiographic abnormalities in elite high school athletes: Comparison to adolescent hypertrophic cardiomyopathy. *Br. J. Sports Med.* **2016**, *50*, 105–110. [[CrossRef](#)] [[PubMed](#)]
135. Malhotra, A.; Sharma, S. Hypertrophic cardiomyopathy in athletes. *Eur. Cardiol.* **2017**, *12*, 80–82. [[CrossRef](#)]
136. Stadiotti, I.; Catto, V.; Casella, M.; Tondo, C.; Pompilio, G.; Sommariva, E. Arrhythmogenic cardiomyopathy: The guilty party in adipogenesis. *J. Cardiovasc. Transl. Res.* **2017**, *10*, 446–454. [[CrossRef](#)] [[PubMed](#)]
137. Xu, Z.; Zhu, W.; Wang, C.; Huang, L.; Zhou, Q.; Hu, J.; Cheng, X.; Hong, K. Genotype-phenotype relationship in patients with arrhythmogenic right ventricular cardiomyopathy caused by desmosomal gene mutations: A systematic review and meta-analysis. *Sci. Rep.* **2017**, *7*, 41387. [[CrossRef](#)]
138. Corrado, D.; Basso, C.; Schiavon, M.; Thiene, G. Does sports activity enhance the risk of sudden cardiac death? *J. Cardiovasc. Med.* **2006**, *7*, 228–233. [[CrossRef](#)]
139. La Gerche, A.; Robberecht, C.; Kuiperi, C.; Nuyens, D.; Willems, R.; de Ravel, T.; Matthijs, G.; Heidbuchel, H. Lower than expected desmosomal gene mutation prevalence in endurance athletes with complex ventricular arrhythmias of right ventricular origin. *Heart* **2010**, *96*, 1268–1274. [[CrossRef](#)]
140. La Gerche, A.; Burns, A.T.; Mooney, D.J.; Inder, W.J.; Taylor, A.J.; Bogaert, J.; Macisaac, A.I.; Heidbuchel, H.; Prior, D.L. Exercise-induced right ventricular dysfunction and structural remodelling in endurance athletes. *Eur. Heart J.* **2012**, *33*, 998–1006. [[CrossRef](#)]
141. Heidbuchel, H.; Hoogsteen, J.; Fagard, R.; Vanhees, L.; Ector, H.; Willems, R.; Van Lierde, J. High prevalence of right ventricular involvement in endurance athletes with ventricular arrhythmias. Role of an electrophysiologic study in risk stratification. *Eur. Heart J.* **2003**, *24*, 1473–1480. [[CrossRef](#)]
142. Heidbuchel, H.; La Gerche, A. The right heart in athletes. Evidence for exercise-induced arrhythmogenic right ventricular cardiomyopathy. *Herzschrittmacherther Elektrophysiol.* **2012**, *23*, 82–86. [[CrossRef](#)] [[PubMed](#)]
143. Leischik, R.; Dworak, B.; Strauss, M.; Horlitz, M.; Pareja-Galeano, H.; de la Guia-Galipienso, F.; Lippi, G.; Lavie, C.J.; Perez, M.V.; Sanchis-Gomar, F. Exercise-induced right ventricular injury or arrhythmogenic cardiomyopathy (ACM): The bright side and the dark side of the moon. *Prog. Cardiovasc. Dis.* **2020**, *63*, 671–681. [[CrossRef](#)]
144. Benito, B.; Gay-Jordi, G.; Serrano-Mollar, A.; Guasch, E.; Shi, Y.; Tardif, J.C.; Brugada, J.; Nattel, S.; Mont, L. Cardiac arrhythmogenic remodeling in a rat model of long-term intensive exercise training. *Circulation* **2011**, *123*, 13–22. [[CrossRef](#)]
145. Kindermann, W.; Corrado, D.; Scharhag, J. The right heart in athletes. Do we really have sufficient evidence for exercise-induced arrhythmogenic right ventricular cardiomyopathy? *Herzschrittmacherther Elektrophysiol.* **2012**, *23*, 144–147. [[CrossRef](#)]
146. Prior, D. Differentiating athlete's heart from cardiomyopathies—the right side. *Heart Lung Circ.* **2018**, *27*, 1063–1071. [[CrossRef](#)] [[PubMed](#)]
147. Gasperetti, A.; James, C.A.; Cerrone, M.; Delmar, M.; Calkins, H.; Duru, F. Arrhythmogenic right ventricular cardiomyopathy and sports activity: From molecular pathways in diseased hearts to new insights into the athletic heart mimicry. *Eur. Heart J.* **2021**, *42*, 1231–1243. [[CrossRef](#)] [[PubMed](#)]
148. Maron, B.J.; Maron, B.A. Revisiting athlete's heart versus pathologic hypertrophy: ARVC and the right ventricle. *JACC Cardiovasc. Imaging* **2017**, *10*, 394–397. [[CrossRef](#)]
149. Zaidi, A.; Sheikh, N.; Jongman, J.K.; Gati, S.; Panoulas, V.F.; Carr-White, G.; Papadakis, M.; Sharma, R.; Behr, E.R.; Sharma, S. Clinical differentiation between physiological remodeling and arrhythmogenic right ventricular cardiomyopathy in athletes with marked electrocardiographic repolarization anomalies. *J. Am. Coll. Cardiol.* **2015**, *65*, 2702–2711. [[CrossRef](#)] [[PubMed](#)]
150. Bauce, B.; Frigo, G.; Benini, G.; Michieli, P.; Basso, C.; Folino, A.F.; Rigato, I.; Mazzotti, E.; D'Aliento, L.; Thiene, G.; et al. Differences and similarities between arrhythmogenic right ventricular cardiomyopathy and athlete's heart adaptations. *Br. J. Sports Med.* **2010**, *44*, 148–154. [[CrossRef](#)] [[PubMed](#)]
151. Finocchiaro, G.; Papadakis, M.; Robertus, J.L.; Dhutia, H.; Steriotis, A.K.; Tome, M.; Mellor, G.; Merghani, A.; Malhotra, A.; Behr, E.; et al. Etiology of sudden death in sports: Insights from a United Kingdom regional registry. *J. Am. Coll. Cardiol.* **2016**, *67*, 2108–2115. [[CrossRef](#)]

152. Zaidi, A.; Ghani, S.; Sharma, R.; Oxborough, D.; Panoulas, V.F.; Sheikh, N.; Gati, S.; Papadakis, M.; Sharma, S. Physiological right ventricular adaptation in elite athletes of African and Afro-Caribbean origin. *Circulation* **2013**, *127*, 1783–1792. [[CrossRef](#)] [[PubMed](#)]
153. Chatterjee, D.; Fatah, M.; Akdis, D.; Spears, D.A.; Koopmann, T.T.; Mittal, K.; Rafiq, M.A.; Cattanach, B.M.; Zhao, Q.; Healey, J.S.; et al. An autoantibody identifies arrhythmogenic right ventricular cardiomyopathy and participates in its pathogenesis. *Eur. Heart J.* **2018**, *39*, 3932–3944. [[CrossRef](#)]
154. Maione, A.S.; Pilato, C.A.; Casella, M.; Gasperetti, A.; Stadiotti, I.; Pompilio, G.; Sommariva, E. Fibrosis in Arrhythmogenic Cardiomyopathy: The phantom thread in the fibro-adipose tissue. *Front. Physiol.* **2020**, *11*, 279. [[CrossRef](#)]
155. Zuhl, M.; Schneider, S.; Lanphere, K.; Conn, C.; Dokladny, K.; Moseley, P. Exercise regulation of intestinal tight junction proteins. *Br. J. Sports Med.* **2014**, *48*, 980–986. [[CrossRef](#)]
156. Li, J. The Role of Autoantibodies in Arrhythmogenesis. *Curr. Cardiol. Rep.* **2020**, *23*, 3. [[CrossRef](#)] [[PubMed](#)]
157. Kasperkiewicz, M.; Ellebrecht, C.T.; Takahashi, H.; Yamagami, J.; Zillikens, D.; Payne, A.S.; Amagai, M. Pemphigus. *Nat. Rev. Dis. Primers* **2017**, *3*, 17026. [[CrossRef](#)]
158. Jennings, J.M.; Tucker, D.K.; Kottke, M.D.; Saito, M.; Delva, E.; Hanakawa, Y.; Amagai, M.; Kowalczyk, A.P. Desmosome disassembly in response to pemphigus vulgaris IgG occurs in distinct phases and can be reversed by expression of exogenous Dsg3. *J. Investig. Dermatol.* **2011**, *131*, 706–718. [[CrossRef](#)] [[PubMed](#)]
159. Sumigray, K.; Zhou, K.; Lechler, T. Cell-cell adhesions and cell contractility are upregulated upon desmosome disruption. *PLoS ONE* **2014**, *9*, e101824. [[CrossRef](#)]
160. Lazzerini, P.E.; Capecchi, P.L.; El-Sherif, N.; Laghi-Pasini, F.; Boutjdir, M. Emerging arrhythmic risk of autoimmune and inflammatory cardiac channelopathies. *J. Am. Heart Assoc.* **2018**, *7*, e010595. [[CrossRef](#)]